

MITSUBISHI CHEMICAL CORPORATION

Mitsubishi Chemical Corporation
Separation Materials Department
1-11-2, Osaki, Shinagawa-ku, Tokyo, 141-0032, Japan
Phone: +81-3-6748-7146 Fax: +81-3-5487-6810

please visit
<http://www.diaion.com>

MCI GELTM XtalSpeedTM

MCI GELTM XtalSpeedTM

TECHNICAL INFORMATION 2017-2019

MITSUBISHI CHEMICAL CORPORATION



MCI GELTM XtalSpeedTM

Excellent performance

spherical and sharp particle size distribution

Persistence and highest quality

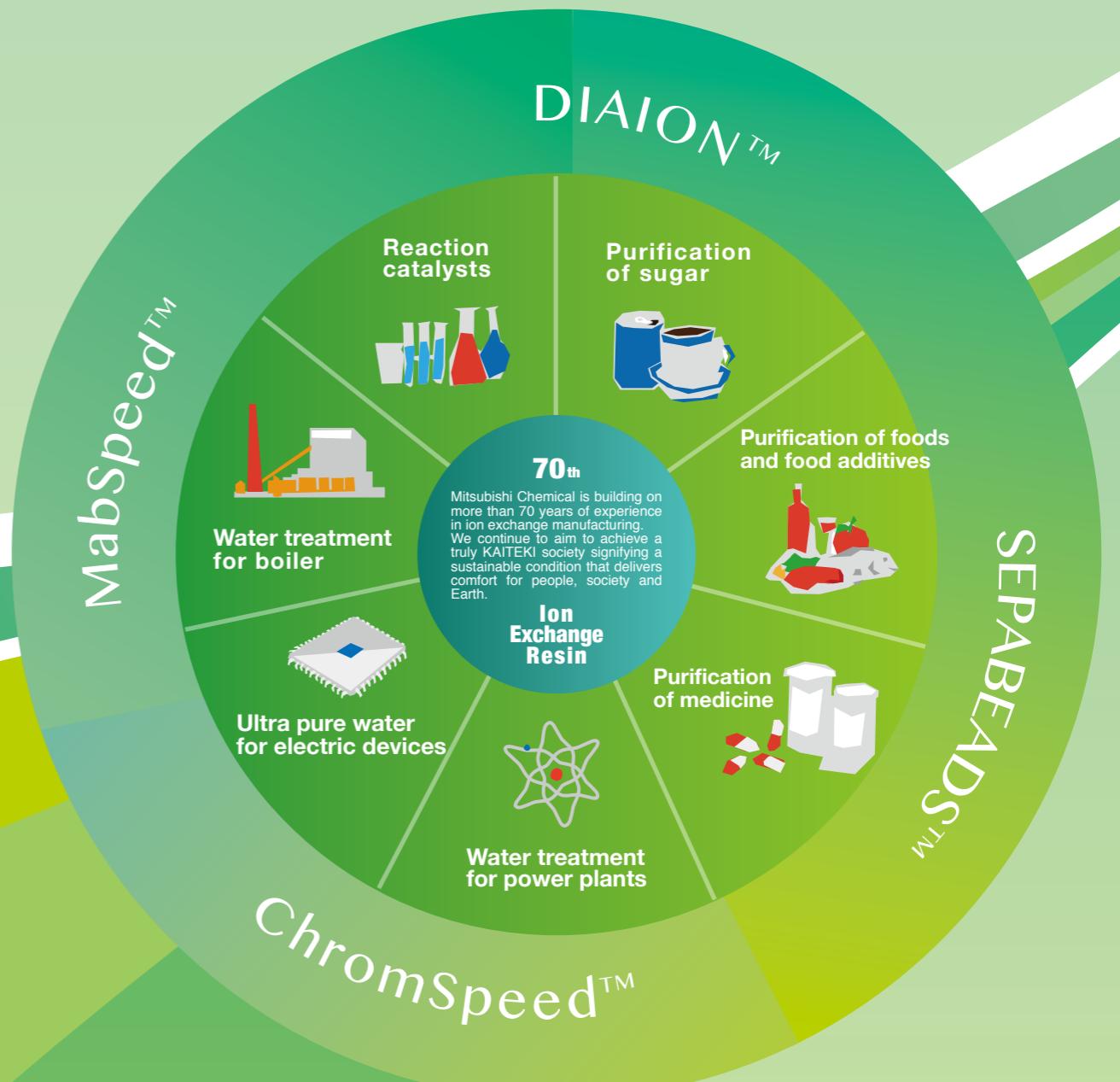
offers packing materials and packed columns,
under strict quality control

Wide range of product line

MCI GELTM has been designed based on technology of
the world famous DiaionTM and SepabeadsTM,
specialized in polymeric packing materials including
from analytical to preparative use,
for ion exchange, reversed-phase mode

Abundant accumulation of technology
and experience

for more than 50 years, MCI GELTM has been used for
HPLC applications



CONTENTS

1 Column selection guide	3~4
Column selection guide	3
Product list by particle size and separation mode	4
USP List	4
2 Ion exchange columns and materials	5~22
Features	5
Column list and materials	6
[Applications] Sugars•Sugar alcohols•Organic acids《CK08,06 series》	7~12
Examples of peak retention time《CK08E series》	13
Oligosaccharides《CK04S•CK04SS•CK02A•CK02AS》	14~16
Amino acids《CK10U》	17~18
Sugars•Organic acids《CA08F》	19~20
Nucleic acids etc. •Sugars•Human urine《CDR10》	21~22
3 Ion chromatography columns and materials	23~27
Column list and materials	23~24
[Applications] Cations《SCK01》	24~25
Cations《CHK45》	25
Anions《SCA04》	26~27
4 Bioseparation columns and materials	28~32
Bioseparation columns	28
Ion exchange chromatography column《XtalSpeed™ series》	29~30
Column list and materials	29
[Applications] Rituxan《XtalSpeed™ series》	30
Aqueous size exclusion columns《CQP series》	31~32
Size exclusion chromatography columns	31
Column list and materials	31
[Applications] Calibration curves《CQP series》	32
Proteins•Water soluble polymers《CQP series》	32
5 Analytical and preparative chromatography columns and materials for pharmaceutical applications	33~53
Polymeric partition chromatography columns and materials MCI GEL™ CHP series	33
Separation mechanism of CHP series	33
Column list	34
Column durability	35
[Applications] Organic compounds•Insulin•Peptide《CHP column series》	36~48
Polymeric chromatography materials《CHP material series》	49~56
Chromatography material list	49
[Applications] Organic compounds《CHP material series》	50~53
6 Chiral separation columns	54~59
Separation mechanism of CRS series	54
[Applications] Optical isomers《CRS10W•CRS15W》	55~58
Separation conditions for various amino acids	59
7 SPE sorbent series	60
Solid phase extraction sorbents	60
Synthetic adsorbents and reversed-phase material list	60
Chelating type list	60
8 MCI GEL™ column list	61~62
9 MCI GEL™ material list	63~66
10 Compounds index	67~75



1 Column selection guide

Nature of sample	Separation mode	MCI GEL™ column	pH range	Applications	Pages
Water Soluble	M.W. >2,000	Size Exclusion	CQP10 CQP30	2 ~12 Proteins, Biopolymers Water soluble polymers	31~32
		Ion Exchange	XtalSpeed™	2 ~12 Proteins, Antibody	29~31
		Reversed-Phase	CMG20 CHPOD	2 ~12 Proteins, Peptides	33~53
			CHP20 CHP07	Full range Proteins, Peptides	33~53
			CK02A CK02AS	6 ~7 Oligosaccharides	14~16
		CK04S CK04SS	6 ~7 Oligosaccharides	14~16	
		CQP06	2 ~12 Peptides	31~32	
		CK10U	1 ~14 Amino acids	17~18	
		CA08F CDR10	1 ~13 Organic acids Saccharides	19~22	
		CDR10	1 ~13 Nucleotides	21~22	
M.W. <2,000	Ion Chromatography	SCA04	3 ~7 Anions	23~27	
		SCK01	1.5~12 Cations	23~25	
		CHK45			
		CMG20 CHPOD	2 ~12 Organic Compounds peptide	33~53	
		CHP20 CHP07	Full range Organic Compounds peptide	33~53	
Sample	Reversed-Phase	CHK40 CHK45	Full range Amino acids, Nucleotide	34,45~48	
		CK08EH	1 ~7 Organic acids	7,11~12	
		CK08E Series	1 ~7 Saccharides	7~13	
		CRS10W CRS15W	5 ~7 Optical isomers (α -amino acids, α -hydroxy carboxylic acids)	54~59	
		CHP20 CMG20 CHP07	Full range Organic Compounds	33~53	
	CMG20 CHPOD	2 ~12 Organic Compounds	33~53		
	CHK40 CHK45	Full range Organic Compounds	34,45,48		
	Normal Phase	CMG20 CHPOD	2 ~12 Organic Compounds	33~53	
	Mix mode	CHK40 CHK45	Full range Organic Compounds	34,45,48	
	Organic Solvent Soluble				

● Product list by particle size and separation mode

Separation mode	Particle size [μm]	Analytical				Preparative	
		5	10	30	50	150	
Ion exchange	CK XtalSpeed™ CDR10	CK CA CDR10	CK CA	CK CA	CK CA	CK CA	
							CQA_P CQK_P
Ion chromatography	CHK45 SCA			SCK			
Size exclusion				CQP			
Normal phase	CHP20/C04 CHP07/C04 CMG20/C04 CHPOD/C04	CHP20/P10 CHP50/P10	CHP20/P20 CHP50/P20	CHP20/P30 CHP50/P30	CHP20/P50	CHP20/P70	CHP20/P120
Mix mode	CHK40 CHK45			CMG20/P30 CHPOD/P30			CMG20/P150 CHP85/P120 CHP87/P120
Ligand exchange	CRS						

● USP LISTING OF MCI GEL™

USP Code	PACKING	MCI GEL™ Column	Page
L17	Strong cation-exchange resin consisting of sulfonated cross-linked styrene-divinylbenzene copolymer in the hydrogen form, 6 to 12 μm in diameter	CK08EH	7
L19	Strong cation-exchange resin consisting of sulfonated cross-linked styrene-divinylbenzene copolymer in the calcium form, 5 – 15 μm in diameter	CK08EC	7
L21	A rigid, spherical styrene-divinylbenzene copolymer, 3 to 30 μm in diameter	CHP20/C04 CHP20/C10	34
L25	Packing having the capacity to separate compounds with a molecular weight range from 100-5000 (as determined by polyethylene oxide), applied to neutral, anionic, and cationic water-soluble polymers. A polymethacrylate resin base, cross-linked with polyhydroxylated ether (surface contained some residual carboxyl functional groups) was found suitable	CQP06	31
L31	A hydroxide-selective, strong anion-exchange resin-quaternary amine bonded on latex particles attached to a core of 8.5-μm macroporous particles having a pore size of 2000 Å units and consisting of ethylvinylbenzene cross-linked with 55% divinylbenzene	SCA04	23
L32	A chiral ligand-exchange resin packing-L-proline copper complex covalently bonded to irregularly shaped silica particles, 5 to 10 μm in diameter	CRS10W CRS15W	54
L37	Packing having the capacity to separate proteins by molecular size over a range of 2,000 to 40,000 Da. It is a polymethacrylate gel	CQP30	31
L38	A methacrylate-based size-exclusion packing for water-soluble samples	CQP10 CQP30	31
L39	A hydrophilic polyhydroxymethacrylate gel of totally porous spherical resin	CMG20/C04 CMG20/C10	34
L58	Strong cation-exchange resin consisting of sulfonated cross-linked styrene-divinylbenzene copolymer in the sodium form, about 6 to 30 μm diameter	CK08S CK08E CK04S CK02A	6
L71	A rigid, spherical polymethacrylate, 4 to 6 μm in diameter	CMG20/C04	34
L73	A rigid, spherical polydivinylbenzene particle, 5 to 10 μ in diameter	CDR10	21

Ion exchange columns and materials

○ Cation exchange resins MCI GEL™ CK series

○ Anion exchange resins MCI GEL™ CA series

Mitsubishi Chemical Ion Exchange Resins

MCI GEL™ specializes in polymer based packing materials. Specifically, polystyrene polymer based ion exchange resins are derived from over 50 years of manufacturing experience of Diaion™ product line. MCI GEL™ ion exchange resins for HPLC have been developed with the same attention to performance and quality. For several decades, Mitsubishi Chemical has been providing MCI GEL™ ion exchange columns are offered in a variety of chemistries, particle sizes and counter ions to support a broad range of applications.

Features

1. Variety of products gel type, porous type, DVB%, particle size, particle size distribution analytical use, preparative use

2. Persistence of high quality, excellent separation performance

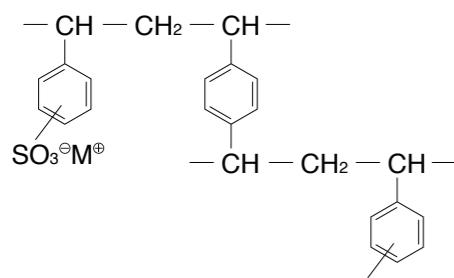
3. Accumulation of abundant knowledge and experience of applications

Ion exchange resins are generally used for analysis of amino acids, sugars, organic acids and amines, etc. MCI GEL™ custom pre-packed columns are specifically designed for each application using the most appropriate packing material among our product line and using the most suitable column dimensions.

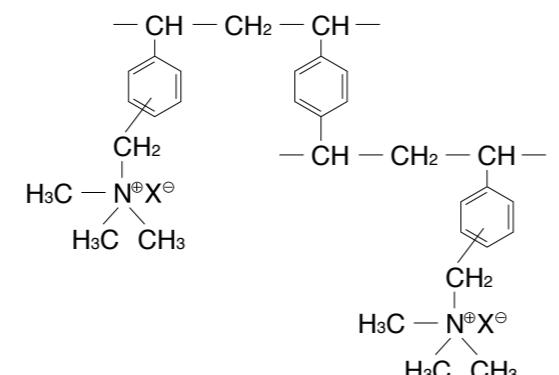
Typical application for each column is shown in this catalog. These data will suggest an appropriate column.

● Chemical structure of ion exchange resin

⟨Strongly acidic cation exchange resin⟩



⟨Strongly basic anion exchange resin⟩



● MCI GEL™ columns for HPLC

Product name	Column dimensions I.D×L [mm]	Description			USP	Typical usage				
		Cross linkage [%]	Counter ion	Particle size [μm]		Amino acid	Mono saccharide	Oligo- saccharide	Carboxylic acid	Amine
MCI GEL™ CK10U	6×120	10	Na ⁺	5		○				○
MCI GEL™ CK08S	8×500	8	Na ⁺	11	L58	○				
MCI GEL™ CK08E	8.0×300 7.8×300	8	Na ⁺	9	L58	○				
MCI GEL™ CK08EC	8.0×300 7.8×300	8	Ca ²⁺	9	L19	○				
MCI GEL™ CK08ES	8×300	8	Ag ⁺	9		○	○			
MCI GEL™ CK08EH	8.0×300 7.8×300	8	H ⁺	9	L17	○	○	○	○	○
MCI GEL™ CK06SC	8×500	6	Ca ²⁺	11		○	○			
MCI GEL™ CK04S	10×200	4	Na ⁺	11	L58	○				
MCI GEL™ CK04SS	10×200	4	Ag ⁺	11		○				
MCI GEL™ CK02A	20×250	2	Na ⁺	20	L58	○				
MCI GEL™ CK02AS	20×250	2	Ag ⁺	20		○				
MCI GEL™ CA08F	4.6×250	8	SO ₄ ²⁻	7		○		○		
MCI GEL™ CDR10	4.6×250	High porous	AcO ⁻	7		○		○		○

● Packing materials

Packing materials are available. Please look at P.64 and P.65.

● Description of a gel type ion exchange column

MCI GEL™ CK08EC

for HPLC use

Cation=K
Anion=A

DVB%

Counter ion
(no letter=Na⁺, C=Ca²⁺
(S=Ag⁺, H=H⁺)

Particle size (mode)
(A=20μm, S=11μm
E=9μm, F=7μm,
U=5μm)

● Note ; Pre-column and guard column

1. Please consider using a guard column concerning purity of injection sample. Guard columns, are listed in the end of this catalog, should be selected in accordance with a main column.

2. As for analysis of amino acids by MCI GEL™ CK10U, MCI GEL™ AFR2-PC is recommended as a pre-column. The AFR2-PC column is very effective to stabilize base line because it can trap ammonium ion in eluent. A peak caused of the ammonium ion may disturb base line stability.

2 MCI GEL™

CK08,06 series

Cation exchange columns
applications; sugars, carboxylic acids, (poly)alcohols, etc.



● Column list

MCI GEL™ column	Counter ion	Application areas	USP
MCI GEL™ CK08S MCI GEL™ CK08E	Na ⁺	General sugar separation columns	L58
MCI GEL™ CK08EC	Ca ²⁺	The most general sugar separation column Highly recommended for fructose and glucose This column conforms to US Pharmacopeia.	L19
MCI GEL™ CK08ES	Ag ⁺	Gel permeation chromatographic effect	
MCI GEL™ CK08EH	H ⁺	Organic acids with H ₃ PO ₄ eluent; sugars with distilled water eluent	L17
MCI GEL™ CK06SC	Ca ²⁺	Use for analysis of mono-saccharides and disaccharides.	

Application data of CK08EC

Fig. 2-1 Sugars

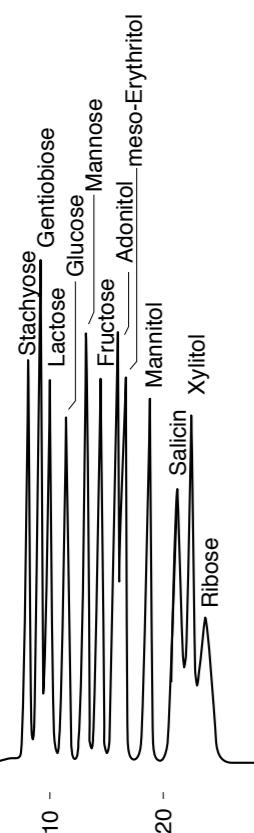
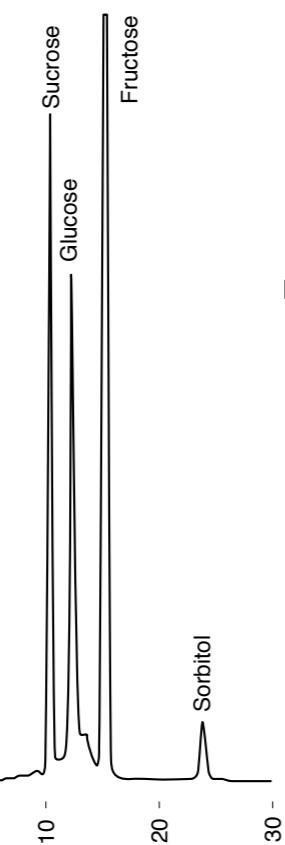
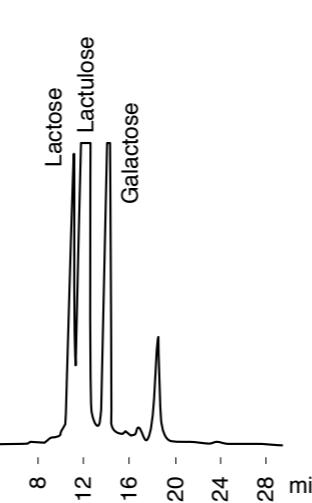


Fig. 2-2 Apple juice



Conditions
Column : MCI GEL™ CK08EC
8mm I.D.×300mm
Eluent : H₂O
Flow rate : 0.6 mL/min
Column temp. : 75°C
Detection : RI

Fig. 2-3 Lactulose syrup



Application data of CK08EC

Fig. 2-4 Sports drink A

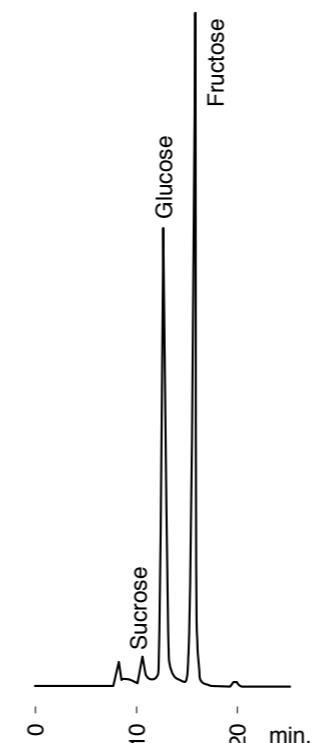


Fig. 2-5 Sports drink B

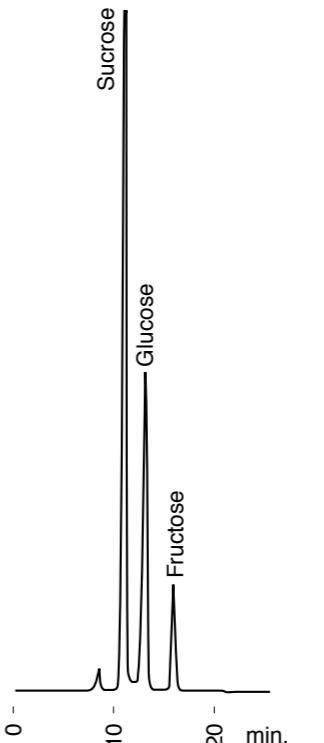


Fig. 2-6 Honey

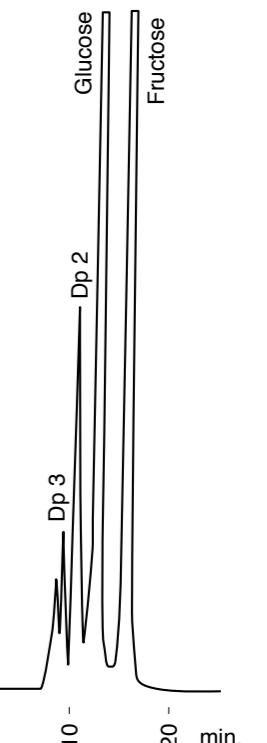


Fig. 2-7 Jam

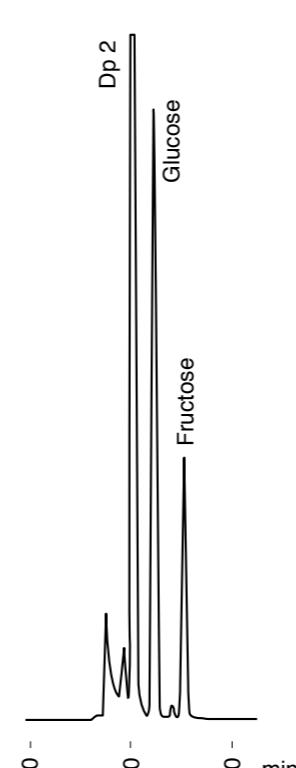
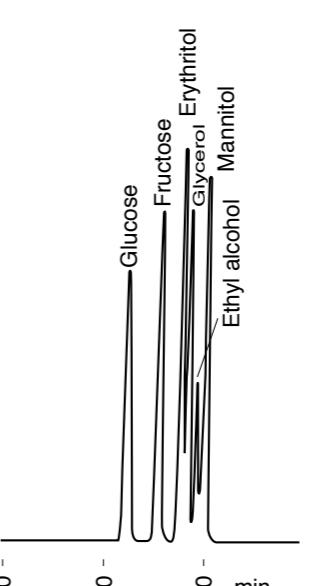


Fig. 2-8 Sugars/Alcohols



Conditions
Column : MCI GEL™ CK08EC
8mm I.D.×300mm
Eluent : H₂O
Flow rate : 0.6 mL/min
Column temp. : 75°C
Detection : RI

Application data of CK08EC

Fig. 2-9 Sugars/Alcoles (Comparison with competitor's column)

Conditions
Column : 7.8x 300 mmI.D. (MCI GEL™ CK08EC / Competitor's Column)
Eluent : Milli Q water
Flow rate : 0.6 mL/min
Temperature : 75 °C
Sample Conc : 40mmol/ml each
Injection : 20μL
Detection : RI

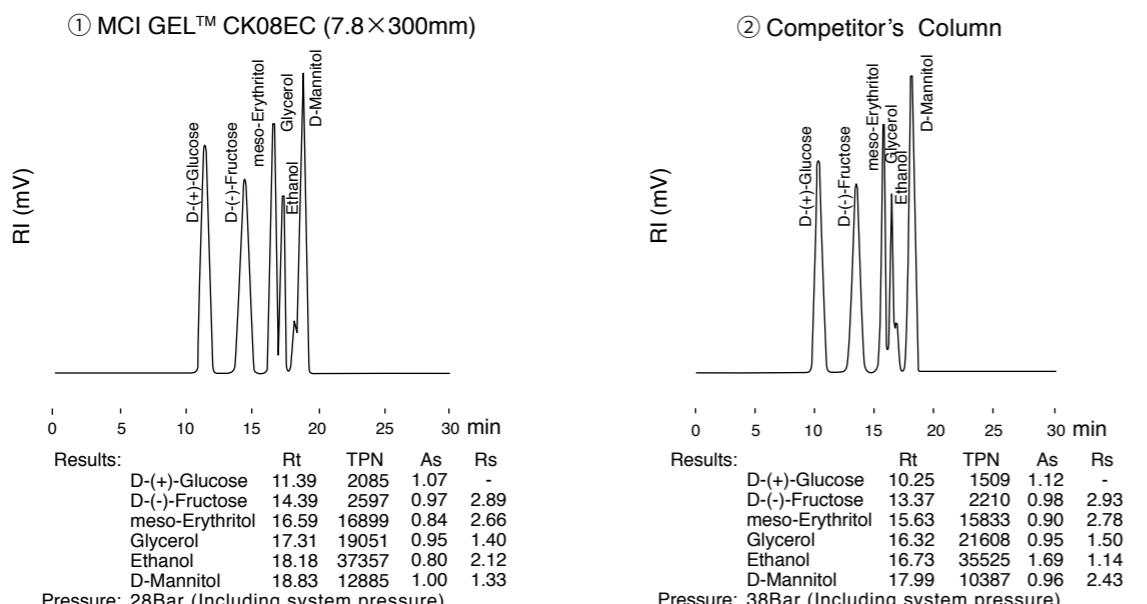
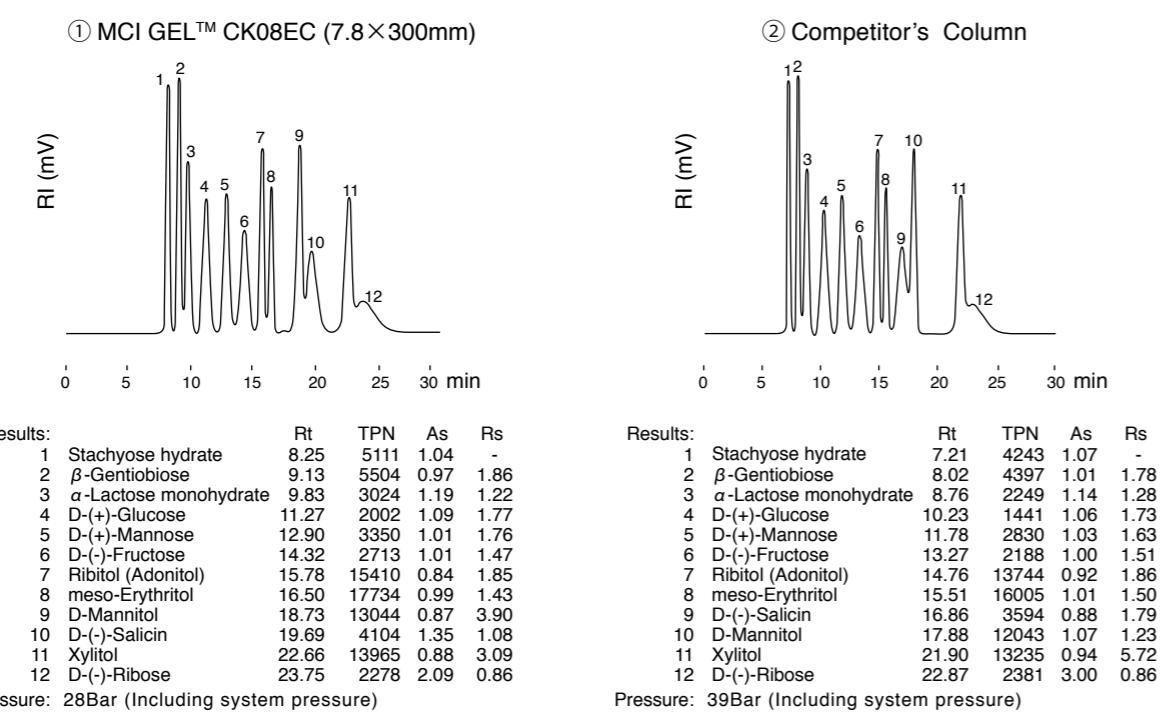


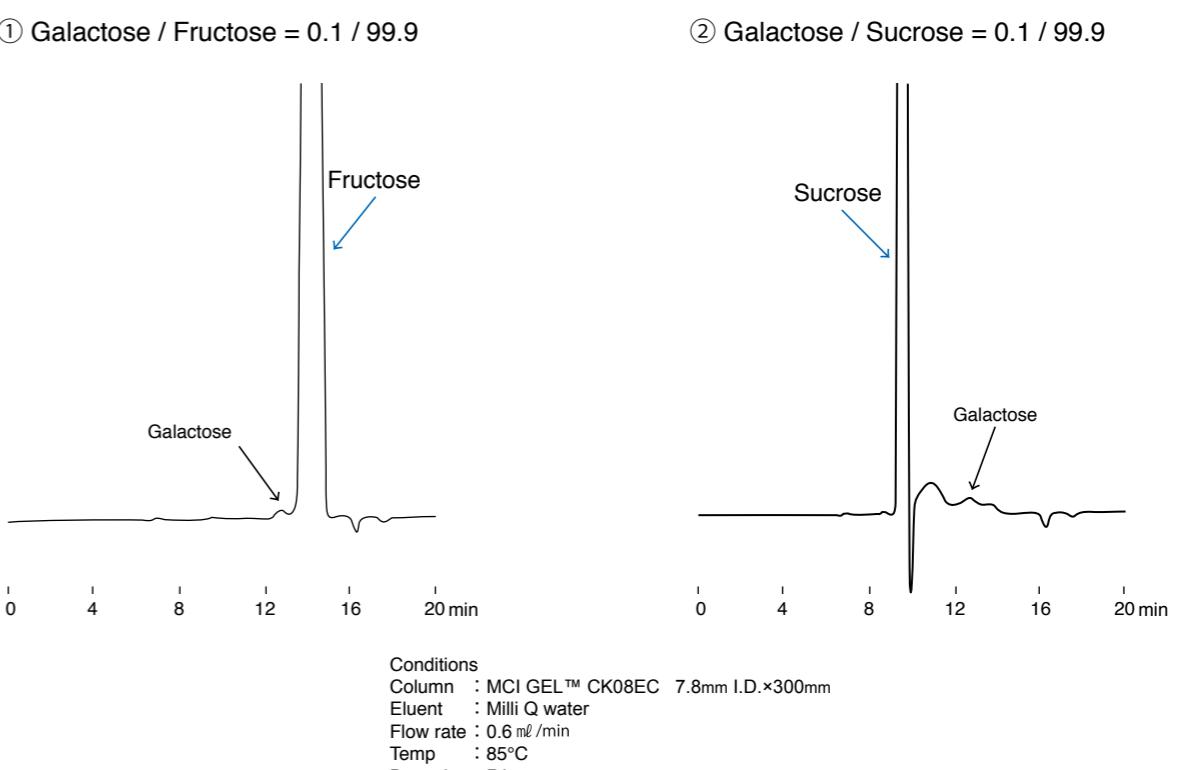
Fig. 2-10 Sugars (Comparison with competitor's column)

Conditions
Column : 7.8x 300 mmI.D. (MCI GEL™ CK08EC / Competitor's Column)
Eluent : Milli Q water
Flow rate : 0.6 mL/min
Temperature : 75 °C
Sample Conc : 40mmol/ml each
Injection : 20μL
Detection : RI



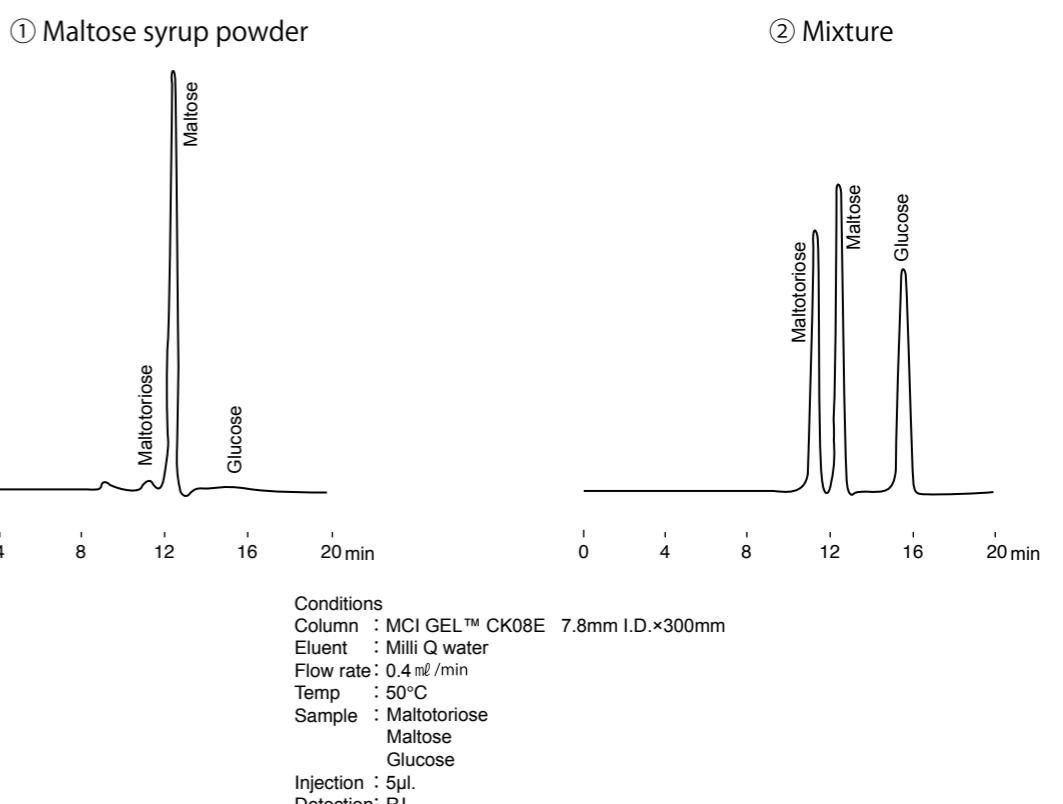
Application data of CK08EC

Fig. 2-11 Analysis of galactose impurity



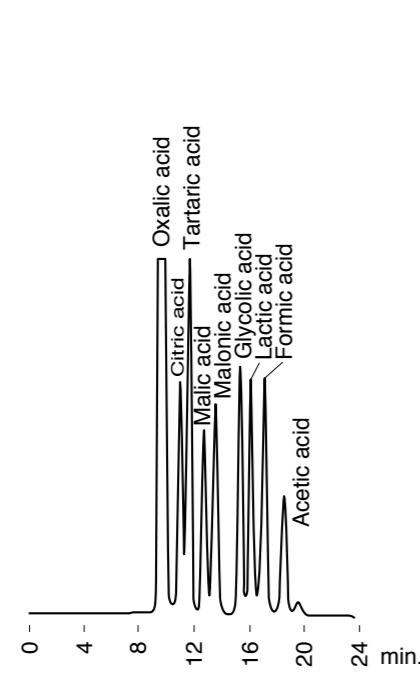
Application data of CK08E

Fig. 2-12 Maltose syrup powder



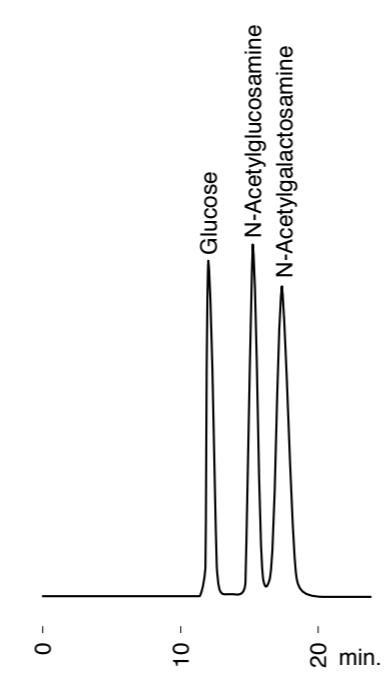
Application data of CK08EH

Fig. 2-13 Carboxylic acids



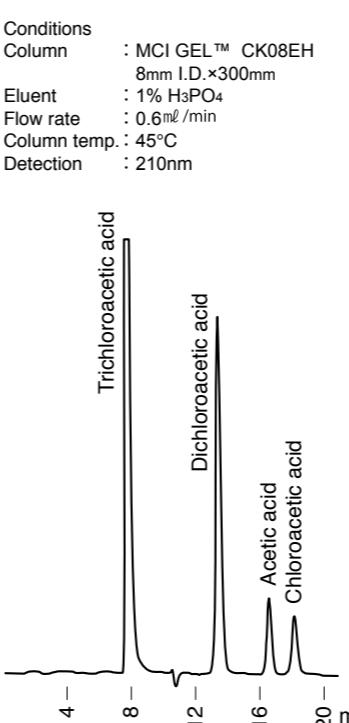
Conditions
Column : MCI GEL™ CK08EH, 8mm I.D.×300mm
Eluent : 1% H₃PO₄ (Fig.2-13,2-14)
Flow rate : 0.6 mL/min
Column temp. : 45°C (Fig. 2-13), ambient (Fig. 2-14)
Detection : 210nm (Fig. 2-13), RI (Fig. 2-14)

Fig. 2-14 Amino sugars



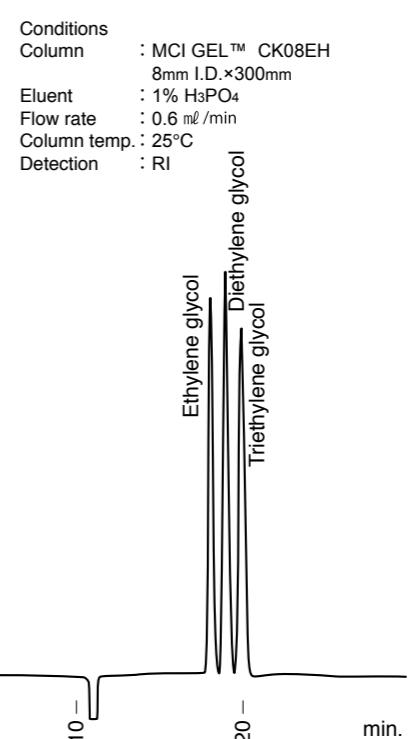
Application data of CK08EH

Fig. 2-16 Chloroacetic acids



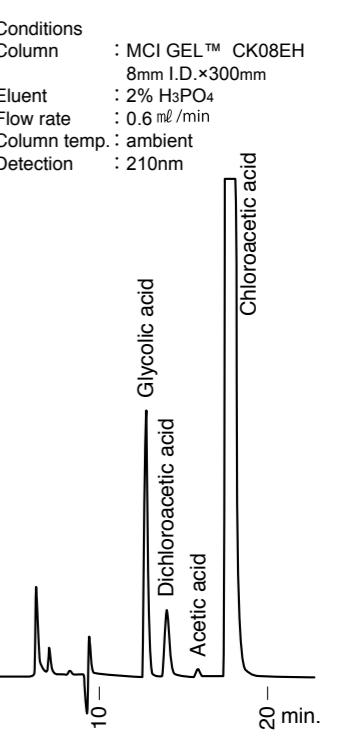
Conditions
Column : MCI GEL™ CK08EH
8mm I.D.×300mm
Eluent : 1% H₃PO₄
Flow rate : 0.6 mL/min
Column temp. : 45°C
Detection : 210nm

Fig. 2-17 Poly alcohols



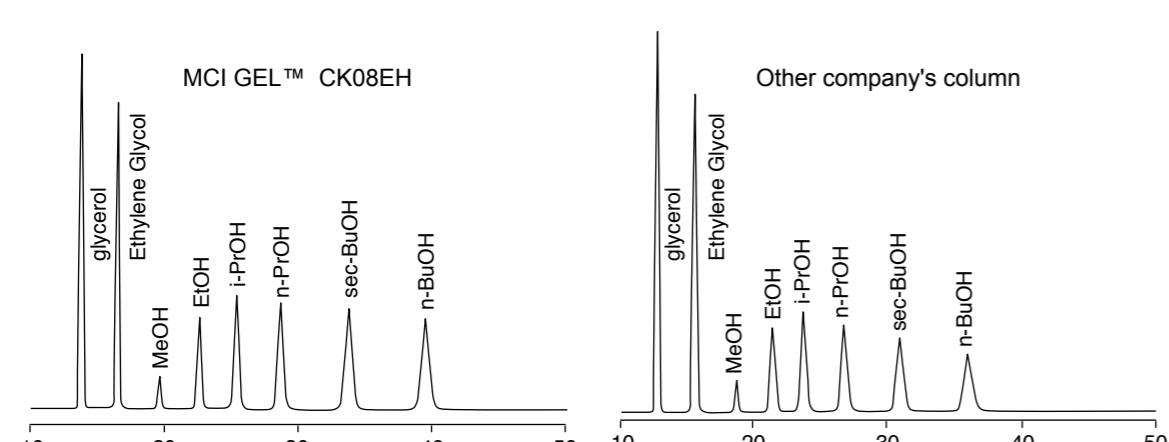
Conditions
Column : MCI GEL™ CK08EH
8mm I.D.×300mm
Eluent : 1% H₃PO₄
Flow rate : 0.6 mL/min
Column temp. : 25°C
Detection : RI

Fig. 2-18 Carboxylic acids



Conditions
Column : MCI GEL™ CK08EH
8mm I.D.×300mm
Eluent : 2% H₃PO₄
Flow rate : 0.6 mL/min
Column temp. : ambient
Detection : 210nm

Fig. 2-15 Alcohols

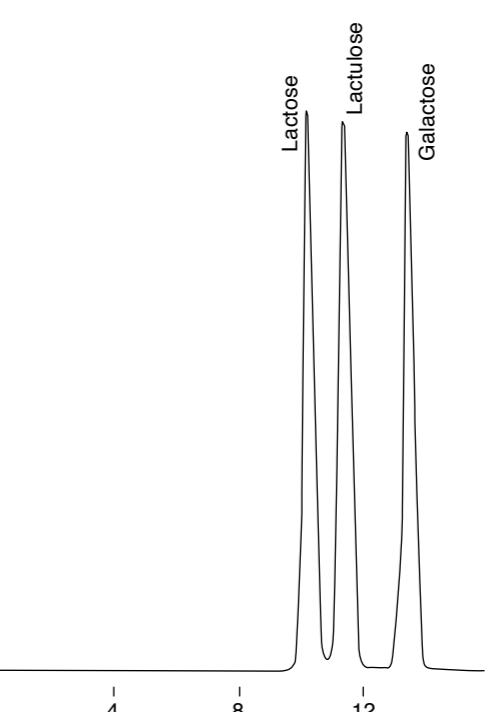


Conditions
Column : MCI GEL™ CK08EH, 7.8mm I.D.×300mm
Eluent : 1% H₃PO₄
Temp : 60 °C
Press : 2.5 bar
Detection : RI
Injection : 10.0 μL

Application data of CK06SC

Fig. 2-19 Lactose

Conditions
Column : MCI GEL™ CK06SC
8mm I.D.×500mm
Eluent : H₂O
Flow rate : 1.0 mL/min
Column temp. : 75°C
Detection : RI
Injection volume : 20μL



Column selection guide

Ion exchange columns and materials

2

Ion chromatography columns and materials

3

Bioseparation columns and materials

4

Analytical and preparative chromatography columns and materials for pharmaceutical applications

5

Chiral separation columns

6

SPE sorbent series

7

MCI GEL™ column list

8

MCI GEL™ material list

10

●Peak retention time for Sugars and Sugar alcohols on various columns [min]

	CK08EC Ca ²⁺	CK08E Na ⁺	CK08ES Ag ⁺
Stachyose	min 9	Stachyose	min 8
Melezitose		Melezitose	* Melezitose
Raffinose		Raffinose	* Stachyose
Gentiobiose	10	Gentiobiose	* Raffinose
Cellobiose		Cellobiose	* Sucrose
Trehalose		Trehalose	Trehalose
Isomaltose		Sucrose	Cellobiose
Sucrose		Isomaltose	Gentiobiose
Maltose		Melibiose	Maltose
Melibiose		Maltose	Isomaltose
Lactose	11	Maltulose	Maltulose
Maltulose		Lactose	Lactose
Lactulose	12	Lactulose	Melibiose
Glucose	13		Lactulose
Xylose	14	Glucose	Adonitol
Galactose		Mannitol	Digitoxose
Mannose		Rhamnose	Rhamnose
Rhamnose	15	Adonitol	Glucose
Fructose	16	Sorbitol	Xylose
Fucose		Digitoxose	Xylitol
Inositol		Mannose	Erythritol
Arabinose		Xylose	Mannitol
Digitoxose		Galactose	Fructose
Adonitol	17	Fructose	Dulcitol
Erythritol	18	Inositol	Xylitol
Mannitol	20	Xylitol	Fucose
Salicin	22	Fucose	Sorbitol
Dulcitol	23	Ribose	Mannose
Xylitol	24	Salicin	Arabinose
Sorbitol			Fucose
Ribose	25		Ribose

Column temp : CK08EC...75°C, CK08E...45°C, CK08ES...75°C

Column size : 8mm I.D.×300mm

Eluent : H₂O

Flow rate : 0.6 mL/min

Sample : 1% aq. solution

Injection vol. : 20μL

* ; These sugars, containing Fructose component, may partially be decomposed by CK08ES and CK08EH.

2 MCI GEL™

CK04S, CK04SS CK02A, CK02AS

Cation exchange columns applications; oligosaccharides

The separation mechanism is based on gel filtration chromatography and elution is achieved via simple distilled water. A larger molecule elutes ahead.



●Separation ability of each column

MCI GEL™ column	Counter ion	Separation ability (degree of polymerization)	USP
MCI GEL™ CK04S	Na ⁺	8~9	L58
MCI GEL™ CK04SS	Ag ⁺	12~13	
MCI GEL™ CK02A	Na ⁺	15~16	L58
MCI GEL™ CK02AS	Ag ⁺	19~20	

Calibration curves of malto-oligosaccharides

Fig. 2-20

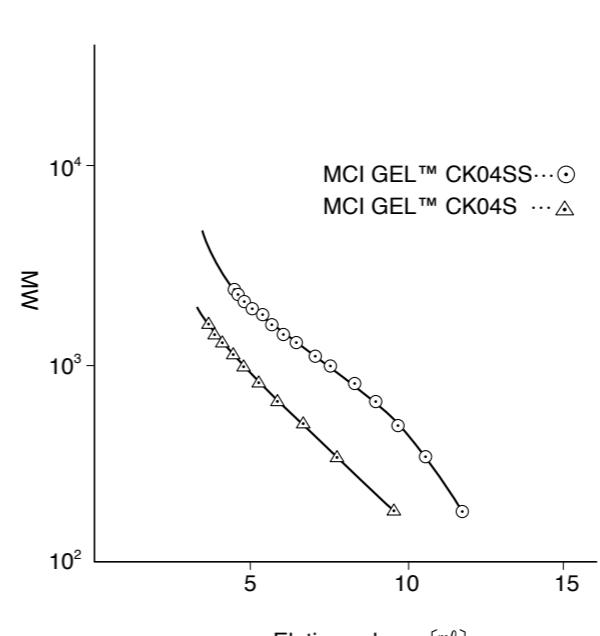
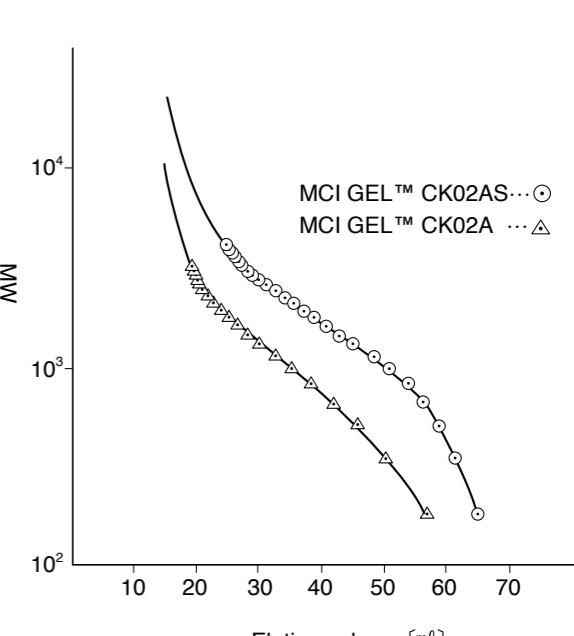
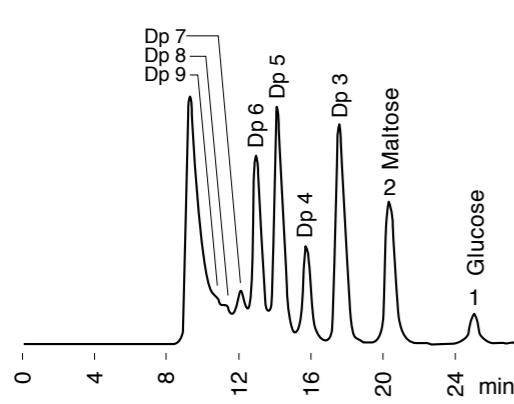
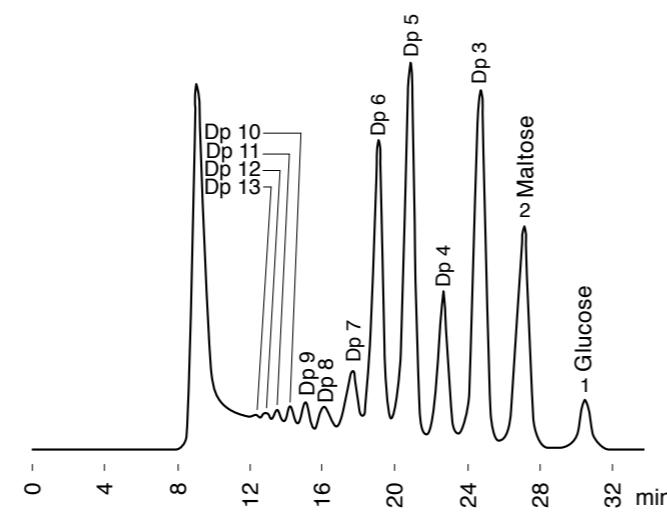
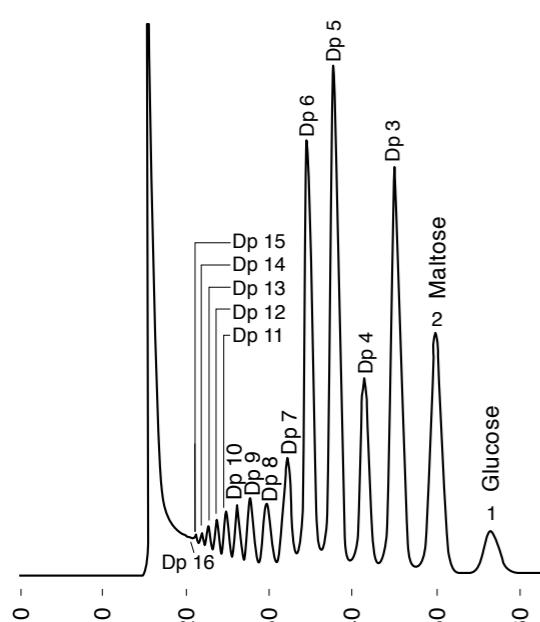
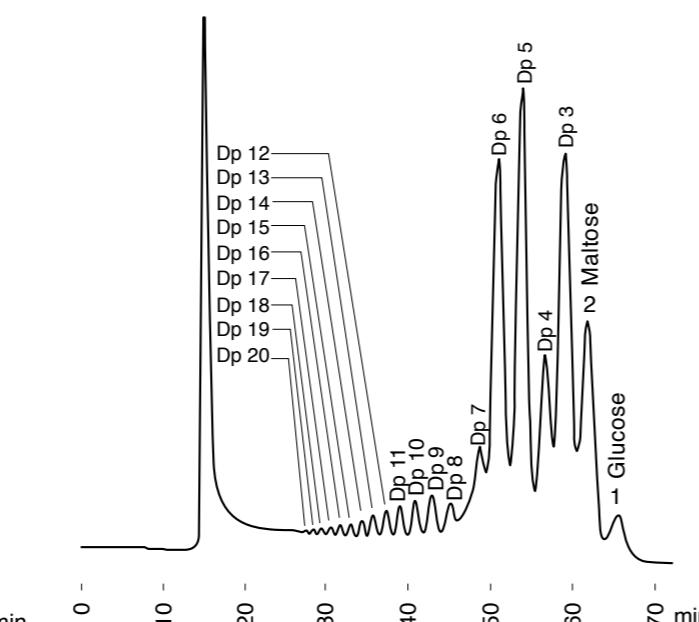


Fig. 2-21

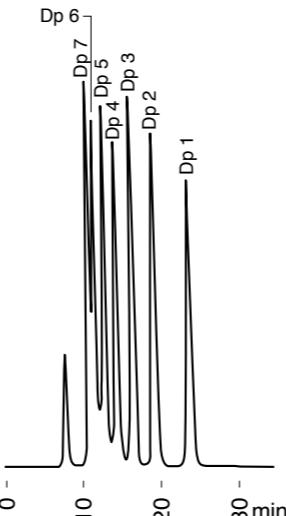
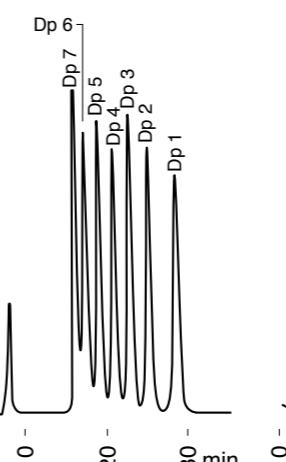
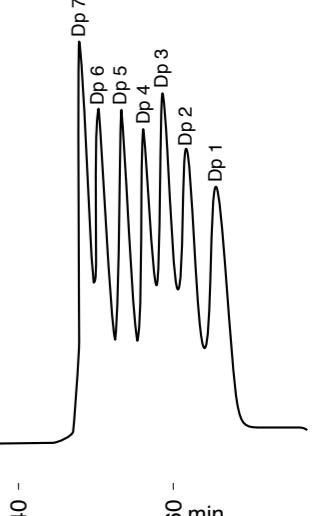


Comparison data of malto-oligosaccharides

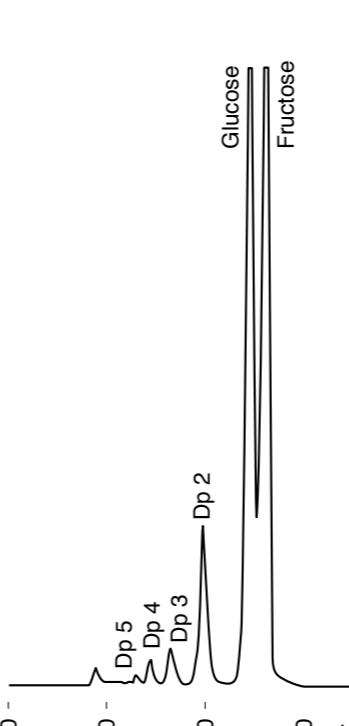
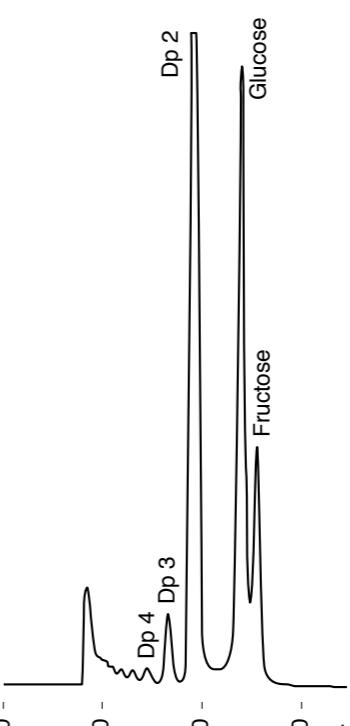
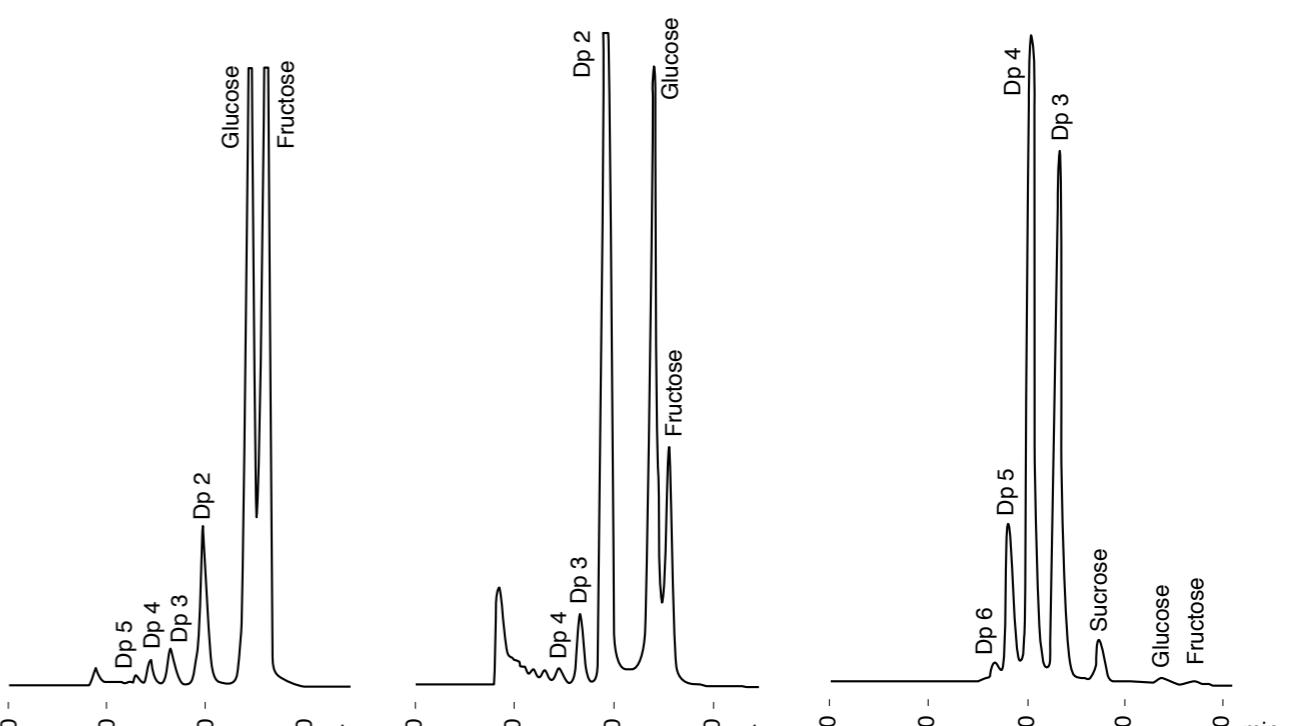
Fig. 2-22 MCI GEL™ CK04S
10mm I.D.×200mm**Fig. 2-23** MCI GEL™ CK04SS
10mm I.D.×200mm**Fig. 2-24** MCI GEL™ CK02A
20mm I.D.×250mm**Fig. 2-25** MCI GEL™ CK02AS
20mm I.D.×250mm

Conditions
Eluent : H₂O
Flow rate : 0.4 mL/min (Fig. 2-22, 2-23, 2-26, 2-27)
1.0 mL/min (Fig. 2-24, 2-25, 2-28)
Column temp. : 85°C
Detection : RI

Comparison data of authentic malto-oligosaccharides samples

Fig. 2-26 MCI GEL™ CK04S
10mm I.D.×200mm**Fig. 2-27** MCI GEL™ CK04SS
10mm I.D.×200mm**Fig. 2-28** MCI GEL™ CK02AS
20mm I.D.×250mm

Application data of CK04S

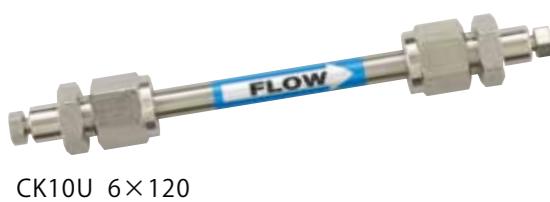
Fig. 2-29 Honey**Fig. 2-30** Jam**Fig. 2-31** Fructo-oligosaccharides

Conditions
Column : MCI GEL™ CK04S
10mm I.D.×200mm
Eluent : H₂O
Flow rate : 0.4 mL/min (Fig. 2-29, 2-30) 0.3 mL/min (Fig. 2-31)
Column temp. : 85°C (Fig. 2-29, 2-30) 45°C (Fig. 2-31)
Detection : RI

2 MCI GEL™

CK10U

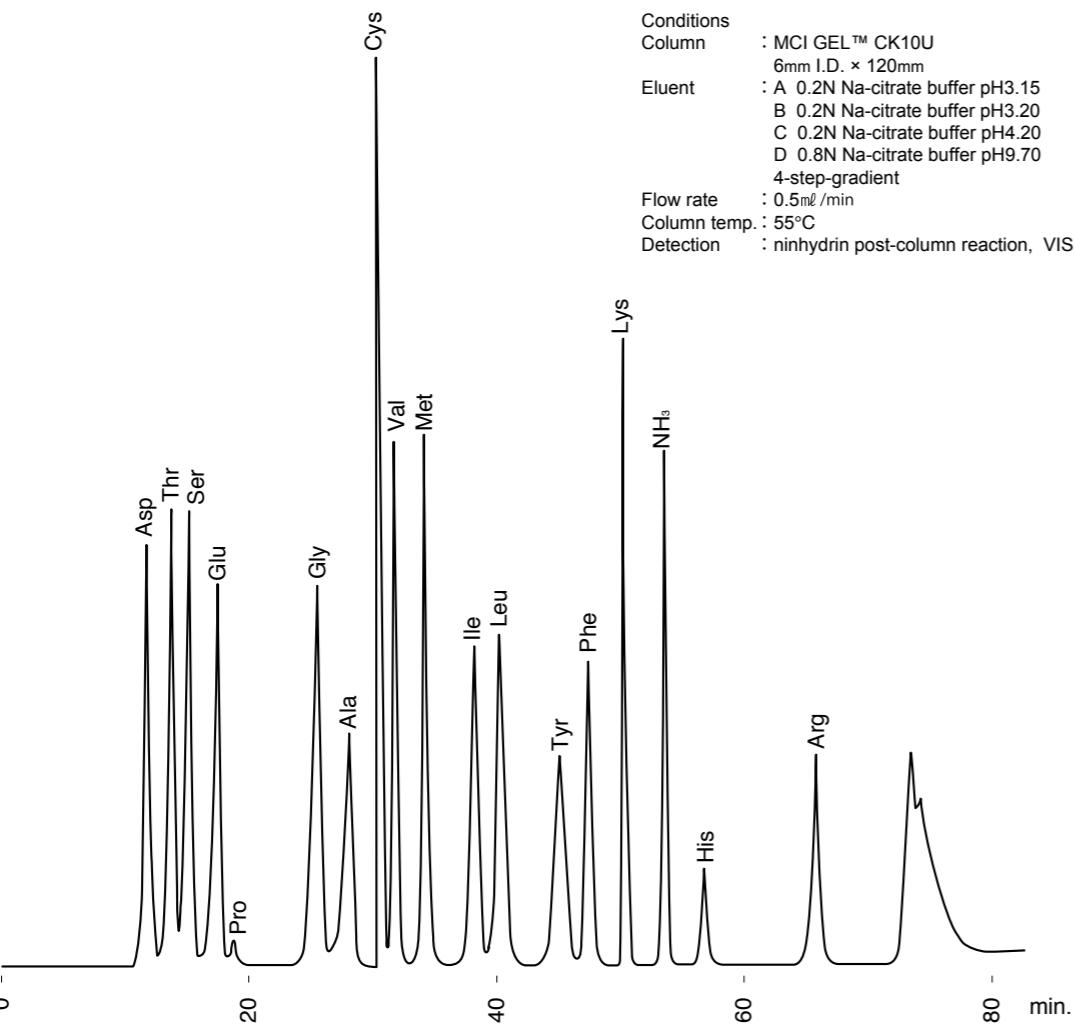
High cross linkage cation exchange column applications; amino acids, amines, etc



CK10U 6×120

Separation of amino acids

Fig. 2-32 Protein hydrolyzates amino acids



As for analysis of amino acids by a cation exchange column such as MCI GEL™ CK10U, MCI GEL™ AFR2-PC is recommended as a pre-column. The AFR2-PC column is very effective to stabilize base line because ammonium in eluent is trapped in this column. The ammonium ion may disturb base line stability. The AFR2-PC should be installed between an outlet of HPLC pump and an inlet of sample injector. A gradient elution, commonly used for amino acid analysis, is influenced by HPLC instrument. So to obtain a satisfactory chromatogram, gradient conditions should be optimized in accordance with the HPLC equipment.

Separation of amino acids

Fig. 2-33 Valine, β -Alanine

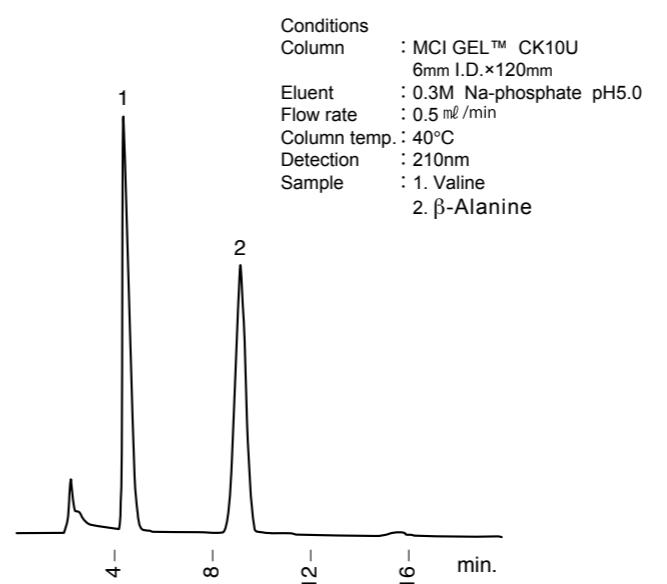


Fig. 2-34 γ -Aminobutyric acid

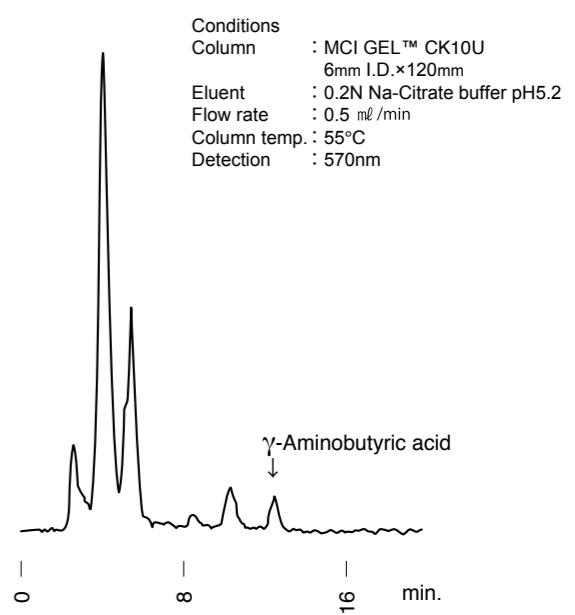


Fig. 2-35 cyclic amino acids

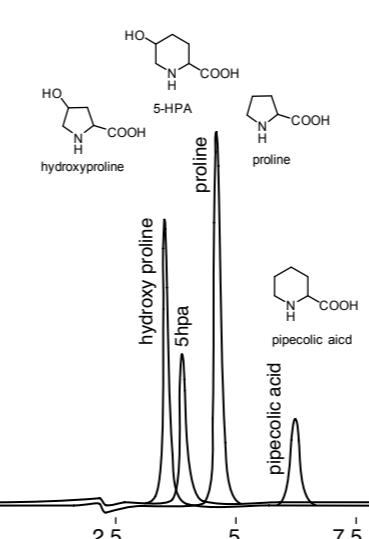


Fig. 2-36 alkyl amino acid

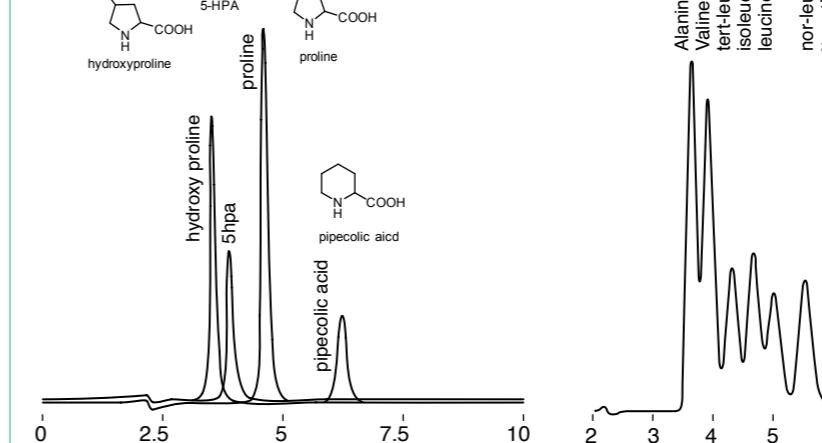
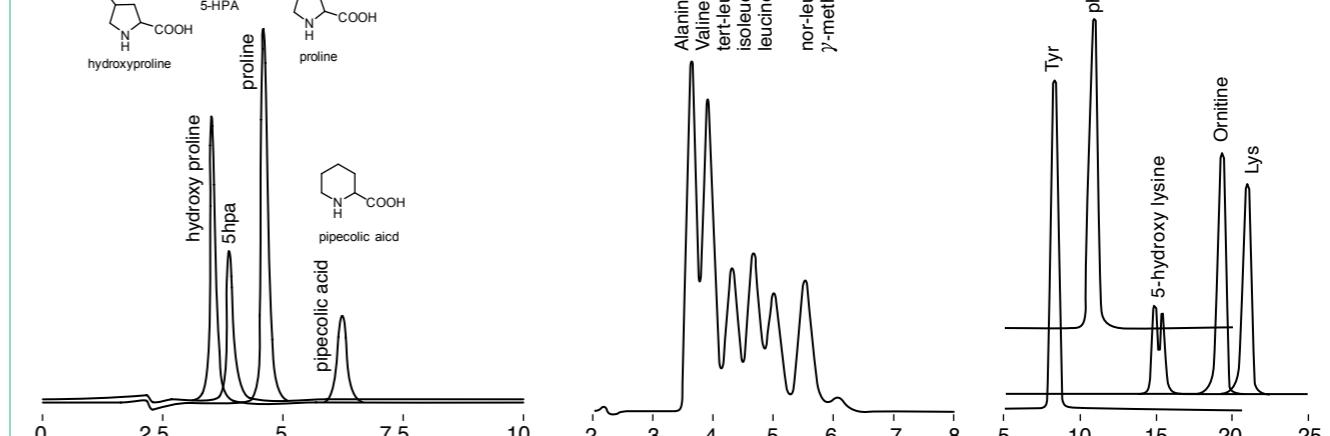


Fig. 2-37 basic amino acid and aromatic amino acids

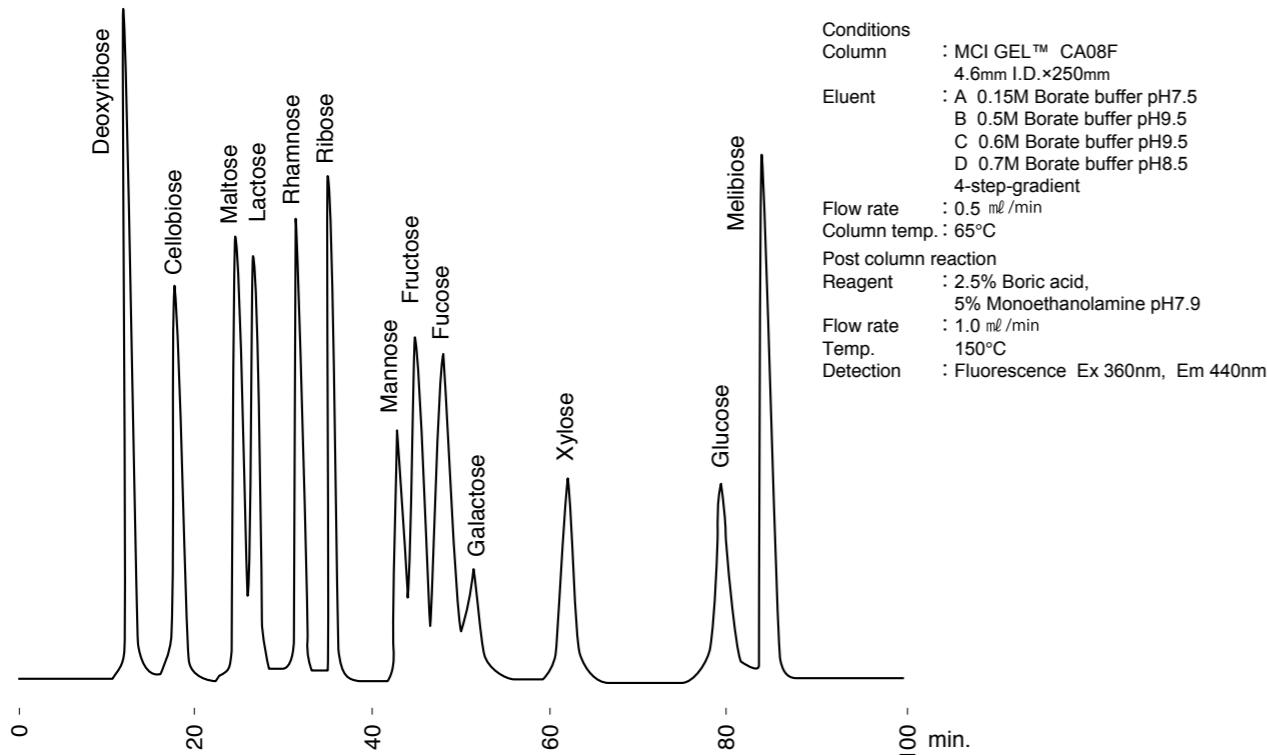
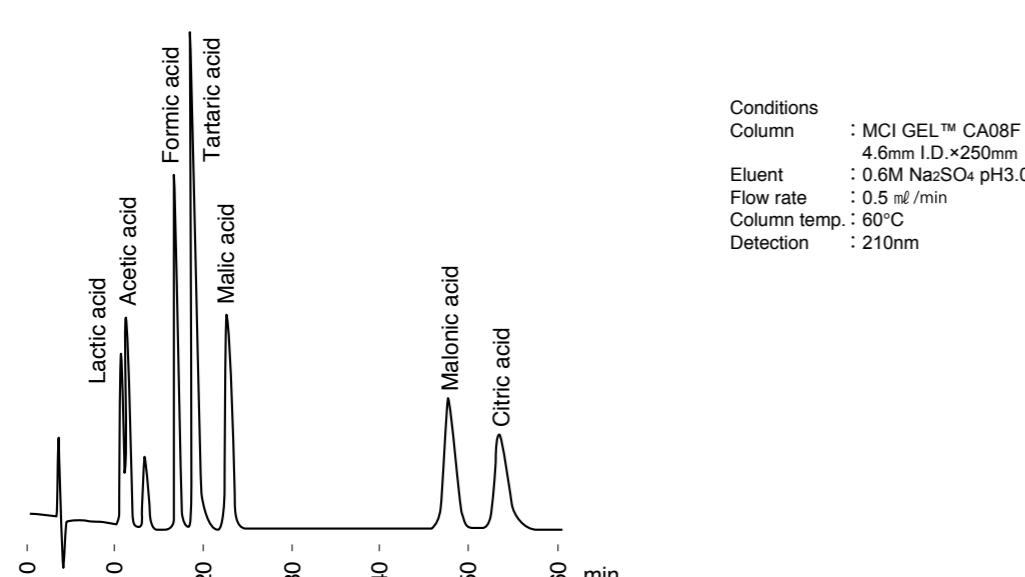
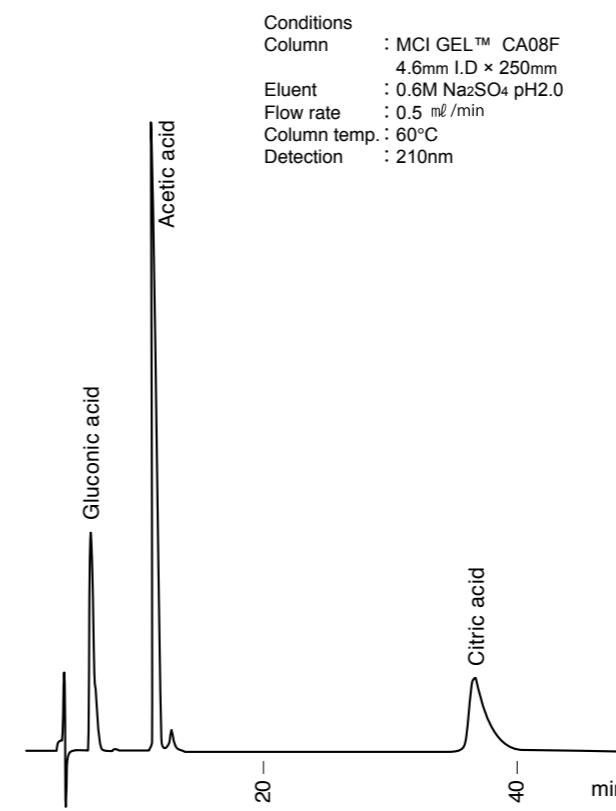
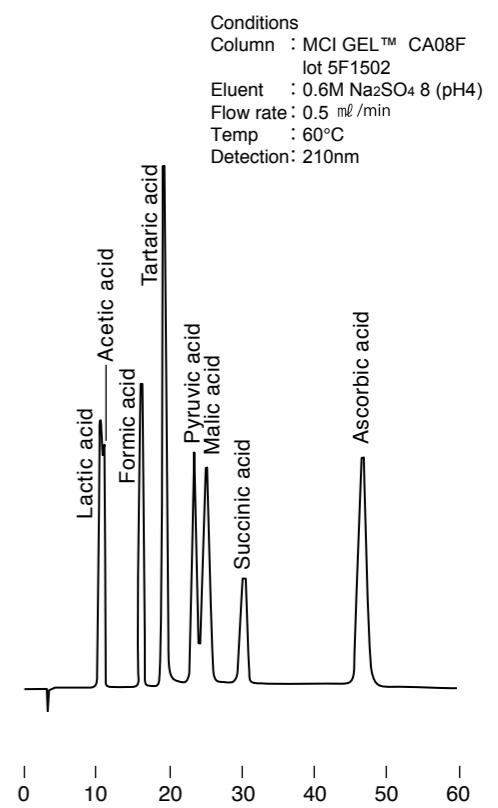
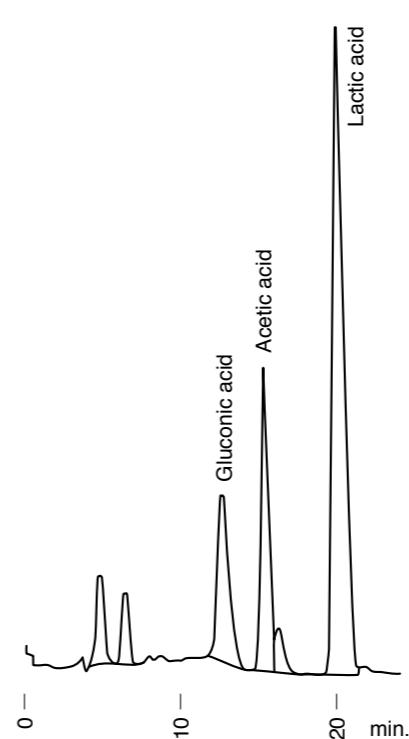


Conditions
Column : MCI GEL™ CK10U
6mm I.D. × 120mm
Eluent : 0.3M NaSO₄ (pH5.7)
Flow : 0.5ml/min
Temp. : 60°C
Detection: UV210nm

CA08F**Anion exchange column applications; sugars, carboxylic acids, nucleotides**

MCI GEL™ CA08F packed column has been designed for the analysis of nucleotides, sugars, and organic acids by anion exchange chromatography mode.

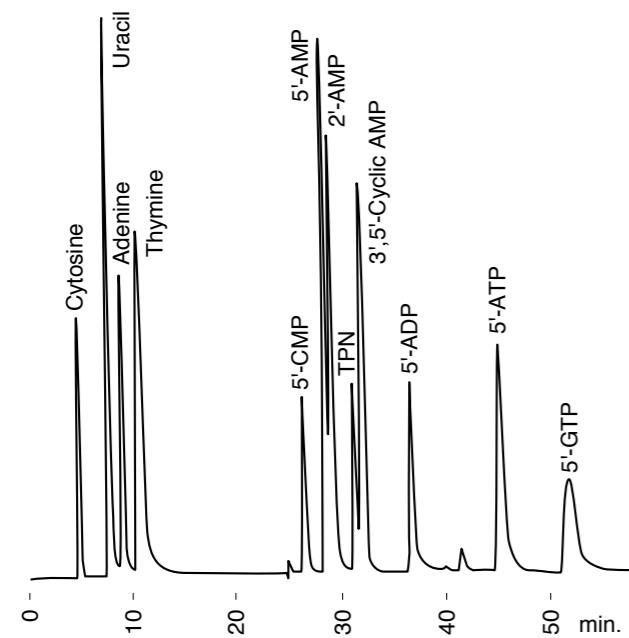
This column will provide excellent separation and short analysis time.

Application data of CA08F**Fig. 2-38 Sugars****Fig. 2-39 Carboxylic acids****Application data of CA08F****Fig. 2-40 Carboxylic acids****Fig. 2-41 Organic acid****Fig. 2-42 Carboxylic acids**

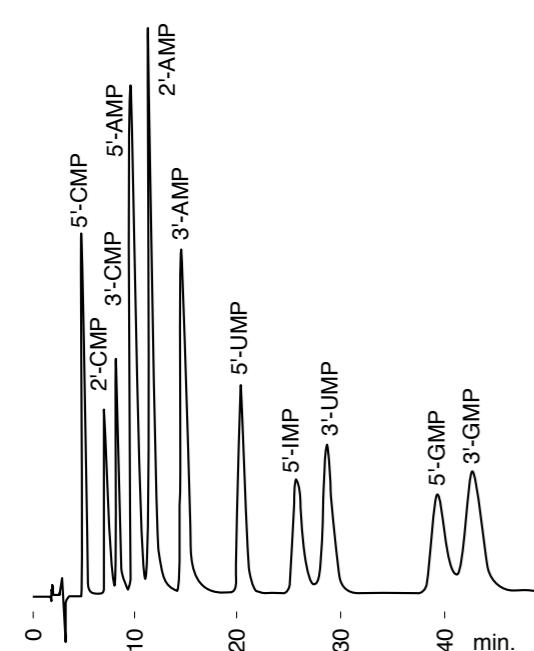
CDR10

High porous type anion exchange column

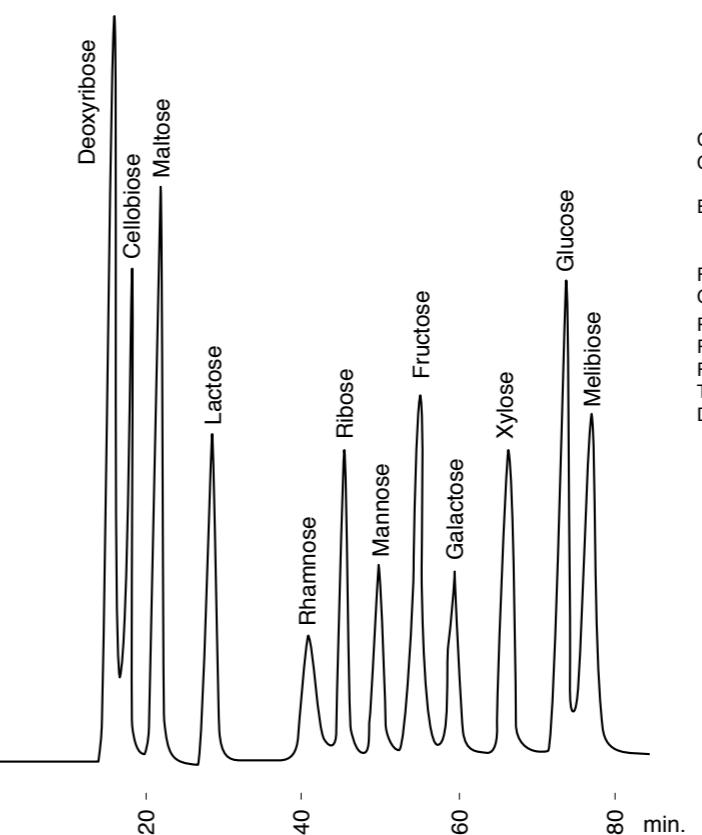
Packing material of MCI GEL™ CDR10 column is based on a high porous polystyrene functionalized with a quaternary ammonium anion exchange resin. Since a high porous type ion exchange resin is rigid, CDR10 allows usage of aggressive gradient elution, for example water to 6M of acetate buffer gradient. MCI GEL™ CDR10 is highly recommended for rapid analysis of physiological fluids like urine and blood.

Application data of CDR10**Fig. 2-43 Nucleic acids and related substances**

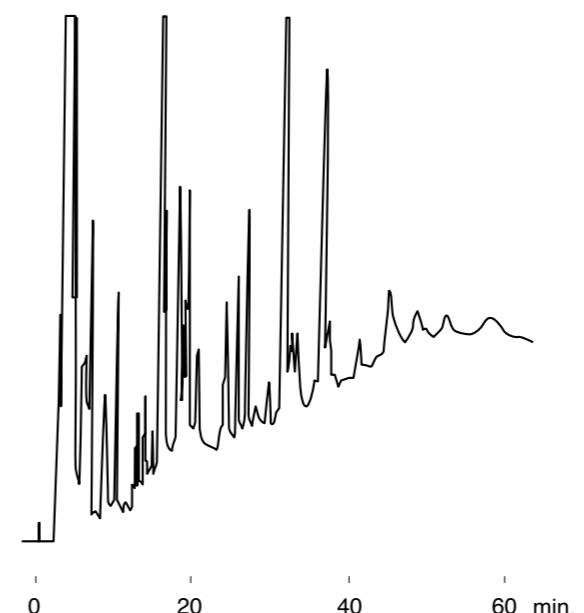
Conditions
Column : MCI GEL™ CDR10
4.6mm I.D.×250mm
Eluent : A H₂O
B 6M Acetate buffer pH4.4
A→B 30min linear gradient
Flow rate : 0.5 mL/min
Column temp. : 60°C
Detection : 254nm

Fig. 2-44 Mono-nucleotides

Conditions
Column : MCI GEL™ CDR10
4.6mm I.D.×250mm
Eluent : 1M Acetate buffer pH3.3
Flow rate : 1.2 mL/min
Column temp. : 60°C
Detection : 254nm

Application data of CDR10**Fig. 2-45 Sugars**

Conditions
Column : MCI GEL™ CDR10
4.6mm I.D.×250mm
Eluent : A 0.15M Borate buffer pH7.5
B 0.6M Borate buffer pH9.5
A→B 60min linear gradient
Flow rate : 0.5 mL/min
Column temp. : 65°C
Post column reaction : 2.5% Boric acid, 5% Monoethanolamine pH7.9
Reagent : 0.5
Temp. : 150°C
Detection : Fluorescence Ex 360nm, Em 440nm

Fig. 2-46 Human urine

Conditions
Column : MCI GEL™ CDR10
4.6mm I.D.×250mm
Eluent : A 0.006M Acetate buffer pH4.4
B 6M Acetate buffer pH4.4
A→B 60min. linear gradient
Flow rate : 1.0 mL/min
Column temp. : 60°C
Detection : 254nm

MCI GEL™

3

Ion chromatography columns and materials

- Cation chromatography column MCI GEL™ SCK01
- Cation chromatography column MCI GEL™ CHK45/C05
- Anion chromatography column MCI GEL™ SCA04

The MCI GEL™ ion chromatography columns are based on surface functionalized cation and anion exchange resins designed for non-suppressed ion chromatography applications. The non-suppressed ion chromatography is an analysis technique of cations and anions with combination of a packed column of low capacity ion exchange resin and low concentration of electrolyte solution as an eluent. The advantage of the ion chromatography is that several ions can be analyzed by only one injection with free of complicated sample pre-treatment.

Cation chromatography column MCI GEL™ SCK01

Packing material of MCI GEL™ SCK01 is crosslinked polystyrene functionalized with sulfonic acid. This column is characterized by excellent resolution and rapid analysis for monovalent and divalent cations. Standard monovalent cations like Li⁺, Na⁺, NH₄⁺, K⁺, Rb⁺, Cs⁺ and simple amines such as mono-, di- and trimethylamine can be resolved using a nitric acid solution as eluent. Divalent cations, such as alkaline earth metals and transition metal elements, can be efficiently resolved using tartaric acid and complexing reagent such as ethylene diamine to selectively elute the metals from the column.

■ Note:

When using the MCI GEL™ SCK01 column for monovalent cations, it is recommended that a pre-column, MCI GEL™ SCK-PC, be used to trap heavy metals which might otherwise poison the SCK01 column resulting in a rapid loss of capacity and chromatographic performance.

Cation chromatography Column MCI GEL™ CHK45/C05

Packing material of MCI GEL™ CHK45/C05 is made of crosslinked polymethacrylate functionalized with carboxylic acid. By use of simple eluent system, MCI GEL™ CHK45/C05 can separate both monovalent and divalent cations in tap water, river water and other environmental water samples.

Anion chromatography column MCI GEL™ SCA04

Packing material of MCI GEL™ SCA04 is based on a hydrophilic vinyl polymer matrix functionalized with quaternary ammonium group and particle size of 5 µm. A solution of potassium hydrogen phthalate and a vanilic acid (VA)/N-methyldiethanolamine (MDEA) solution both can be used as a mobile phase. The unique VA/MDEA eluent, is developed for the SCA04 column, which allows users to determine 7 standard anions in 14 minutes without system peak.

■ Note:

A pre-column, MCI GEL™ SCA-PC is recommended for prevention of contamination to the SCA04 column when the VA/MDEA eluent is used. The SCA-PC is effectively prolong SCA04 column life. The SCA-PC should be installed between an outlet of HPLC pump and an sample injector.



SCA04 4.6×150 PEEK

Column list

Cation analysis	MCI GEL™ SCK01	6mm I.D×50mm	Stainless steel column
Cation analysis	MCI GEL™ SCK01	4.6mm I.D×150mm	Stainless steel column
Pre-column for cation analysis	MCI GEL™ SCK-PC	6mm I.D×50mm	Stainless steel column
Cation analysis	MCI GEL™ CHK45/C05	4.6mm I.D×150mm	Stainless steel column
Anion analysis	MCI GEL™ SCA04	4.6mm I.D×150mm	PEEK column
Pre-column for anion analysis	MCI GEL™ SCA-PC	8mm I.D×10mm	Stainless steel column

*USP L31 column

Application data of SCK01

Fig. 3-1 Monovalent cations

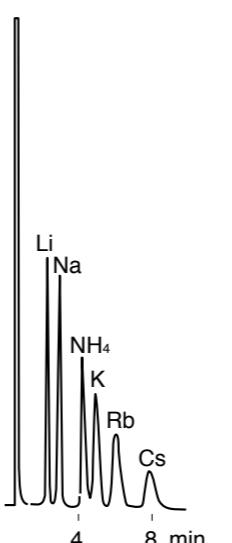


Fig. 3-2 Amines

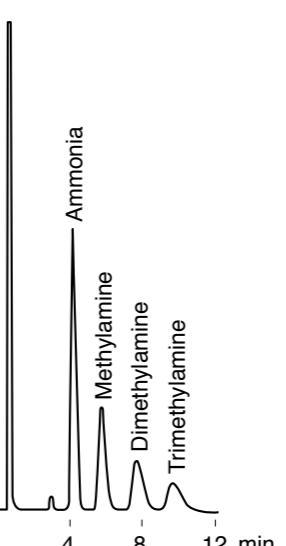


Fig. 3-3 Monovalent cations in rain

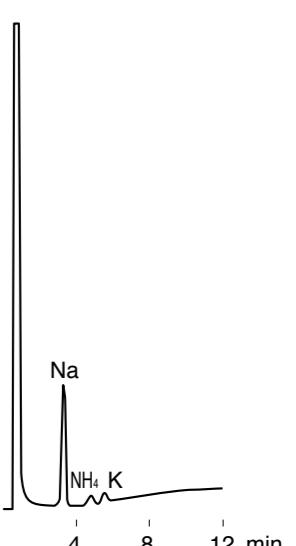


Fig. 3-4 Monovalent cations in tap water

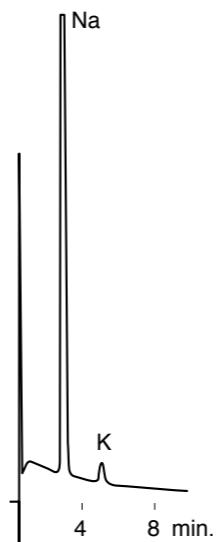


Fig. 3-5 Sports drink

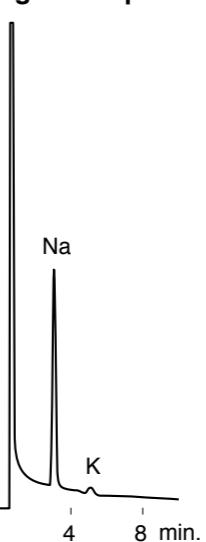
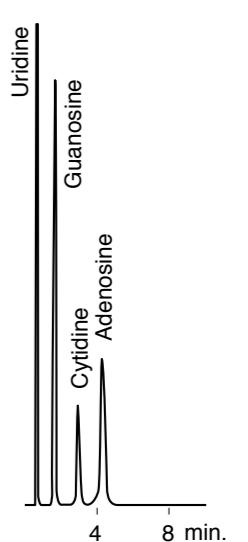
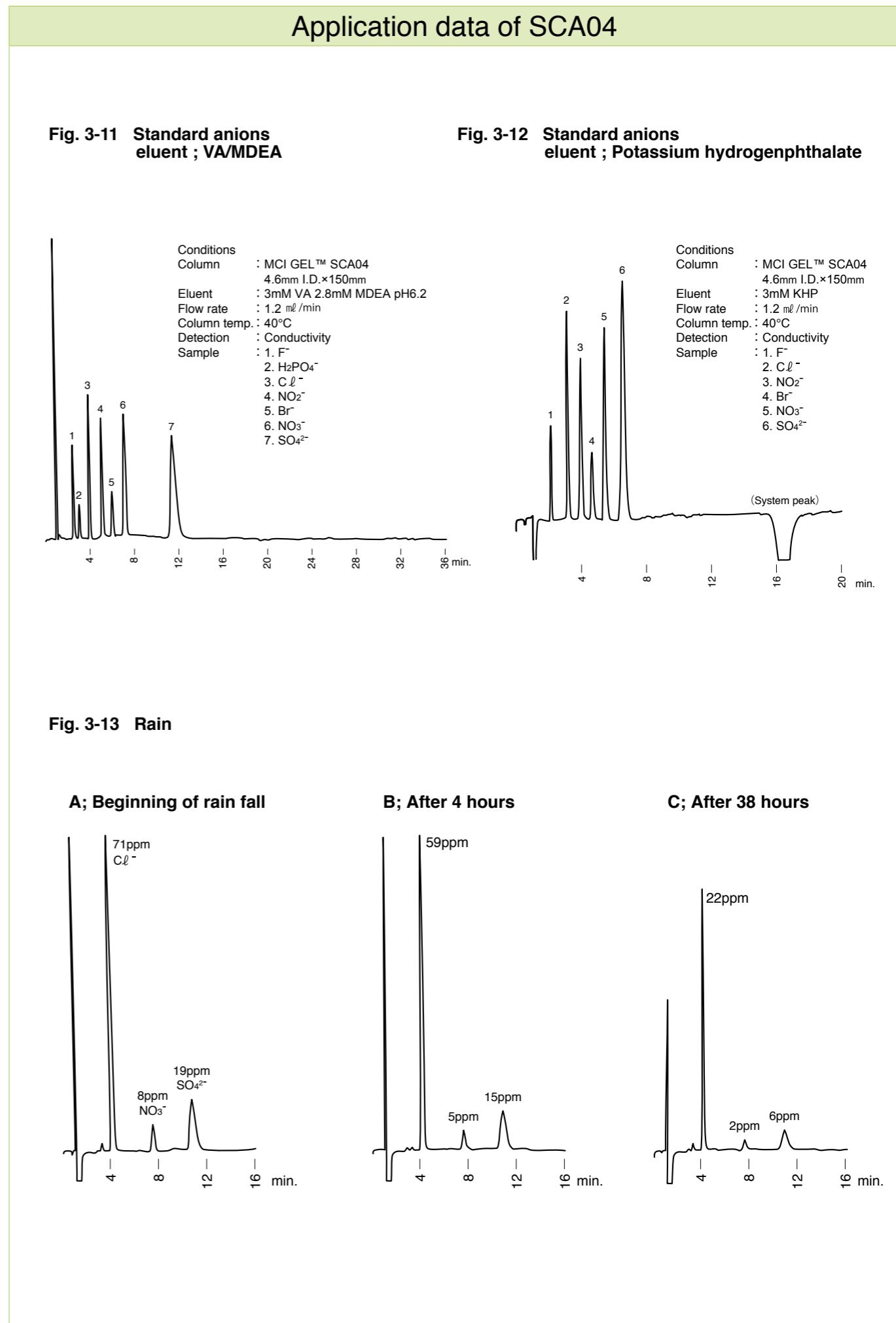
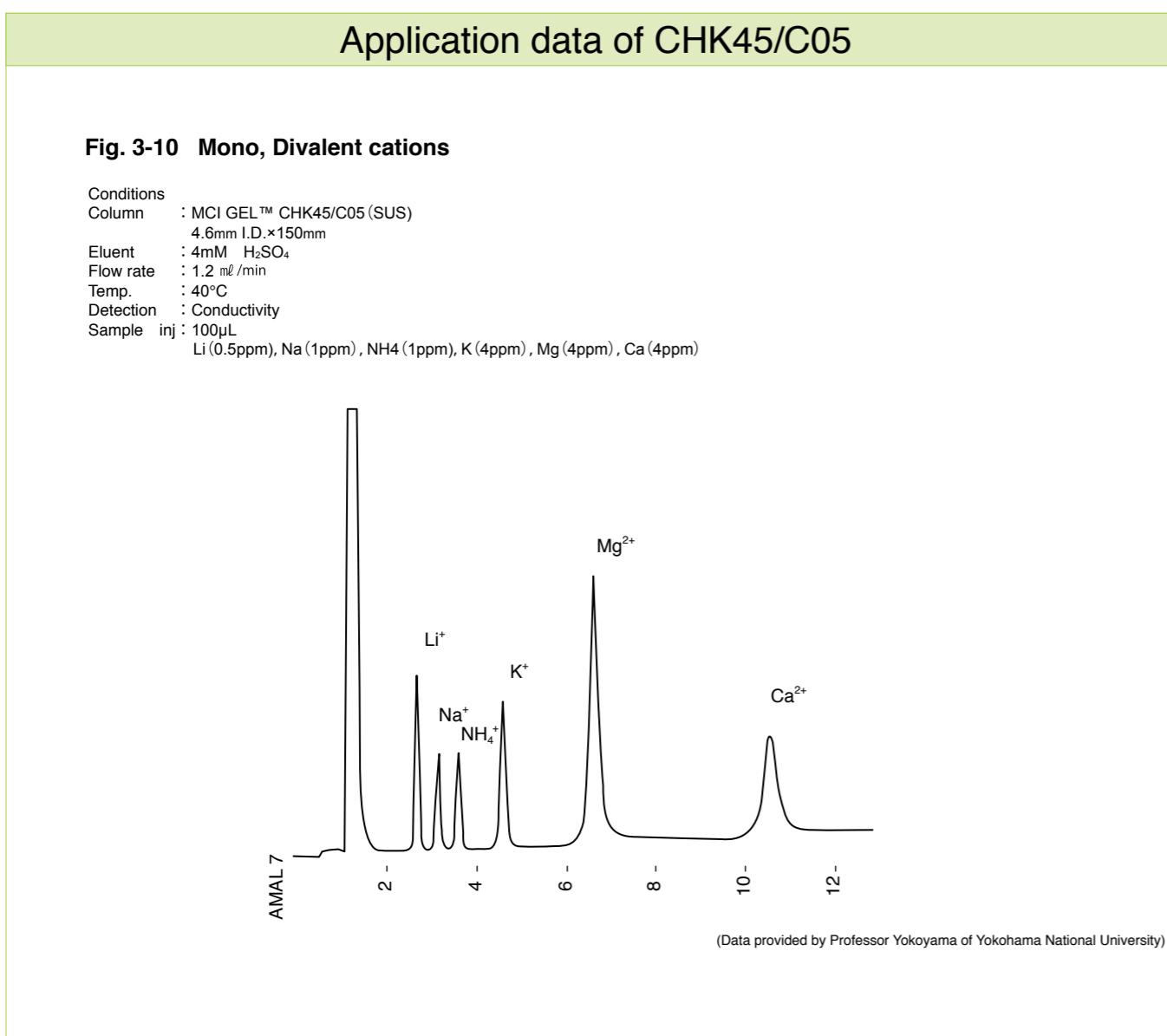
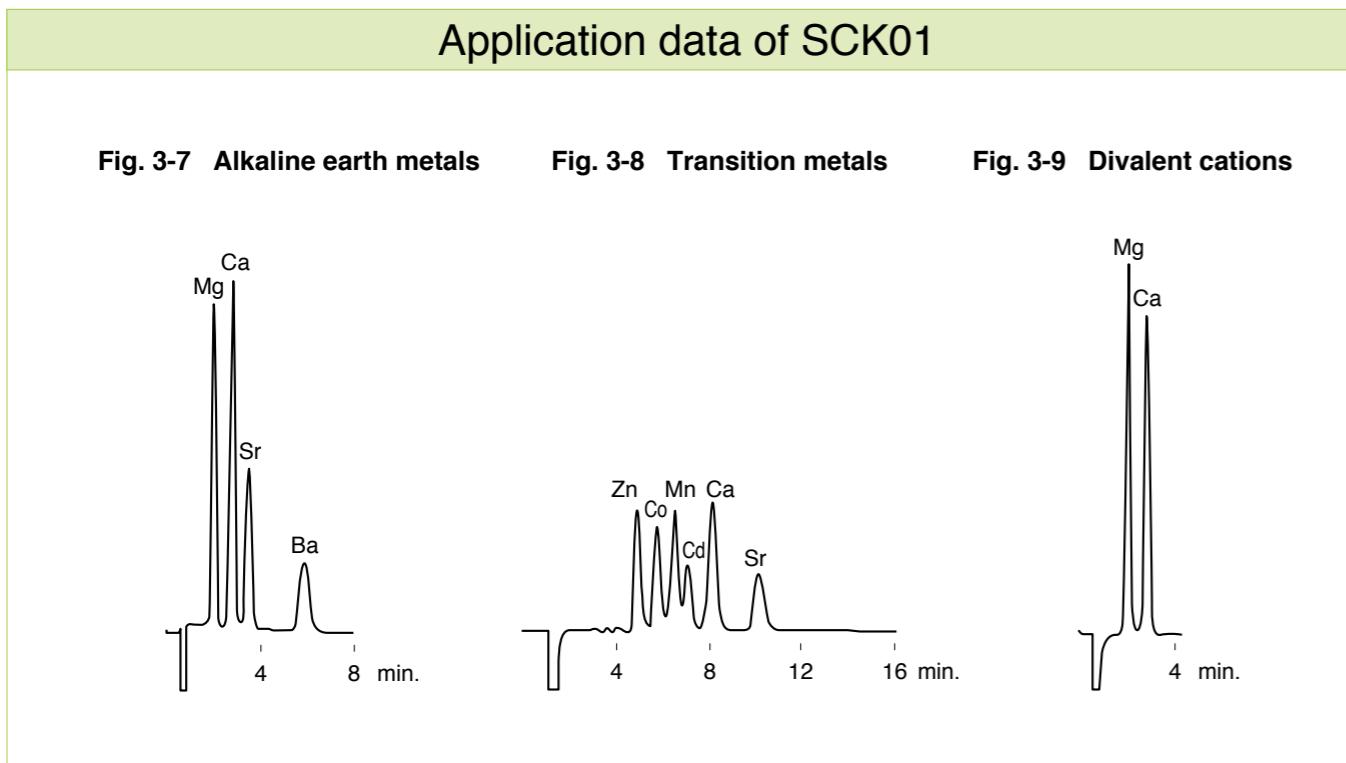


Fig. 3-6 Nucleoside



Conditions
Column : MCI GEL™ SCK01 6mm I.D.×50mm
Eluent : 5mM HNO₃
Flow rate : 1.0 mL/min
Column temp. : 40°C
Detection : Conductivity (Fig. 3-1, 3-2, 3-3, 3-4, 3-5) 254nm (Fig. 3-6)



Application data of SCA04

Fig. 3-14 River water

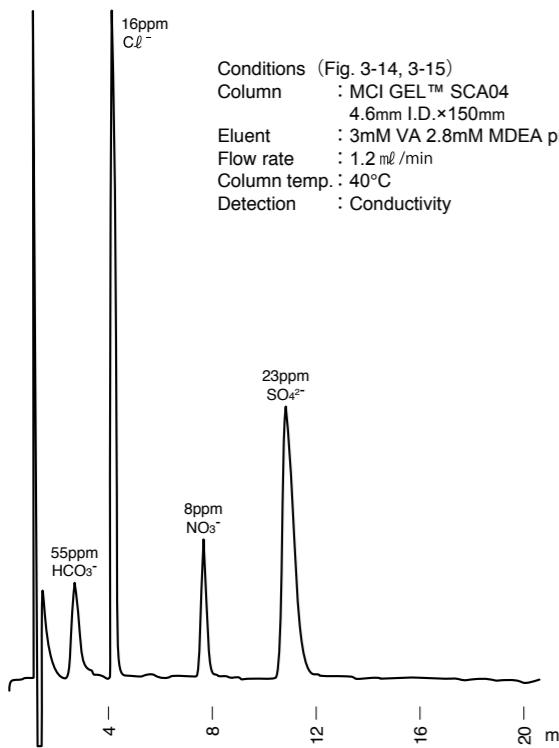


Fig. 3-15 Sulfur compounds

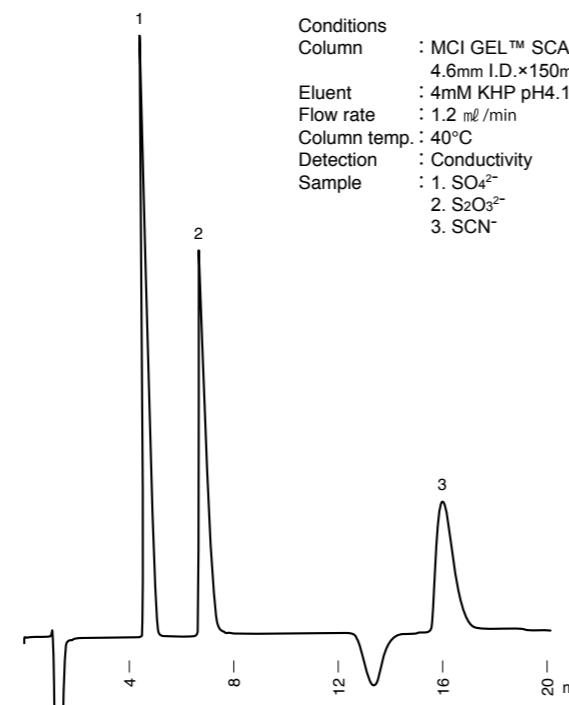
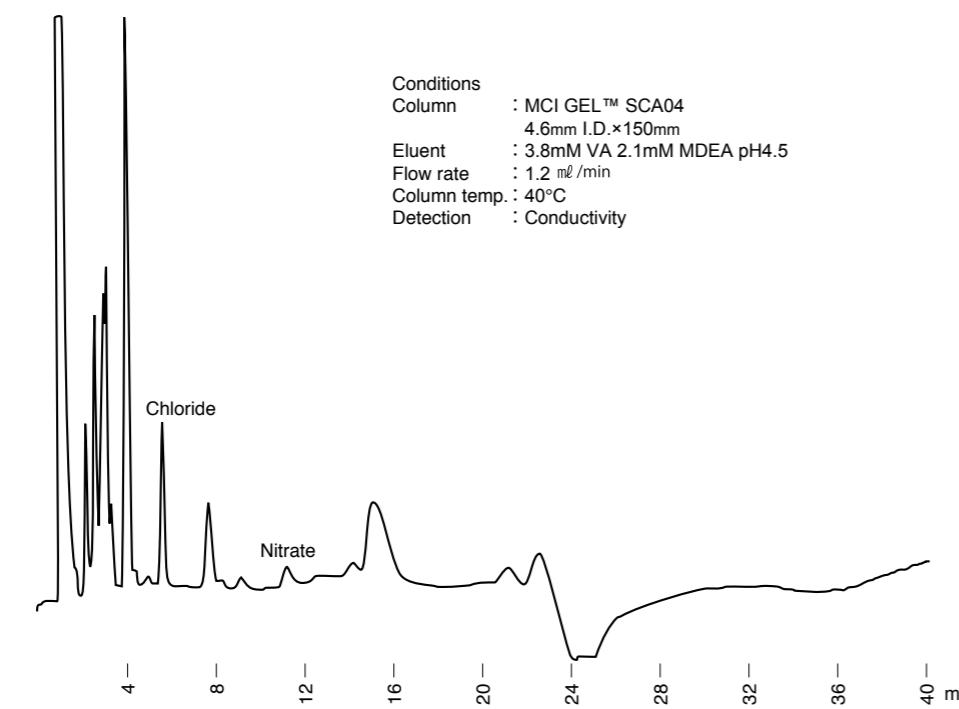


Fig. 3-16 Instant coffee

Ion-exchange chromatography columns
XtalSpeed™ seriesSize-exclusion chromatography columns
MCI GEL™ CQP series

Bioseparation columns

MCI GEL™ bioseparation columns are based on a hydrophilic, wide-pore, and rigid polymer designed for analytical chromatography of proteins, peptides, enzymes, and other biomolecules.

MCI GEL™ CQP series are used for size-exclusion chromatography.

XtalSpeed™ series are ion-exchange columns used for protein purification. High-quality target proteins are obtained with this column at a high recovery rate. XtalSpeed™ series are used for both analytical and preparative purpose in protein crystallography and NMR research.

XtalSpeed™ series are also used for antibody variant analysis and protein isoform analysis.

Column name	USP	Separation mode	Functional Group
XtalSpeed™ SP01	—	Cation exchange	Sulfopropyl(SP)
XtalSpeed™ DA01	—	Anion exchange	Diethyl amino ethyl(DEAE)
XtalSpeed™ CM01	—	Cation exchange	carboxymethyl(CM)
XtalSpeed™ Q01	—	Anion exchange	Quaternary ammonium(QA)
MCI GEL™ CQP06	L25	Size exclusion	—
MCI GEL™ CQP10	L38	Size exclusion	—
MCI GEL™ CQP30	L37, L38	Size exclusion	—



4 MCI GEL™

XtalSpeed™ series

Ion exchange chromatography columns

Ion exchange chromatography columns

XtalSpeed™ series columns are ion-exchange columns used for protein purification. They have been designed especially for protein crystallography and NMR research, and enable to purify target proteins with high quality at a high recovery rate and in a very short time and obtain protein crystals for further analysis.

We developed hydrophilic and chemically stable polymer layers based on highly porous polymer beads, reducing non-specific binding to the lowest level.

To eliminate other interactions and allow target proteins participate only in the ion-exchange mechanism, this column was able to separate similar proteins that other columns never succeeded to separate.

Even under large sample loading, this column maintains excellent selectivity. Taking these aspects into consideration, XtalSpeed™ series can be used as preparative columns for protein.

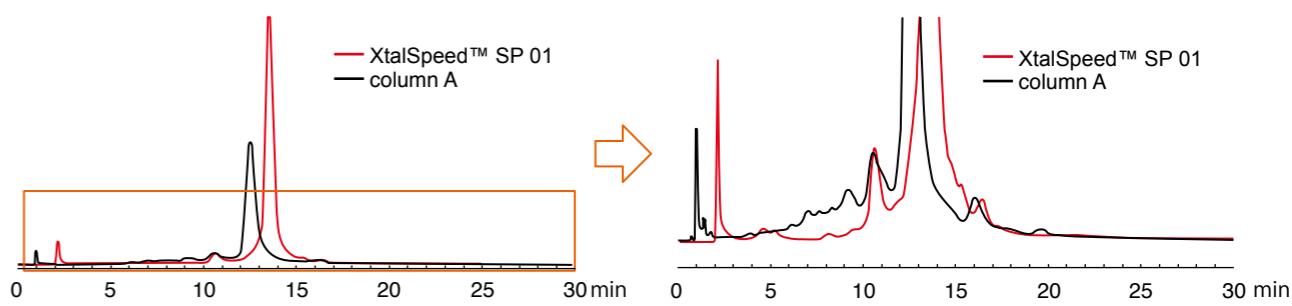
XtalSpeed™ SP01 is also used for antibody variant analysis.

Column list

Column name	Column size	Code	Housing	Functional Group
SP01	Φ4.6mm×50mm	0-047-11	PEEK	Sulfopropyl (SP)
	Φ4.6mm×100mm	0-047-12		
	Φ7.5mm×100mm	0-047-13		
	Φ11.5mm×100mm	0-047-14		
DA01	Φ4.6mm×50mm	0-047-01	PEEK	Diethylaminoethyl (DEAE)
	Φ4.6mm×100mm	0-047-04		
	Φ7.5mm×100mm	0-047-02		
	Φ11.5mm×100mm	0-047-03		
CM01	Φ4.6mm×50mm	0-047-31	PEEK	Carboxymethyl (CM)
	Φ4.6mm×100mm	0-047-32		
	Φ7.5mm×100mm	0-047-33		
	Φ11.5mm×100mm	0-047-34		
Q01	Φ4.6mm×50mm	0-047-21	PEEK	Quaternary ammonium (QA)
	Φ4.6mm×100mm	0-047-22		
	Φ7.5mm×100mm	0-047-23		
	Φ11.5mm×100mm	0-047-24		

Application data of XtalSpeed™ series

Fig. 4-1 Analysis of Rituximab



Conditions
Column : XtalSpeed™ SP01
100 x 4.6mm.I.D
Column A (250 x 4mm.I.D)
Eluent : A, 20mM Na phosphate (pH7.0)
B, A + 1.0M NaCl
Flow rate : 0.529mL/min for XtalSpeed and 1.0 mL/min for ProPac
Gradient : (A), 2.5-20.0%B over 30min + 20.0-100%B over 5min
(C), 2.5-5.0%B over 30min + 5.0-100%B over 5min

Fig. 4-2 Analysis of Hemoglobin A1C

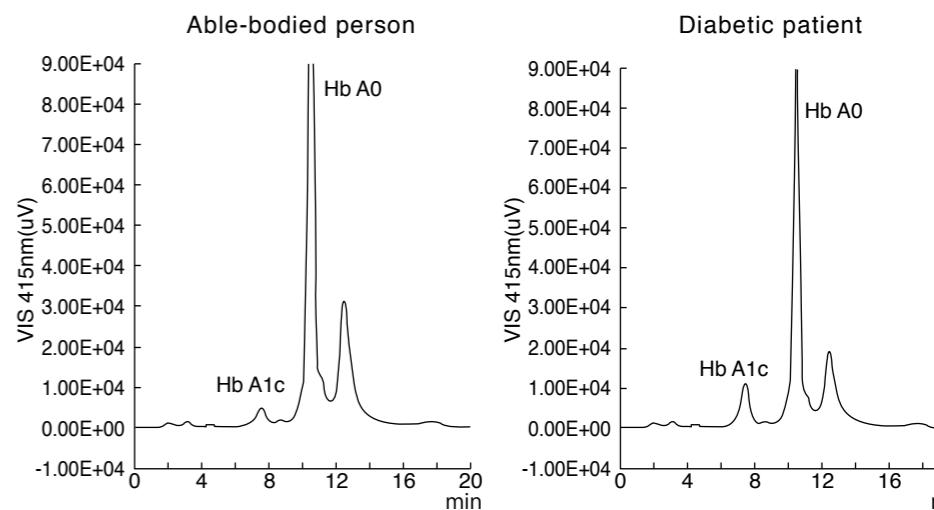
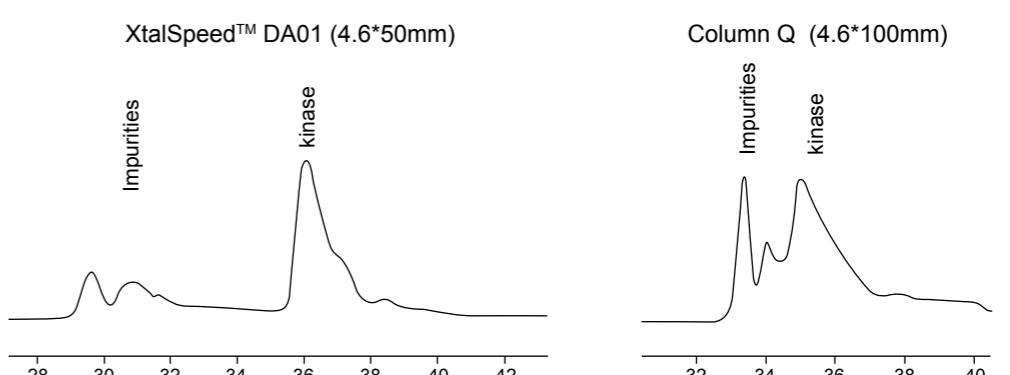


Fig. 4-3 Comparison of loadability



Conditions
Eluent : Buffer A:50mM HEPES-NaOH(pH7)
Buffer B:50mM HEPES-NaOH(pH7),1M NaCl
Flow rate : XtalSpeed™ DA01 :0.6 mL/min
Competitor's Column Q:1 mL/min
Gradient : 0-40% B (0-400mM NaCl)/40CV
Sample : 50 micro grams of human kinase

4 MCI GEL™

CQP series

Aqueous size exclusion columns

Size exclusion chromatography columns

Size exclusion chromatography is a liquid chromatographic technique which separates solute molecules according to their size in solution. The column is packed with porous particles and separation takes place as a result of the differential solute distribution outside and within the pores of the packing material. Solute molecules which are larger than the pores of the packing material will be excluded and therefore will elute first and have a lower retention time than the smaller one. The CQP series columns based on a hydrophilic polymer are designed for analysis of water soluble polymers such as oligosaccharides and PEG, etc.

Column list

●CQP series

MCI GEL™ column	USP	Column dimensions	Packing materials		Theoretical plates number [TP/column]	Exclusion limit [PEG]
			Particle size [μm]	Pore size [nm]		
MCI GEL™ CQP06	L25	7.5mm I.D. ×600mm	10	12	10000	~1×10 ³
MCI GEL™ CQP10	L38	7.5mm I.D. ×600mm	10	20	6000	~1×10 ⁴
MCI GEL™ CQP30	L37, L38	7.5mm I.D. ×600mm	10	60	6000	~1×10 ⁶

●Guard columns

MCI GEL™ column	Column dimensions
MCI GEL™ CQP06G	4.0mm I.D. ×50mm
MCI GEL™ CQP10G	4.0mm I.D. ×50mm
MCI GEL™ CQP30G	4.0mm I.D. ×50mm

Application data of CQP series

Fig. 4-4 Calibration curve

Conditions
Column : MCI GEL™ CQP06
Eluent : H₂O
Flow rate : 1.0 ml/min
Column temp. : 25°C
Detection : RI
Sample : PEG 100μl inj.

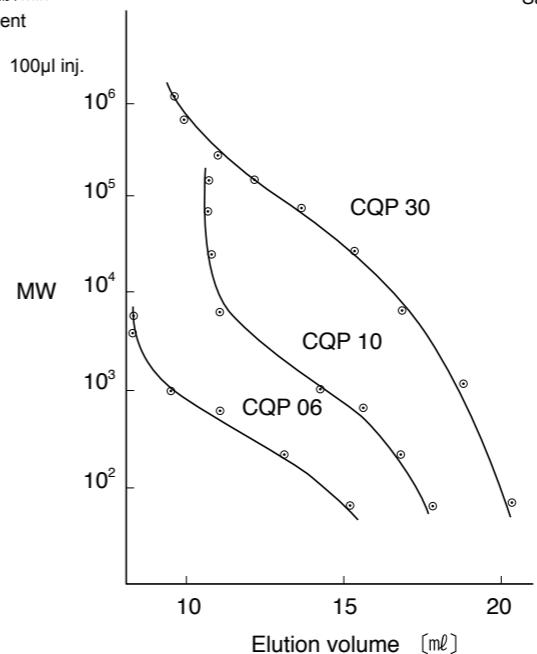


Fig. 4-5 Separation of PEG mixture

Conditions
Column : MCI GEL™ CQP30 7.5mm I.D. ×600mm
Eluent : H₂O
Flow rate : 1.0 ml/min
Column temp. : 25°C
Detection : RI
Sample : 1. PEG 145,000
2. 40,000
3. 6,000

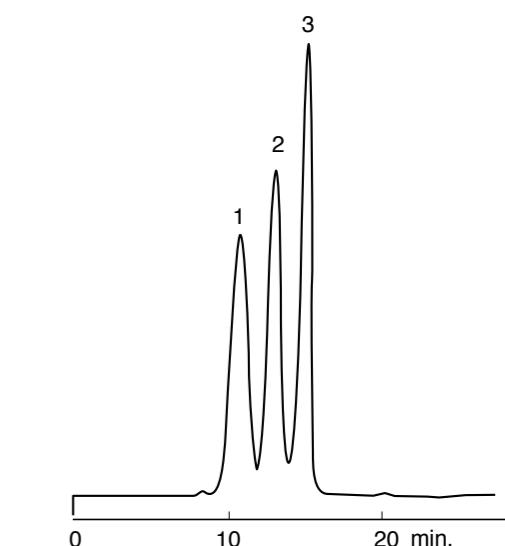


Fig. 4-6 Separation of protein mixture

Conditions
Column : MCI GEL™ CQP30 7.5mm I.D. ×600mm
Eluent : 14mM Tris-HCl/O₄ buffer
Flow rate : 1.0 ml/min
Column temp. : ambient
Detection : 280nm
Sample : 1. Ferritin (MW440,000)
2. Ovalbumin (MW43,000)
3. Myoglobin (MW17,500)
4. Cytochrome c (MW12,400)

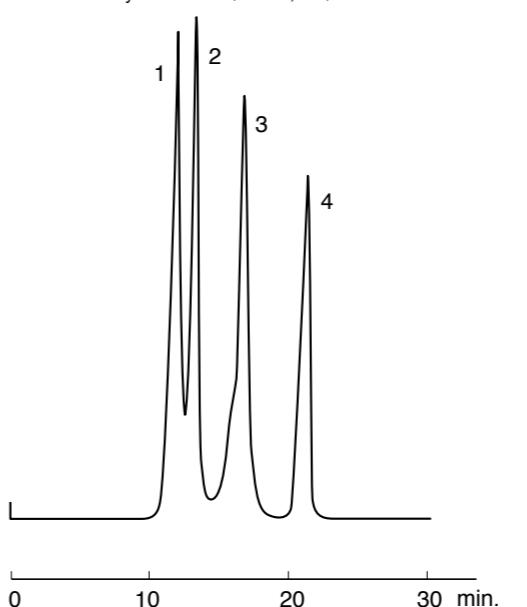
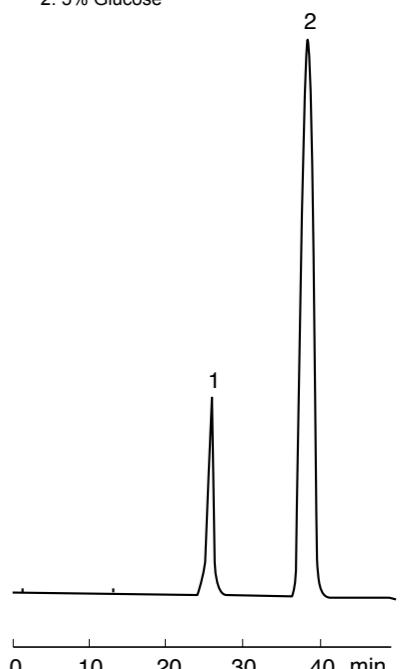


Fig. 4-7 Separation of gluconic acid and glucose

Conditions
Column : MCI GEL™ CQP06 7.5mm I.D. ×600mm
Eluent : H₂O
Flow rate : 0.8 ml/min
Column temp. : ambient
Detection : RI
Sample : 1. 5% Gluconic acid
2. 5% Glucose



5**MCI GEL™****Analytical and preparative chromatography columns
and materials for pharmaceutical applications****○Polymeric partition chromatography columns and materials
MCI GEL™ CHP series****Separation mechanism of CHP series**

High performance liquid chromatography relies on one of the following physical phenomena for efficient separation of solutes: partition, adsorption, size exclusion, or ion exchange. Of these, partition chromatography is the most commonly used method, and it separates solutes based on their difference in partitioning between a stationary phase and a mobile phase. This technique has currently become the mainstay in industry for the separation of organic compounds such as pharmaceuticals, agricultural chemicals, and other intermediates. Practically, partition chromatography can be performed in two different modes depending on the relative polarities of the stationary and mobile phases. In the normal phase (NP) mode, the mobile phase is less polar than the stationary phase while the situation is reversed in the reverse phase (RP) mode, where the mobile phase is significantly more polar than the stationary phase.

MCI GEL™ specializes in polymer-based packing materials. The use of polymer-based columns has become more widespread thanks to the many advantages of the polymer matrix like excellent selectivity, the absence of specific adsorption which is found commonly with silica-based packing, operability in a wide pH range and good chemical stability due to the inert nature of polymeric materials. The MCI GEL™ partition chromatography columns are based on a polystyrene and polymethacrylate porous polymer. As RP columns, they are applied to the separation of a wide variety of organic compounds, both in the isocratic and gradient elution mode. The compounds include peptides, insulin, small molecule APIs, nutraceutical compounds, water-soluble vitamins and nucleotides. As NP columns, they are used in the separation of various carotenoids, fat-soluble vitamins, steroids, and food additives. These columns tolerate various organic solvents like hexane, heptane, methylene chloride, and alcohols.

As NP columns, they are used in the separation of various carotenoids, fat-soluble vitamins, steroids, and food additives. Various organic solvents like Hexane Heptane, methylene chloride and alcohols can be used.

The MCI GEL™ packing materials are based on the same chemistries offered in the Diaion™ and Sepabeads™ synthetic adsorbent resins. These polymer chemistries, like Diaion™ HP series and Sepabeads™ SP series, are widely used and documented in the biopharmaceutical industry for fermentation extraction, the food industry and in industrial chromatographic separations. The MCI GEL™ packing materials are available as packed columns for analytical applications, and as bulk packing materials for analytical, preparative and production chromatography applications.

●Description of MCI GEL™ columns and materials**MCI GEL™ CHP20/C04**

Matrix type _____

Particle size _____

{ C=Column
P=Material }**5 MCI GEL™****CHP column series**Analytical and preparative chromatography columns and
materials for pharmaceutical applications

MCI GEL™ CHP series are suitable for RP and NP chromatography. There are four kinds of columns of various hydrophobicities; porous polystyrene, modified porous polystyrene, polymethacrylate, and modified porous polymethacrylate. This range of packing materials offers tremendous scope for a proper selection of columns based on the properties of the target compounds.

Polystyrene packing: MCI GEL™ CHP20/C04, CHP20/C10

Modified polystyrene packing: MCI GEL™ CHP07/C04, CHP07/C10, CHK40/C04

Polymethacrylate packing: MCI GEL™ CMG20/C10

Modified polymethacrylate packing: MCI GEL™ CHPOD/C04, CHK45/C05

The hydrophobicities of the columns are in the following orders:

MCI GEL™ CHP07/C04, C10 > CHP20/C04, C10 > CHPOD/C04 ≥ ODS columns ≥ CMG20/C04, C10

Polymer columns for HPLC, with their superior chemical resistance, can be used with various mobile phases of broad pH range, acidic through alkaline. They have the following advantages due to their high hydrophobicities:

- 1) In reverse phase chromatographic methods to separate acidic or alkaline compounds, eluents that can suppress the ionic properties of such compounds are generally used. Polymer columns can be applied in these cases where ODS columns would be unsuitable.
- 2) Some extremely hydrophilic compounds, e.g., oligosaccharides, can be separated using strongly hydrophobic CHP07/C04 or CHP07/C10 columns.
- 3) Polymer columns can be washed with acidic and/or basic solutions in case of contamination.

Polymethacrylate columns, CMG20/C04 and CMG20/C10, can be applied both for reverse phase and normal phase chromatography.

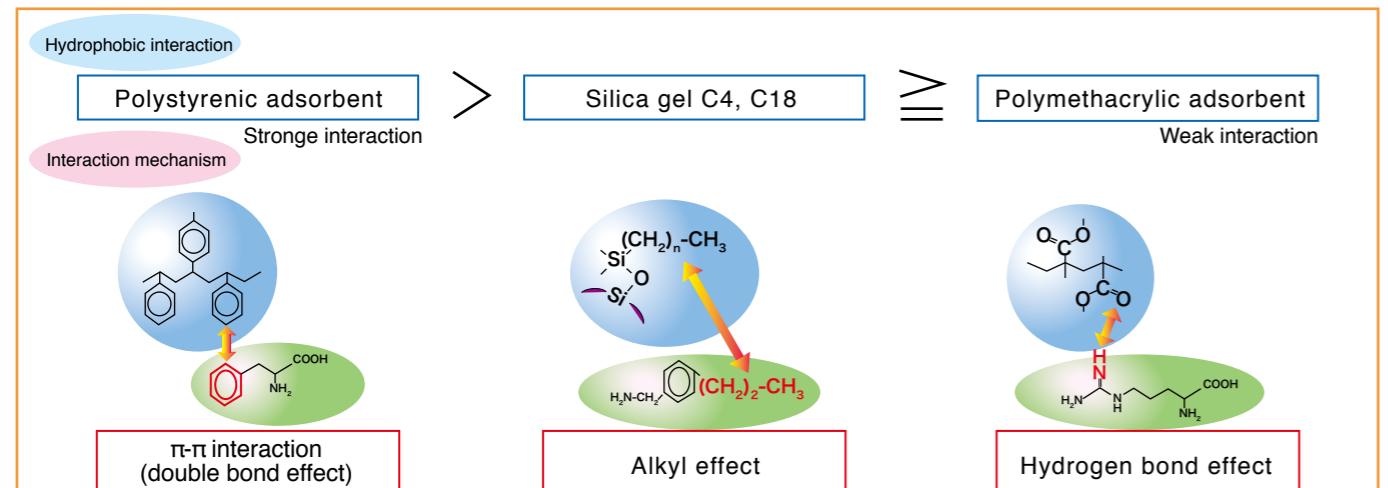
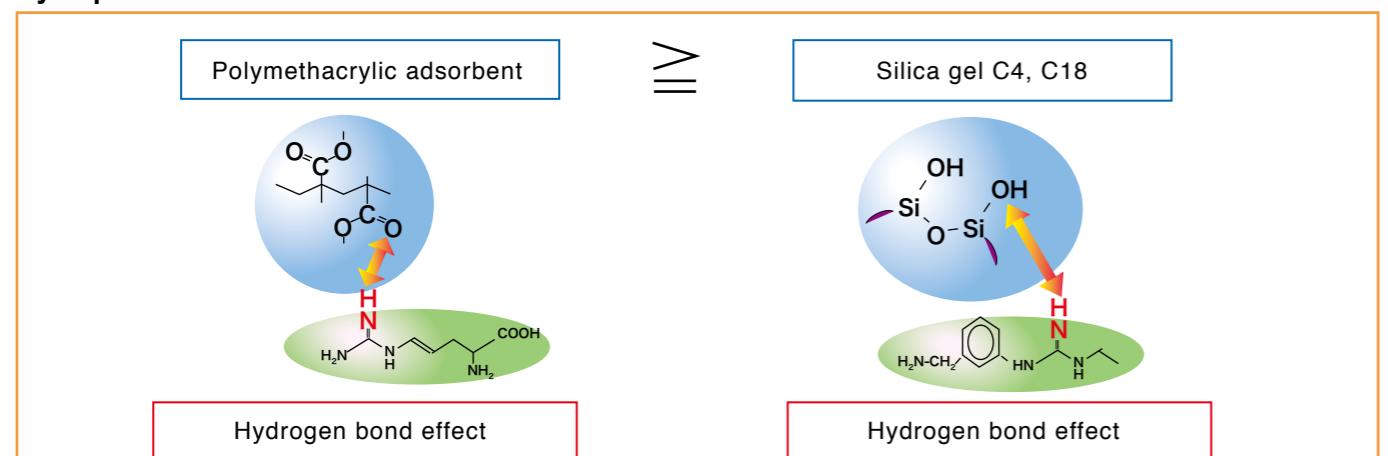
Modified polystyrene packing, CHK40/C04, is a mixed-mode type material; both hydrophobic and hydrophilic interactions occur between the packing material surface and the analytes. This material is useful for compounds that are difficult to separate using existing ODS or other polymer-based columns. This column is also used in the normal phase mode and shows a unique separation profile.

All polymeric columns exhibit superior stability and yield in comparison to ODS columns, which may have free silanol groups even when end-capping agents have been used.

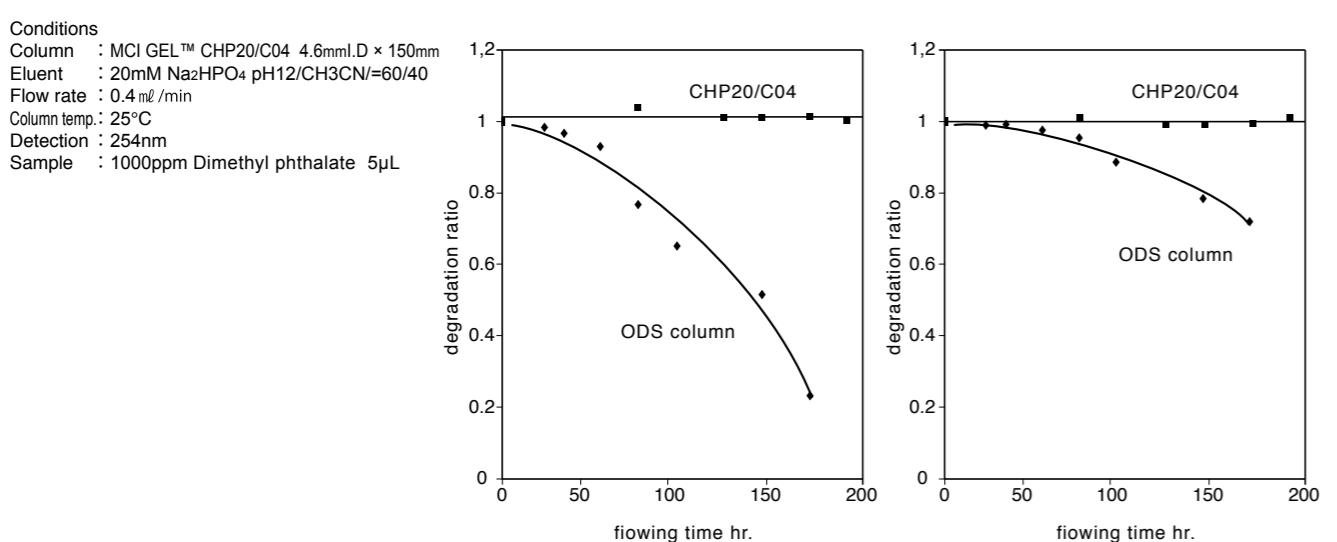
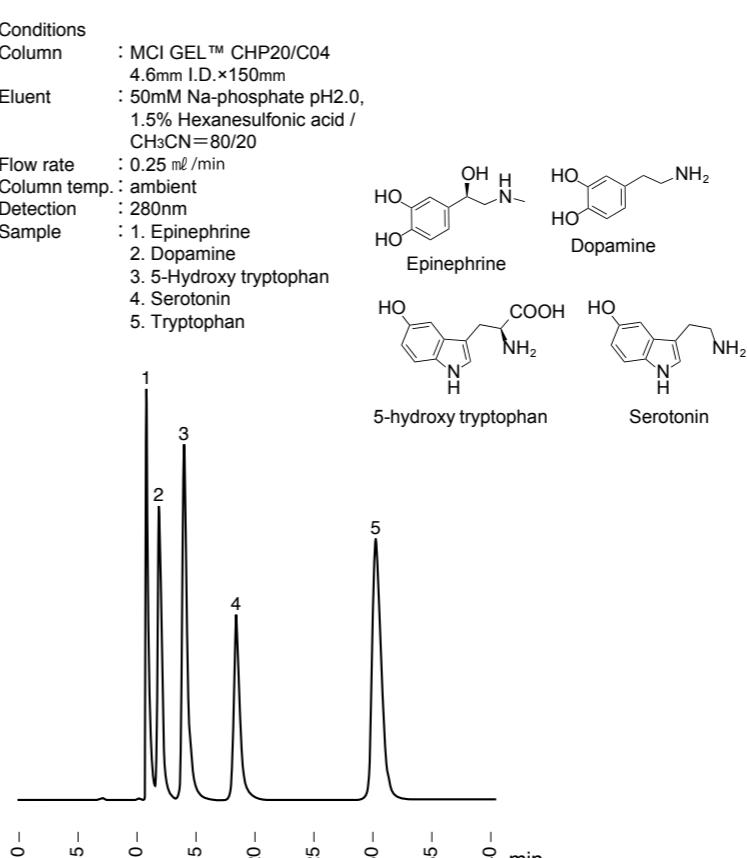
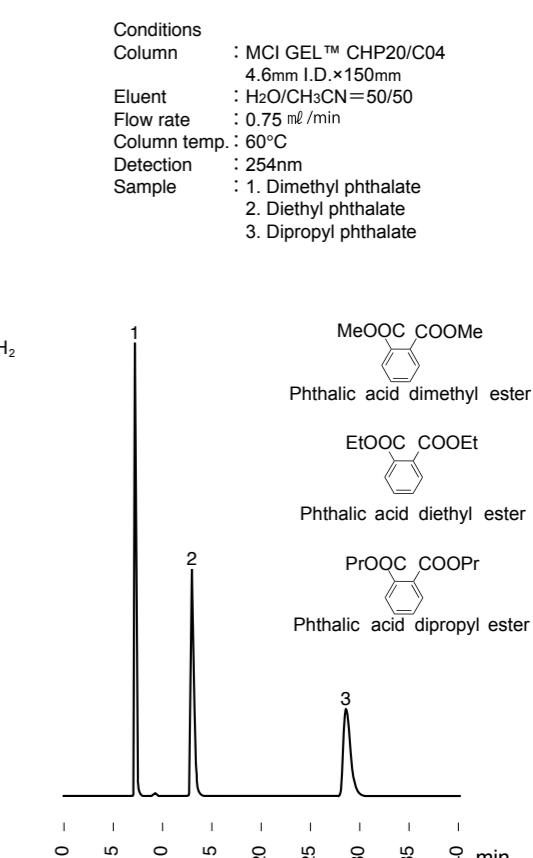
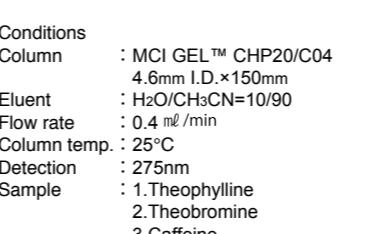
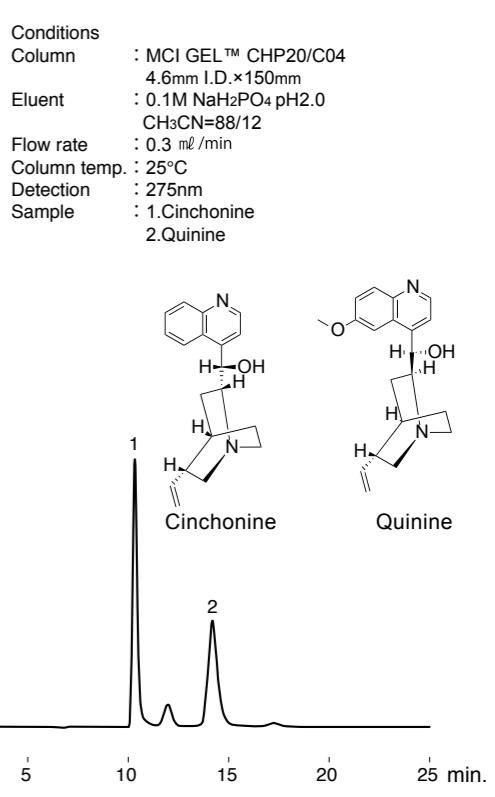
Column list**●CHP column series**

Matrix Type	Functional group	Product name	Particle size [μm]	Column size [mm I.D.×mm]	pH range	USP
Styrene Divinylbenzene	None	CHP20/C04	4	4.6×150 20×150	Full range	L21
		CHP20/C10	10	4.6×150 4.6×250 10×250 20×150 20×250		
	Br	CHP07/C04	4	4.6×150 20×200		
		CHP07/C10	10	4.6×150 4.6×250 10×150 20×150 20×250		
	Cation exchange group	CHK40/C04	4	4.6×150		
Methacrylates	None	CMG20/C04	4	4.6×150 20×150	2~12	
		CMG20/C10	10	4.6×150 4.6×250 10×250 20×150 20×250		
	C18	CHPOD/C04	4	4.6×150 20×200		
	Weak cation exchange group	CHK45/C05	5	4.6×150		

*CHP20/C04, CHP20/C10: USP classification is L21

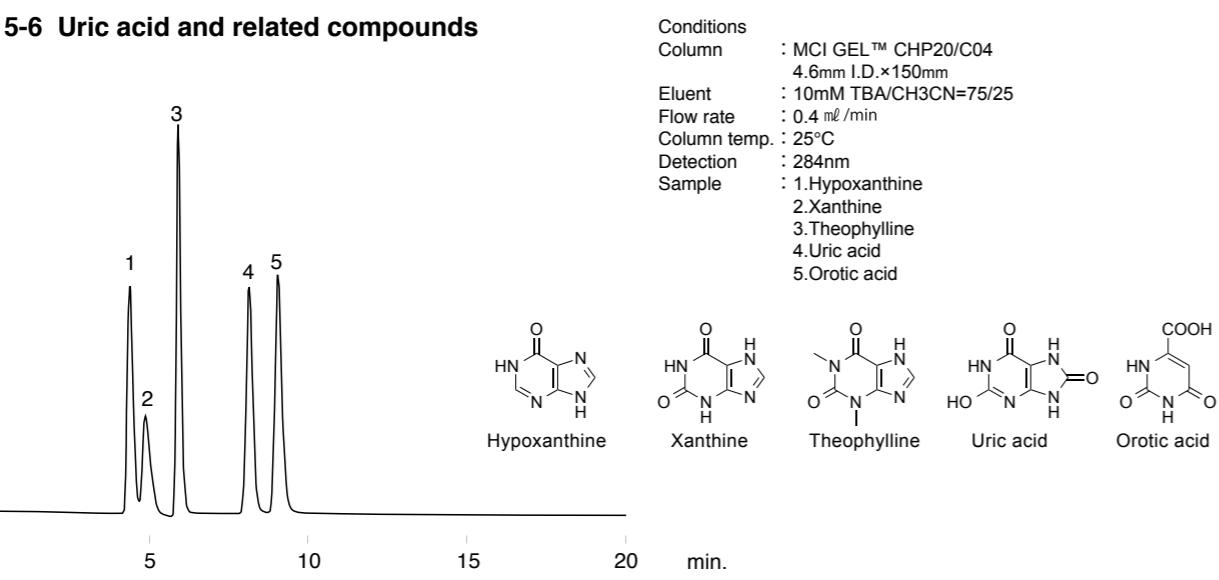
Retentiveness in reverse phase mode**Hydrophobic interaction Interaction mechanism****Durability of polymeric column**

The polymeric RP columns are chemically stable. Specifically, the columns have resistance to an alkaline eluent. The following graphs demonstrate stability of the polymeric columns. After feeding a solution of pH 12 into the MCI GEL™ CHP20/C04, there is no change of column performance.

Fig. 5-1 Column durability at pH12 comparison between CHP20/C04 and an ODS column**Application data of CHP series****Fig. 5-2 Separation of catecholamines****Fig. 5-3 Separation of phthalic acid esters****Fig. 5-4 Purine alkaloids****Fig. 5-5 Cinchona alkaloids**

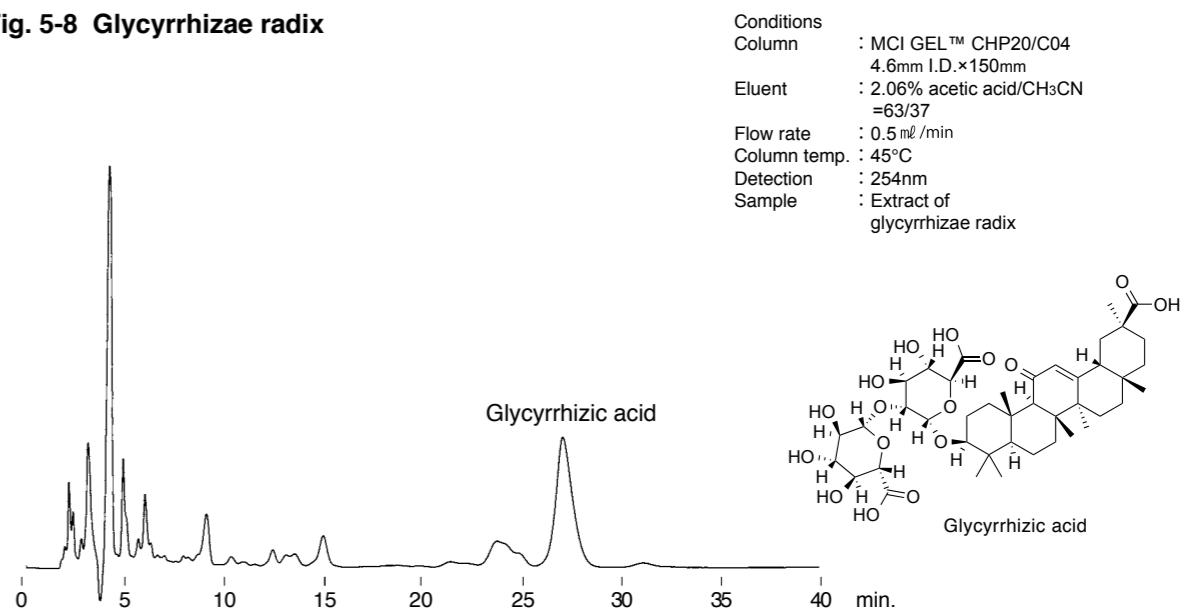
Application data of CHP series

Fig. 5-6 Uric acid and related compounds



Application data of CHP series

Fig. 5-8 Glycyrrhizae radix



Comparison with an ODS column

Fig. 5-7 Bile acids

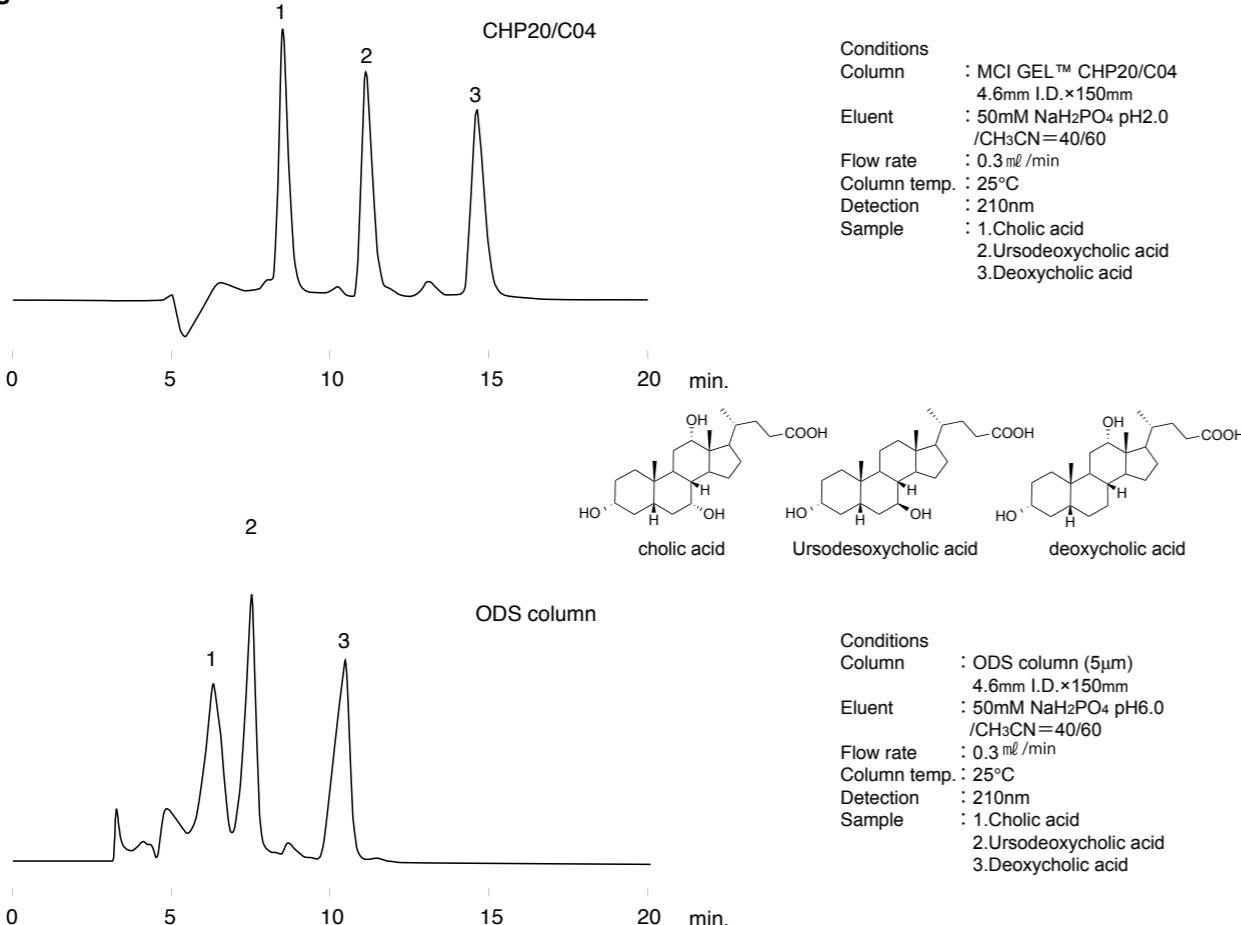
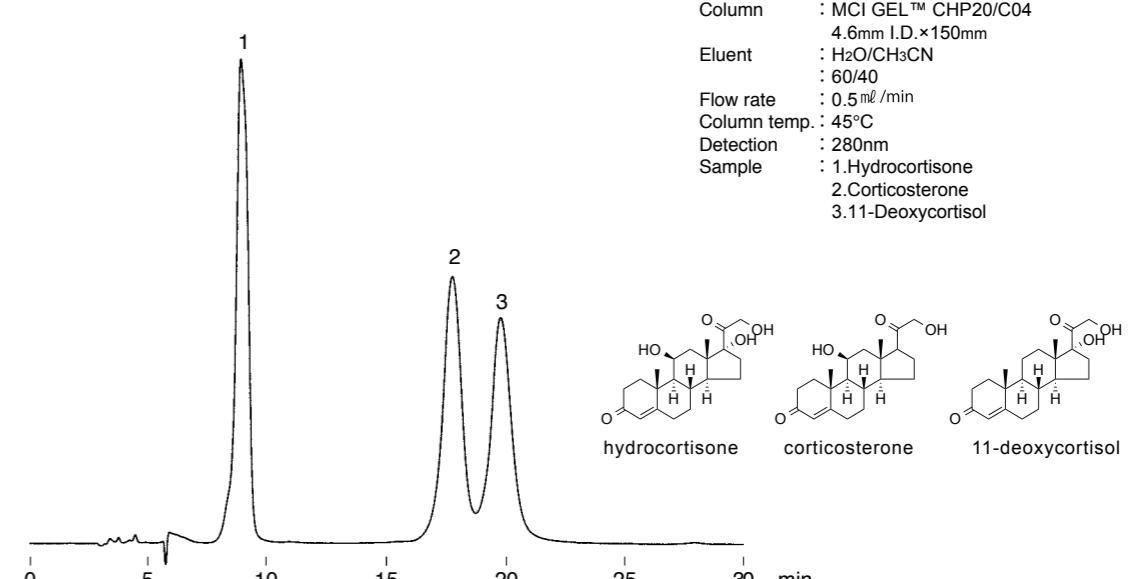
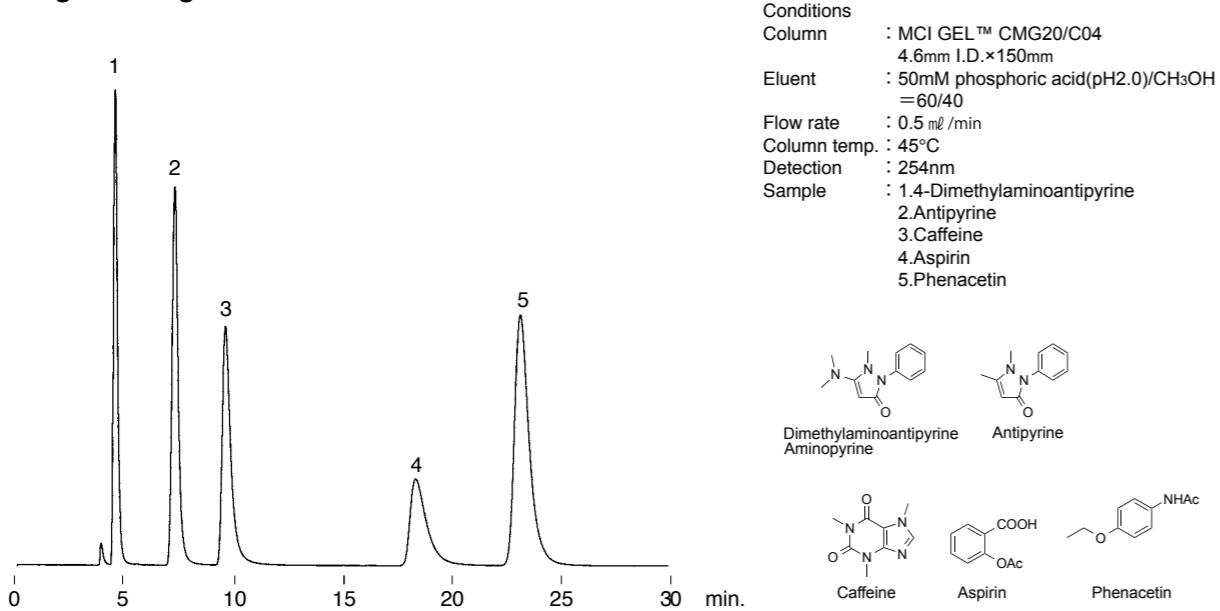


Fig. 5-9 Adrenal cortex hormones



Application data of CHP series

Fig. 5-10 Ingredients of medicine



Application data of CHP series

Fig. 5-12 Peptides

Conditions

- Column : MCI GEL™ CMG20/C04
4.6mm I.D.×150mm
- Eluent : 0.1%TFA/CH₃CN =70/30
- Flow rate : 0.5 mL/min
- Column temp. : 25°C
- Detection : 220nm
- Sample : 1.Gly-Tyr
2.Met Enkephalin
3.Leu Enkephalin
4.Angiotensin II

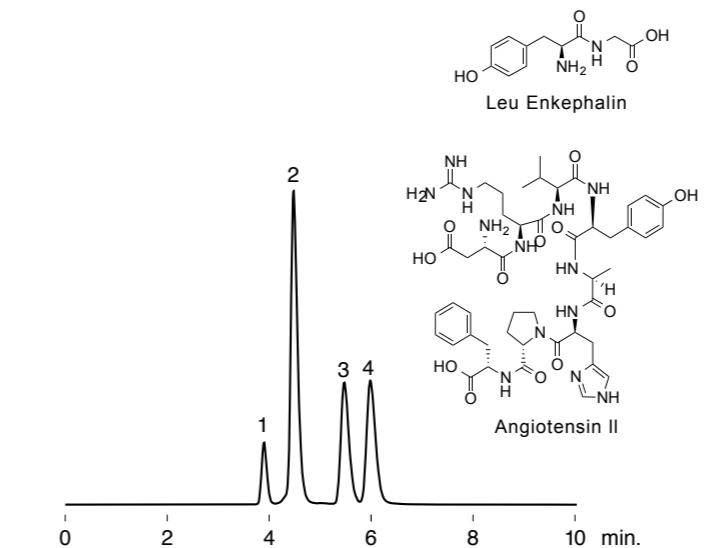
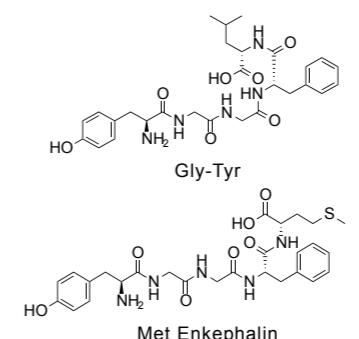
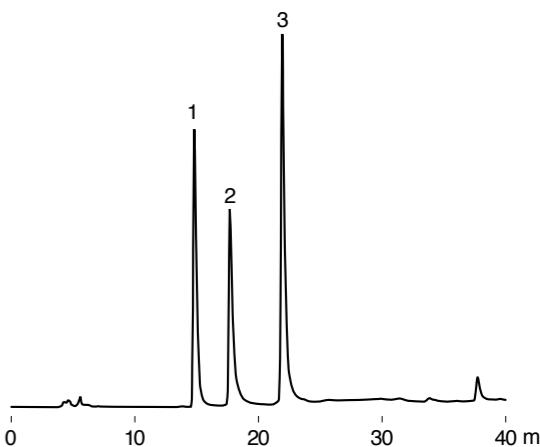


Fig. 5-13 Proteins

Conditions

- Column : MCI GEL™ CMG20/C04
4.6mm I.D.×150mm
- Eluent : A;0.05%TFA/CH₃CN =80/20
B;0.05%TFA/CH₃CN =20/80
A→B 30min.linear
- Flow rate : 0.5 mL/min
- Column temp. : 25°C
- Detection : 280nm
- Sample : 1.Ribonuclease A
2.Cytochrome c
3.α-Chymotrypsinogen A



Comparison with an ODS column

Fig. 5-11 Sulfa drugs

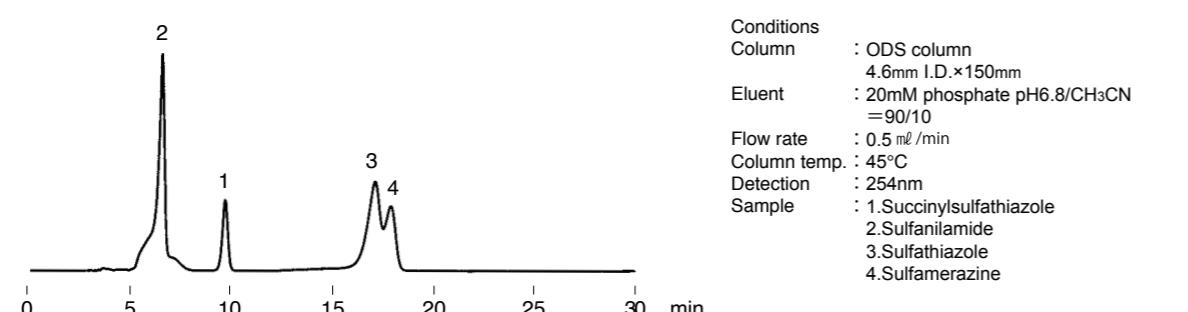
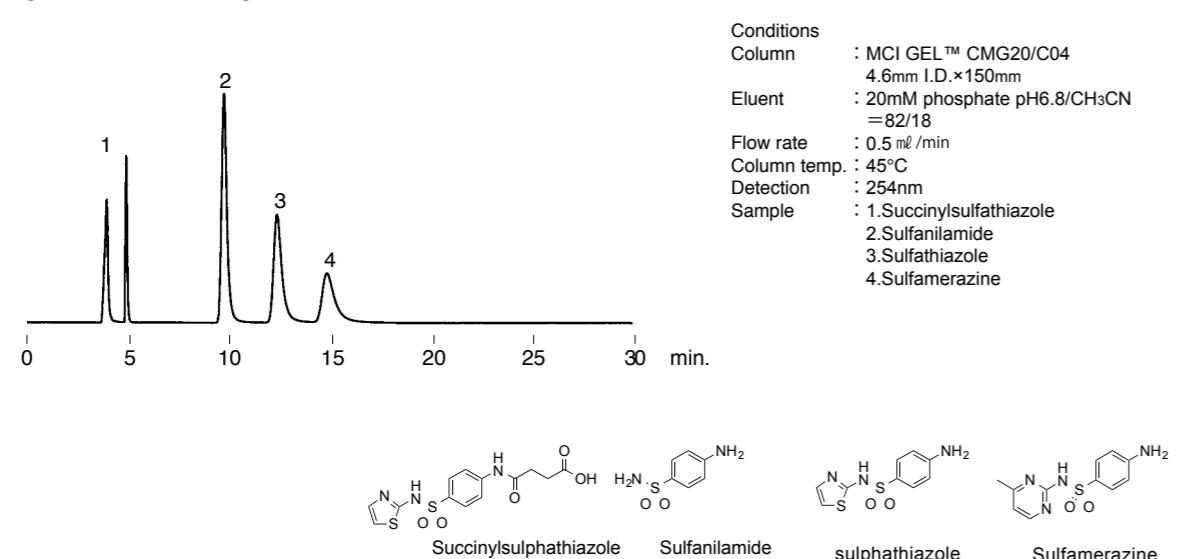


Fig. 5-14 Procainamide, Procaine

Conditions

- Column : MCI GEL™ CMG20/C04
4.6mm I.D.×150mm
- Eluent : 20mM phosphate pH7.2/CH₃CN =65/35
- Flow rate : 0.5 mL/min
- Column temp. : 45°C
- Detection : 254nm
- Sample : 1.Procainamide
2.Procaine

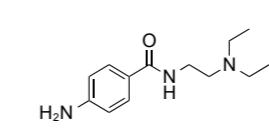
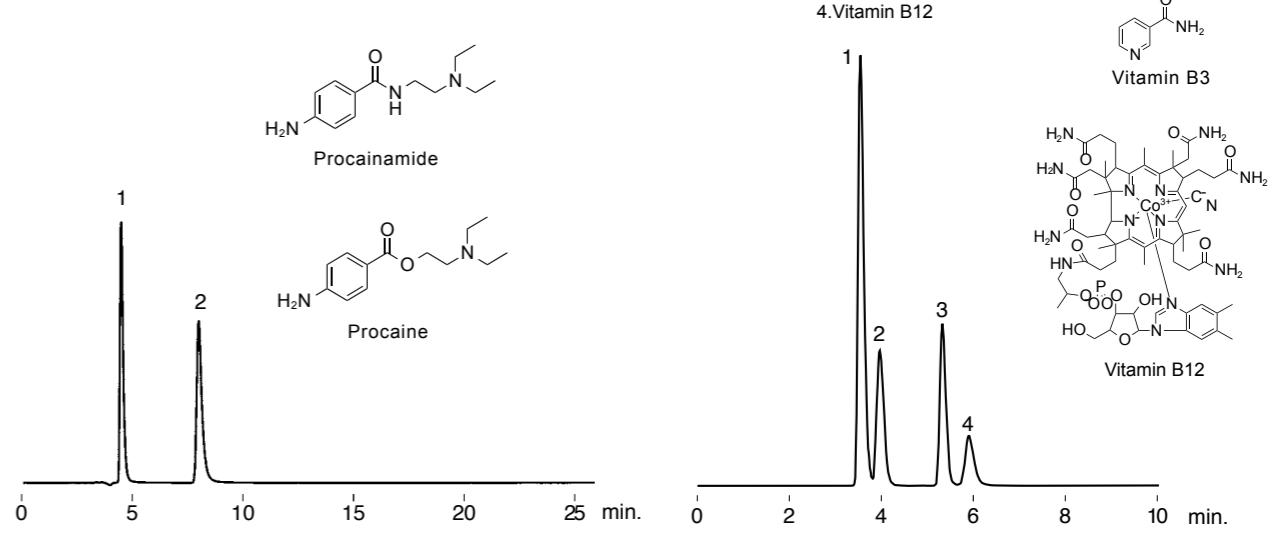


Fig. 5-15 Water-soluble vitamins

Conditions

- Column : MCI GEL™ CMG20/C04
4.6mm I.D.×150mm
- Eluent : 8mM Na₂HPO₄ pH7.0/CH₃CN =85/15
- Flow rate : 0.5 mL/min
- Column temp. : 25°C
- Detection : 254nm
- Sample : 1.Vitamin C
2.Vitamin B6
3.Vitamin B3
4.Vitamin B12



Application data of CHP series

Fig. 5-16 Pravastatin sodium

Conditions
Column : MCI GEL™ CHP20/C10 (10μm 250 ×4.6mm I.D.) and ODS (10μm 250 ×4.6mm I.D.)
Eluent : A :0.1% Formic acid;
B :0.1% Formic acid in AcCN;
Gradient : 45%B-95%B over 29min.
Flow rate : 1.00 ml/min
Column temp.: 25°C
Detection : UV238nm
Sample : Pravastatin sodium, Mevastatin and Simvastatin, 1mg/ml each;
Injection : 5μl

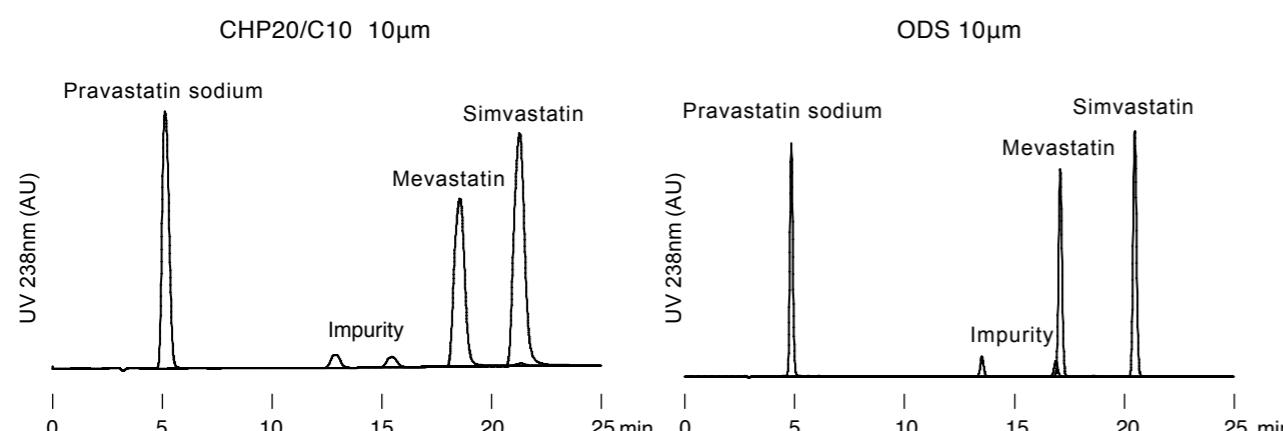
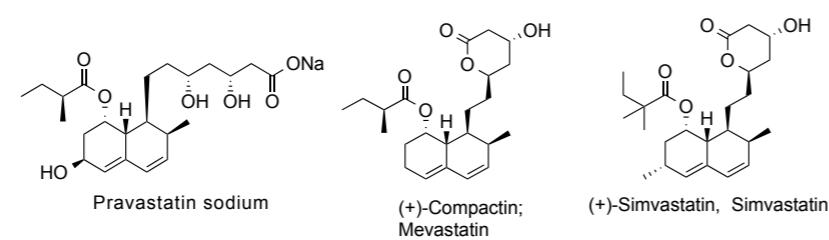
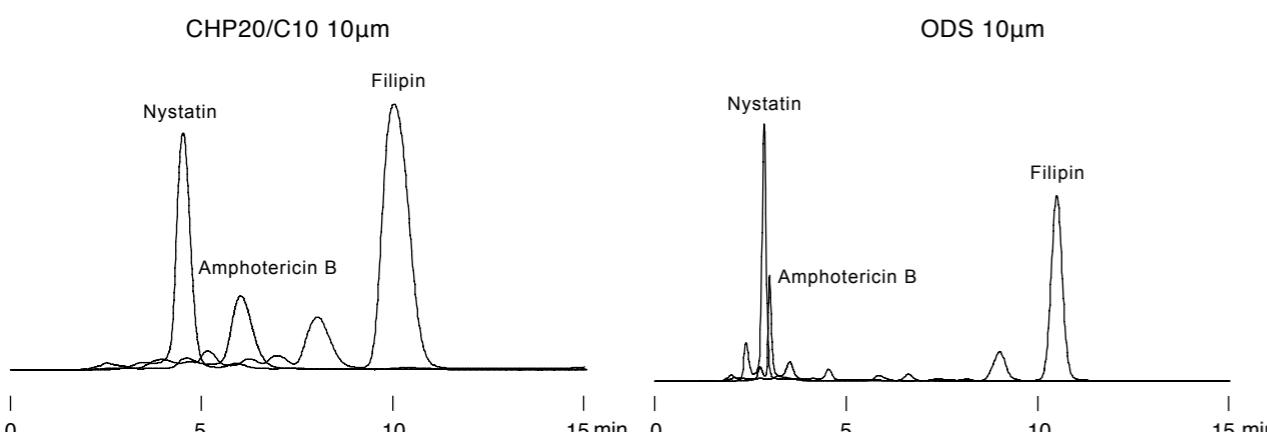
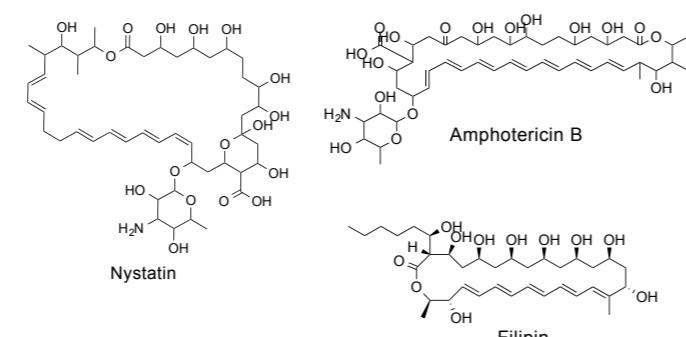


Fig. 5-17 Polyene antibiotics

Conditions
Column : MCI GEL™ CHP20/C10 (10μm 250 ×4.6mm I.D.) and ODS (10μm 250 ×4.6mm I.D.)
Eluent : A :0.1% Formic acid;
B :0.1% Formic acid in AcCN; A/B=60/40;
Flow rate : 1.00ml/min
Column temp.: 25°C
Detection : UV305nm for Nystatin, VIS405nm for Amphotericin B and UV340nm for Filipin;
Sample : Pravastatin sodium, Mevastatin and Simvastatin, 1mg/ml each.;
Injection : 10μl



Application data of CHP series

Fig. 5-18 Proteins

Conditions
Column : MCI GEL™ CMG20/C10 4.6mm I.D.×250mm
Eluent : A 0.05% TFA/CH₃CN=80/20
B 0.05% TFA/CH₃CN=30/70
A → B 45min linear gradient
Flow rate : 0.5 ml/min
Column temp.: 25°C
Detection : 280nm
Sample : 1. Ribonuclease A
2. Cytochrome C
3. Transferrin
4. α-Chymotrypsinogen A
5. β-Lactoglobulin

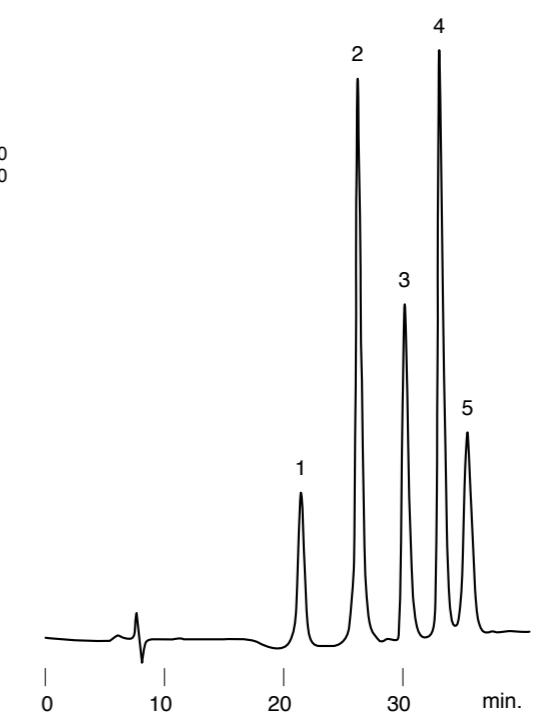
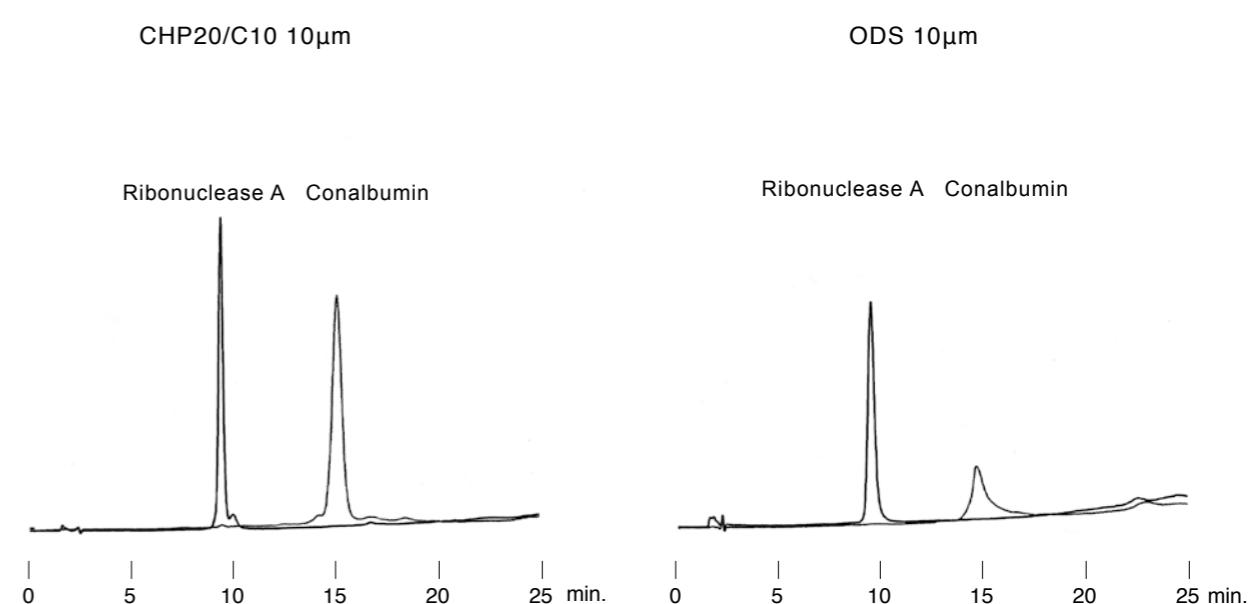


Fig. 5-19 Proteins

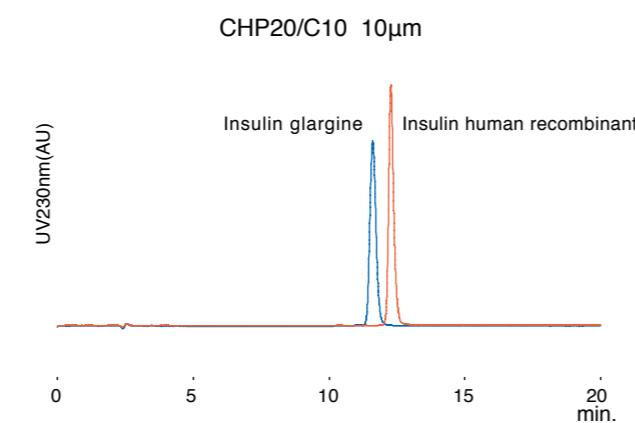


Conditions
Column : 150 ×4.6mm I.D.
Eluent : A :0.1% TFA;
B :0.1% TFA in AcCN
Flow rate : 1.00 ml/min
Column temp.: 20%B-60%B over 20min;
Detection : UV280nm;
Sample : Ribonuclease A and Conalbumin 2mg/ml.;
Injection : 10μl

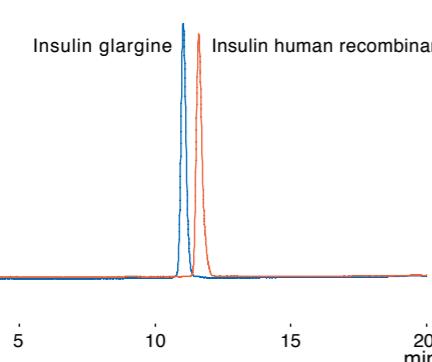
Application data of CHP series

Fig. 5-20 Insulin

Conditions
Column : MCI GEL™ CHP20/C10
MCI GEL™ CMG20/C10
ODS 10 μ m
4.6mm I.D. \times 150mm
Eluent : A) 0.1%TFA, H₂O
B) 0.1%TFA, CH₃OH
Gradient : 20% B \rightarrow 60% B over 20min.
Flow rate : 1.0 mL/min
Column temp. : 40°C
Detection : 280nm
Sample : Insulin Glargine and human recombinant, 1mg/ml each
Injection : 10 μ l



CMG20/C10 10 μ m



ODS 10 μ m

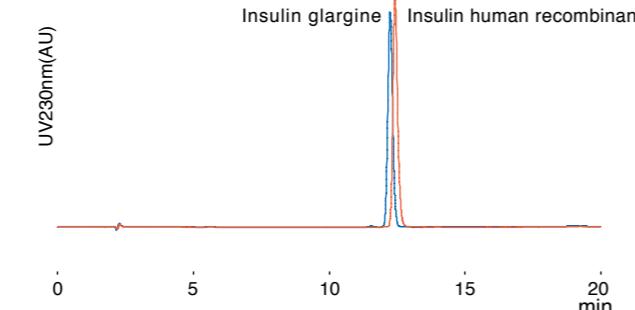
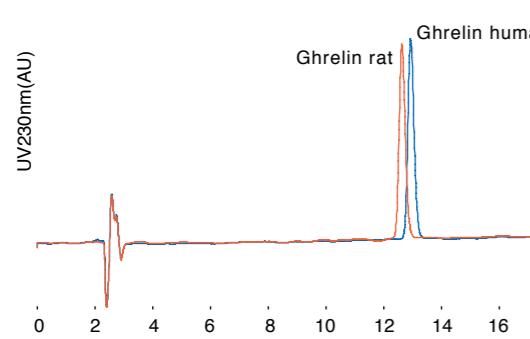


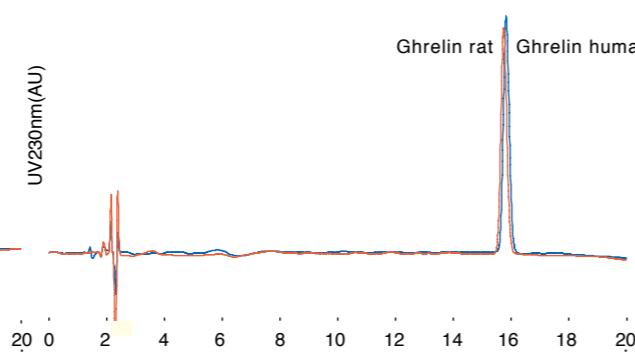
Fig. 5-21 Ghrelin

Conditions
Column : MCI GEL™ CMG20/C10
ODS 10 μ m
4.6mm I.D. \times 150mm
Eluent : A) 0.1%TFA, H₂O
B) 0.1%TFA, AcCN
Gradient : 10% B \rightarrow 60% B over 25min.
Flow rate : 1.0 mL/min
Column temp. : 40°C
Detection : 230nm
Sample : Ghrelin rat and Ghrelin human, 0.1mmol/l each
Injection : 10 μ l

CMG20/C10 10 μ m



ODS 10 μ m

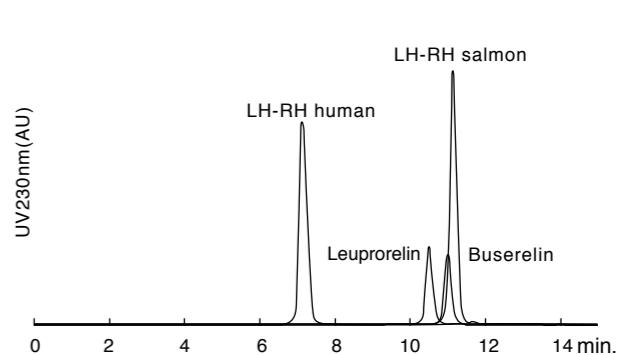


Application data of CHP series

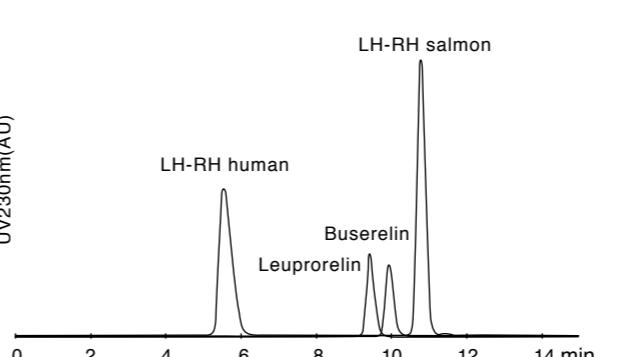
Fig. 5-22 Leuprorelin

Conditions
Column : MCI GEL™ CHP20/C10
MCI GEL™ CMG20/C10
ODS 10 μ m
4.6mm I.D. \times 150mm
Eluent : A) 0.1%TFA, H₂O
B) 0.1%TFA, AcCN
Gradient : 20% B \rightarrow 60% B over 20min.
Flow rate : 1.0 mL/min
Column temp. : 40°C
Detection : 280nm
Sample : Leuprorelin, LHRH human, LHRH salmon and Buserelin, 1mg/ml each
Injection : 10 μ l

CHP20/C10 10 μ m



CMG20/C10 10 μ m



ODS 10 μ m

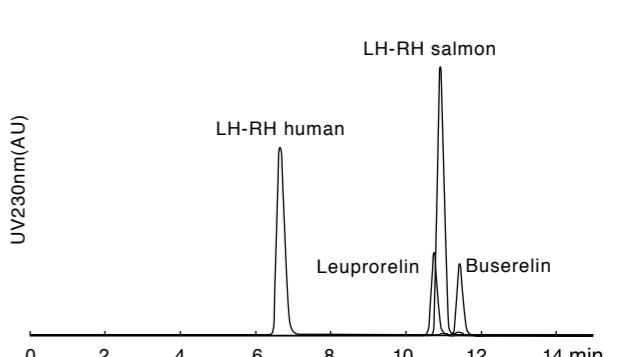
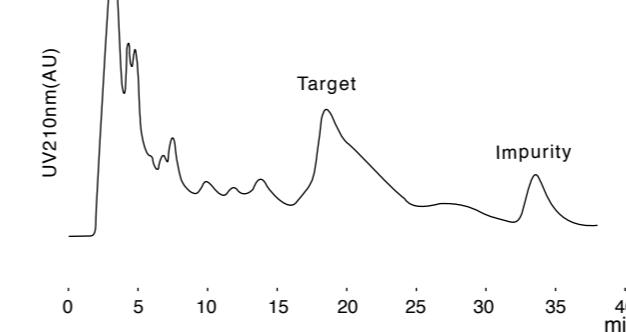


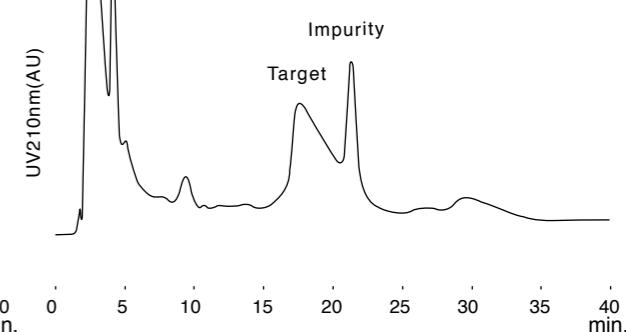
Fig. 5-23 Sifuvirtide

Conditions
Column : MCI GEL™ CMG20/C10
ODS 10 μ m
4.6mm I.D. \times 150mm
Eluent : 0.1%TFA/CH₃CN=68/32
Flow rate : 1.0 mL/min
Column temp. : 40°C
Detection : 210nm
Sample : Sifuvirtide crude(purity 35.5%) 2.1mg/ml
Injection : 0.4ml

CMG20/C10 10 μ m



ODS 10 μ m



Application data of CHP series

Fig. 5-24 ssRNA Ladder Marker

Conditions
Column : MCI GEL™ CMG20/C10
ODS 10μm
4.6mm I.D.×150mm
Eluent : A)100mM TEAA, H₂O
B)100mM TEAA, CH₃CN
Gradient : CHP10/C10 10%B→40%B over 30min
ODS 10μm 8%B→40%B over 30min
Flow rate : 1.0 ml/min
Column temp.: 40°C
Detection : 260nm
Sample : 14-30 ssRNA Ladder Marker [max.0.04mg/ml]
Injection : 5μl

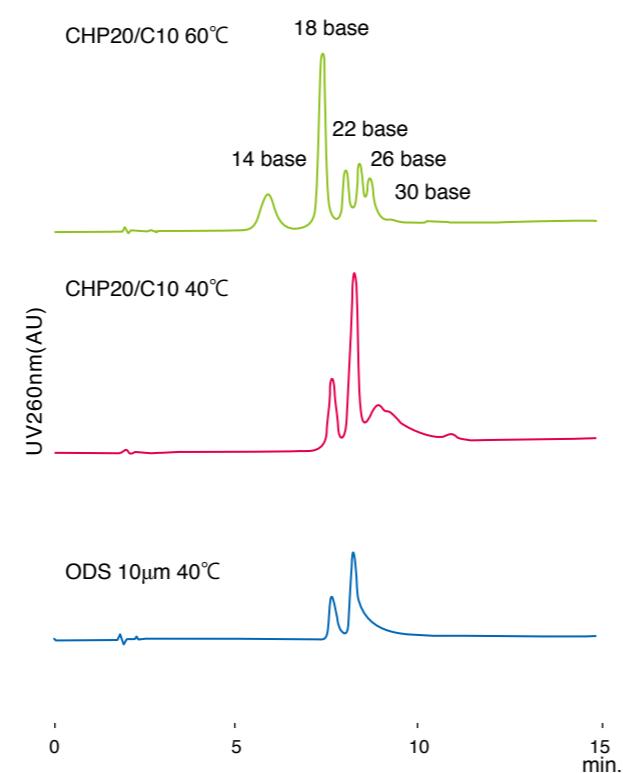
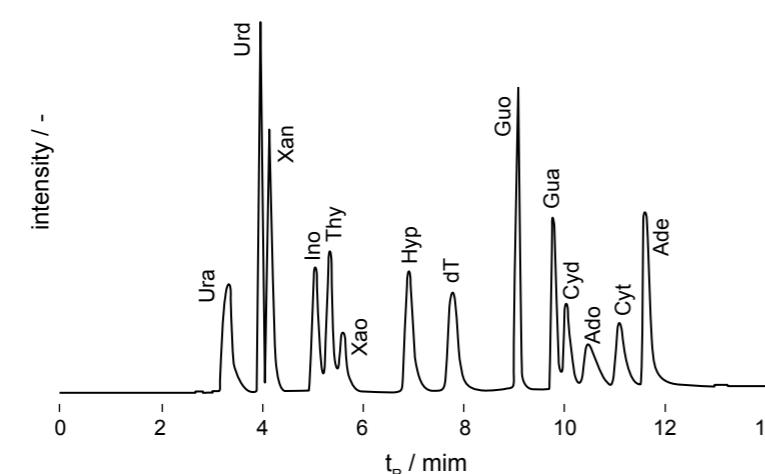


Fig. 5-25 Nucleotide

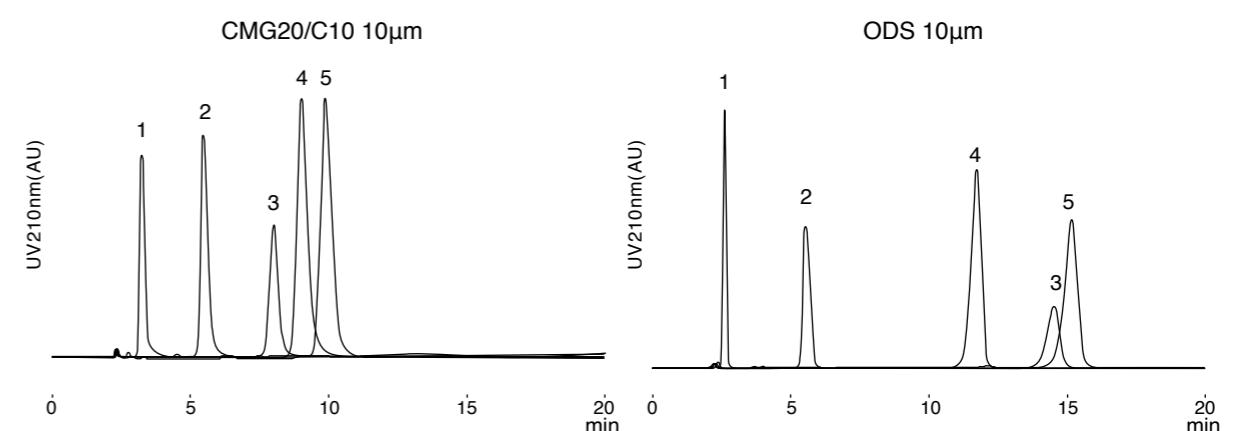
Conditions
Column : MCI GEL™ CHK40/C04
4.6mm I.D.×150mm
Eluent : A)19 mM H₃PO₄ / 1 mM NaH₂PO₄ / 5.0% ACN
B)20 mM Na₂HPO₄ / 100 mM NaClO₄ / 30% ACN
Gradient : 0-4.0min 0% B 4.0-5.0min 0→30% B 5.0min-6.0min 30% B 6.0min-7.0min 30→50% B
7.0min-10.0min 50→65% B 10.0min-11.0min 65% B 11.0min- 0% B
Flow rate : 0.8 ml/min
Column temp.: 50°C
Detection : UV260nm
Sample : 1.Ura, 2.Xan, 3.Thy, 4.Hyp, 5.Gua, 6.Cyt, 7.Ade, 8.Urd, 9.Xao, 10.dT, 11.Ino, 12.Guo, 13.Cyd, 14.Ado
Injection : 20μl



(Data provided by Professor Yokoyama of Yokohama National University)

Application data of CHP series

Fig. 5-26 Linalool

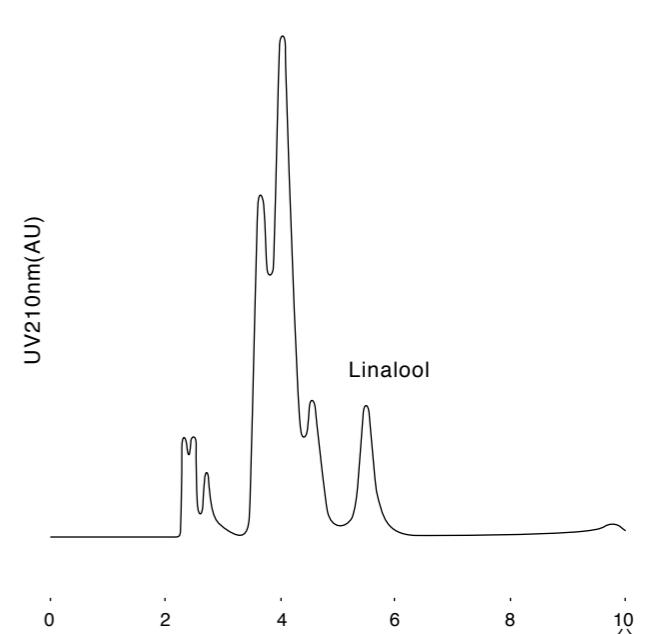


Conditions
Column : MCI GEL™ CMG20/C10
ODS 10μm
4.6mm I.D.×150mm
Eluent : Hexan/Ethanol=99.5/0.5
Flow rate : 1.0 ml/min
Column temp.: 40°C
Detection : 210nm
Sample : 1:Linalyl Acetate 1mg/ml
2:Linalool 1mg/ml
3:β-Citronellool 1mg/ml
4:Nerol 0.5mg/ml
5:Geraniol 0.5mg/ml
Injection : 10μl

Fig. 5-27 Coriander

Conditions
Column : MCI GEL™ CMG20/C10
4.6mm I.D.×150mm
Eluent : Hexan/Ethanol=99.5/0.5
Flow rate : 1.0 ml/min
Column temp.: 40°C
Detection : 210nm
Sample : Coriander
Injection : 10μl

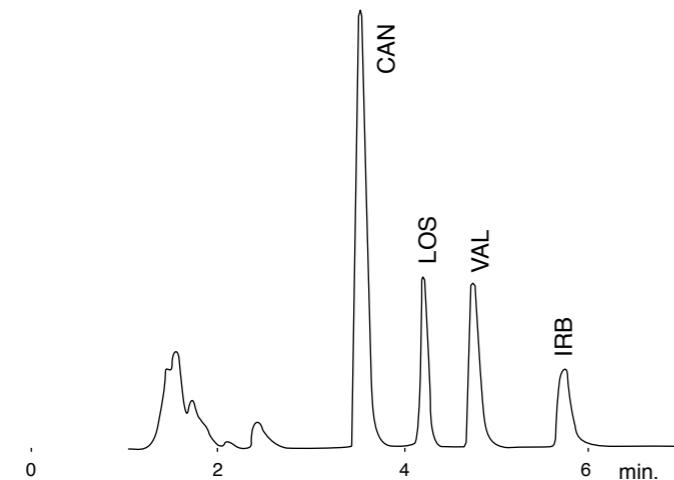
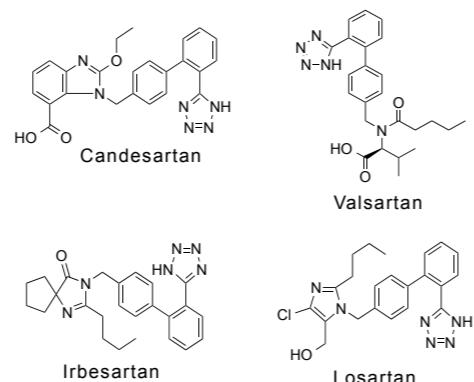
CMG20/C10 10μm



Application data of CHP series

Fig. 5-28 Application data of CHK40/C04: Separation of Sartans

Conditions
Column : MCI GEL™ CHK40/C04
4.6mm I.D.×150mm
Eluent : A) 10 mM NaH₂PO₄ + 0.2 mM Na₂HPO₄ (25%ACN)
B) 10 mM NaH₂PO₄ + 1.0 mM Na₂HPO₄ (40%ACN)
Gradient : 0.5min 0%B 0.5-2.0min 50%B
2.0min- 90%B
Flow rate : 1.0 mL/min
Column temp. : 50°C
Detection : UV
Sample : Candesartan(CAN), Losartan(LOS),
Valsartan(VAL), Irbesartan(IRB)
Injection : 20μL



(Data provided by Professor Yokoyama of Yokohama National University)

(Polyphenon 60)

Fig. 5-29 Modified Styrene Divinylbenzene CHP07/C04

Conditions
Column : MCI GEL™ CHP07/C04
4.6mm I.D.×150mm
Eluent : CH₃OH/10mM-Acetic acid=60/40
Flow rate : 0.46 mL/min
Column temp. : 60°C
Detection : 280nm
Sample : Polyphenon 60(10mg/mL) each 10μL

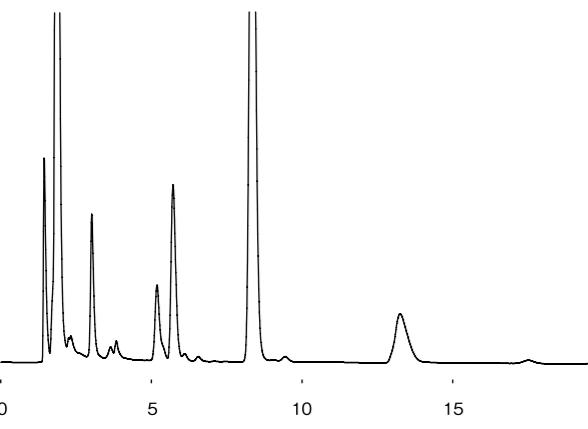
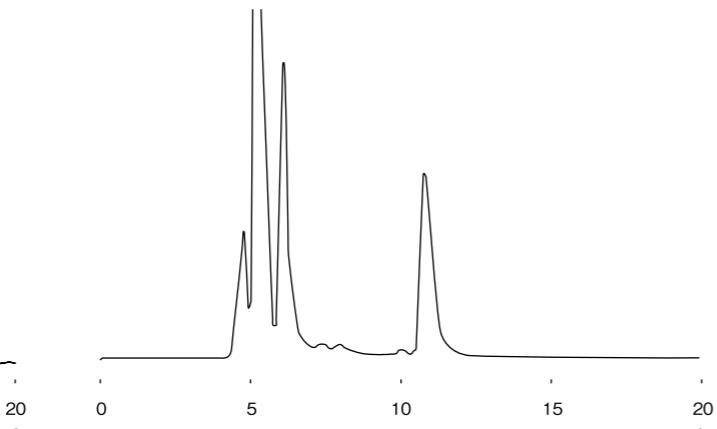


Fig. 5-30 Styrene Divinylbenzene CHP20/C04

Conditions
Column : MCI GEL™ CHP20/C04
4.6mm I.D.×150mm
Eluent : CH₃OH/10mM-Acetic acid=60/40
Flow rate : 0.46 mL/min
Column temp. : 60°C
Detection : 280nm
Sample : Polyphenon 60(10mg/mL) each 10μL



Application data of CHP series

(Triton X-100)

Fig. 5-31 C18-alkylated aliphatics CHPOD/C04

Conditions
Column : MCI GEL™ CHPOD/C04
4.6mm I.D.×150mm
Eluent : 50vol%CH₃CN
Flow rate : 0.50 mL/min
Column temp. : 40°C
Detection : 254nm
Sample : Triton X-100
(Polyoxyethylene octyl phenyl ether)
1% each 10μL

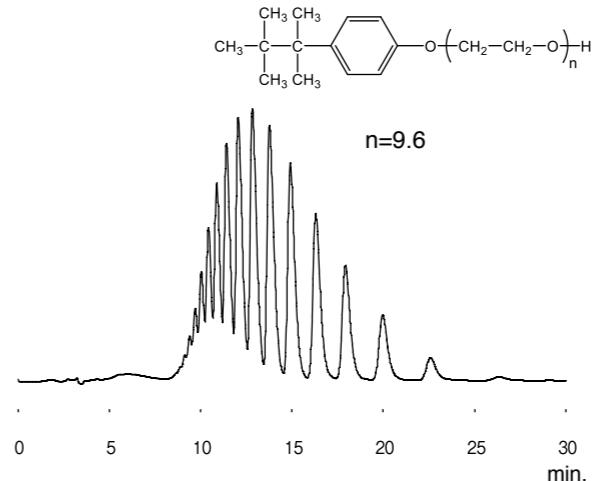


Fig. 5-32 ODS-1HU (ODS)

Conditions
Column : MCI GEL™ ODS-1HU
4.6mm I.D.×250mm
Eluent : 50vol%CH₃CN
Flow rate : 1.00 mL/min
Column temp. : 40°C
Detection : 254nm
Sample : Triton X-100
(Polyoxyethylene octyl phenyl ether)
1% each 10μL

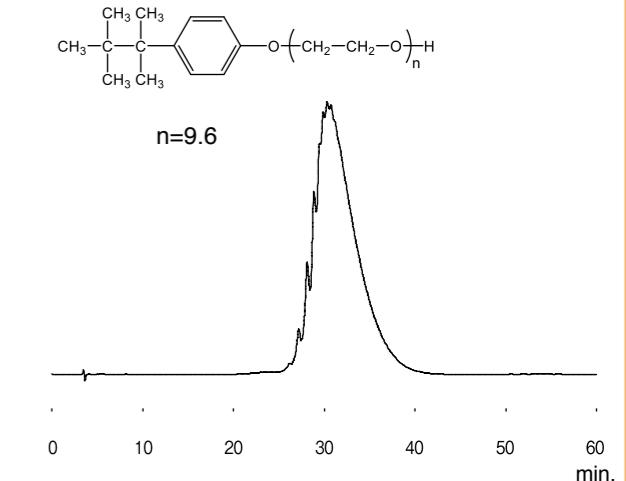
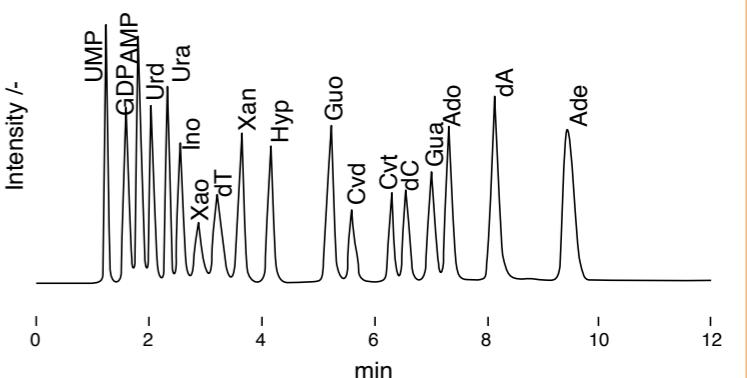
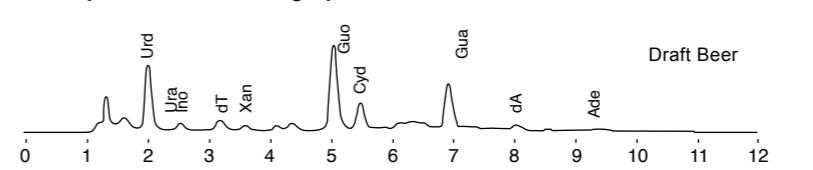


Fig. 5-33 Application data of Nucleic base/Nucleoside and Beer

Conditions
Column : MCI GEL™ CHK45/C05
4.6mm I.D.×150mm
Eluent : A) 8 mM H₃PO₄
B) 10 mM H₃PO₄/30% ACN
Gradient : 0-0.7min 0% B 0.7-3.0min 0→40% B 3.0-3.2min 40% B
3.2-3.5min 40→80% B 3.5-8.0min 80% B 8.0min- 0% B
Flow rate : 1.3 mL/min
Column temp. : 45°C
Detection : UV260nm
Injection : 20μL



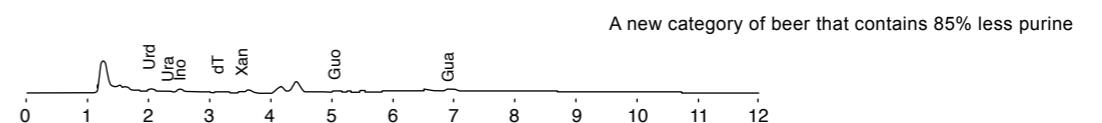
Analysis of various category beer



Draft Beer



Carbohydrate-free "happoshu-type" beer



A new category of beer that contains 85% less purine

(Data provided by Professor Yokoyama of Yokohama National University)

CHP material series

Analytical and preparative chromatography columns and materials for pharmaceutical applications

MCI GEL™ CHP material series are chromatography materials of porous type polymers.

Because polymeric materials are chemically stable, wide pH range, from acidic to alkaline eluents are able to be applied to MCI GEL™ CHP material series.

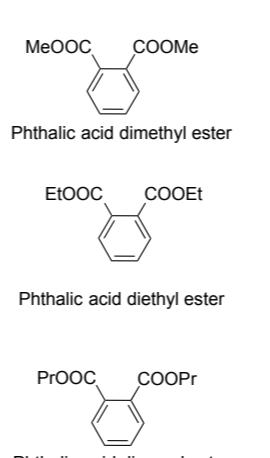
MCI GEL™ CHP50 series and CHP20 series are both ST/DVB polymers, but they differences in porosity. Pore size of CHP20 series is fairly larger than that of CHP50 series. Appropriate packing material can be selected in accordance with molecular size of injection samples.

● CHP material series

Base polymer	Functional group	Product name	Particle size [μm]	Pore diameter [nm]	Main application	Equivalent HPLC column	
Styrene Divinylbenzene	None	CHP20/P20	20	45	drug compounds	CHP20/C04	
		CHP20/P30	30			CHP20/C10	
		CHP20/P50	50		Peptides	—	
		CHP20/P70	70			CHP20/C10	
		CHP20/P120	120			CHP07/C04	
	Br	CHP50/P20	20	25	Proteins	CHP07/C10	
		CHP50/P30	30			CHP07/C10	
		CSP50/P10	10			CMG20/C04	
		CHP07/P120	120	25		CMG20/C10	
		CHP07/P120	120			CMG20/C10	
Polymethacrylate	None	CMG20/P10	10	25			
		CMG20/P30	30				
		CMG20/P150	150				

Fig. 5-34 Phthalic acid esters

Conditions
Column : MCI GEL™ CHP50/P20, 10mm I.D.×250mmL
Eluent : H₂O/CH₃CN=20/80
Flow rate : 0.75 mL/min
Column temp. : 25°C
Detection : 254nm,
Sample : 1.Dimethyl phthalate 0.5%
2.Dipropyl phthalate 0.5%
3.Dibutyl phthalate 0.5%
Injection : 100μL



Application data of CHP 50

Fig. 5-35 Penicillin antibiotics

Conditions
Column : MCI GEL™ CHP series, 10mm I.D.×250mmL
Eluent : CH₃OH/0.05M Phosphate buffer (pH8.0)=60/40
Flow rate : 2.18 mL/min
Column temp. : 25°C
Detection : 254nm,
Sample : 1.6-Aminopenicillanic acid 1000ppm
2.Penicillin G 1000ppm
3.Penicillin V 1000ppm
Injection : 100μL

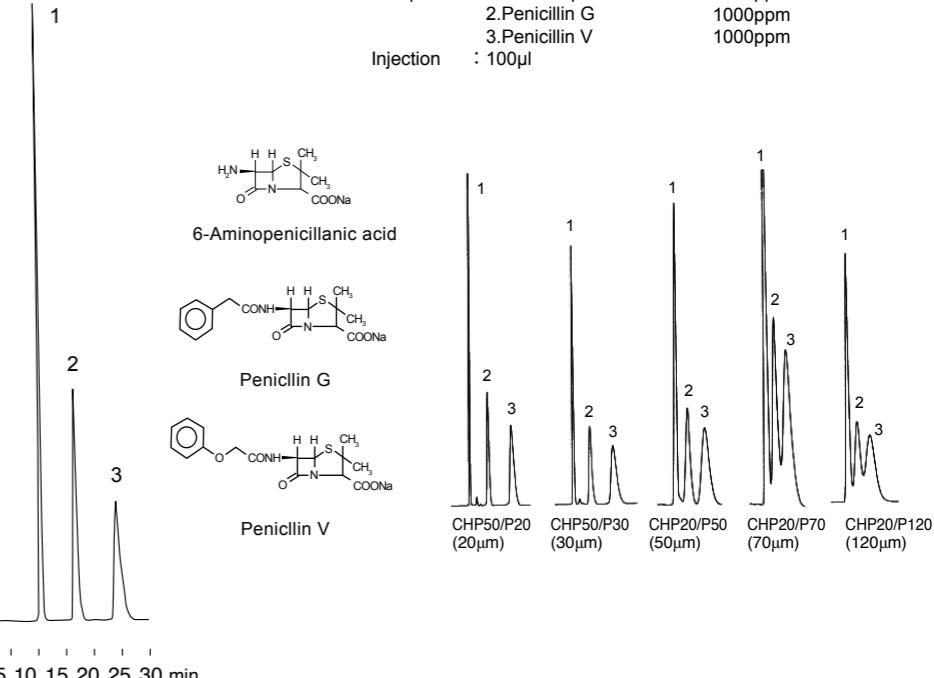
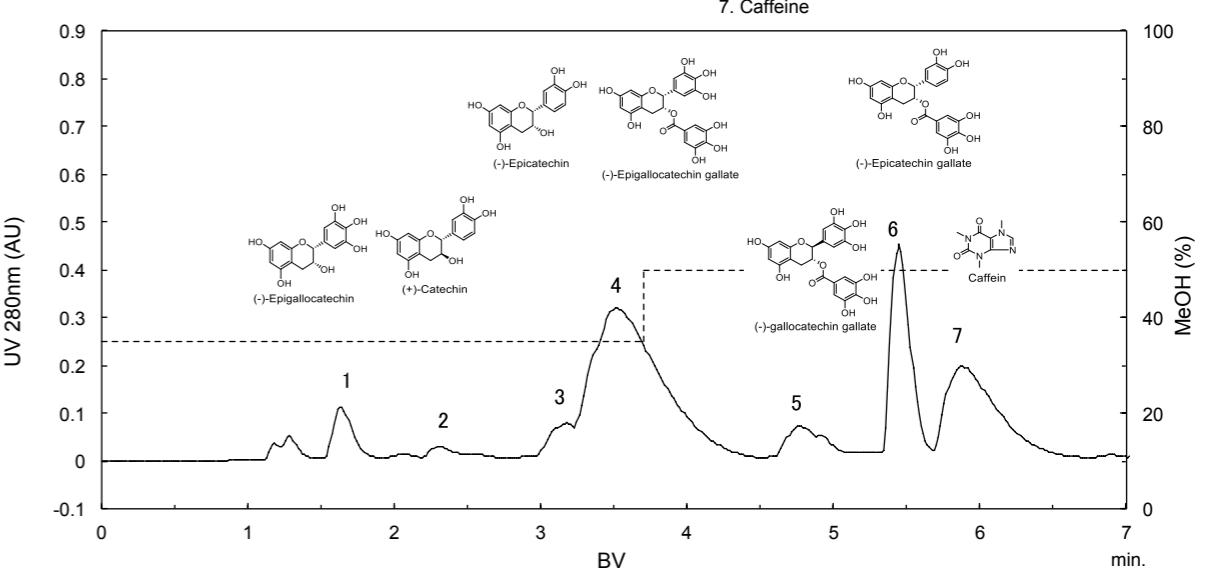


Fig. 5-36 Extract of green tea leaves

Conditions
Column : MCI GEL™ CHP50/P20, 32mmI.D.×465mm
Eluent : 0~185min, CH₃OH:0.01M Acetic acid(35:65)
185~350min, CH₃OH:0.01M Acetic acid(50:50)
Flow rate : 7.48 mL/min
Detection : 280nm
Sample : extract of green tea leaves, injection volume 18.7 mL
1. Epigallocatechin
2. Catechin
3. Epicatechin
4. Epigallocatechin gallate
5. Gallocatechin
6. Epicatechin gallate
7. Caffeine



Application data of CHP series

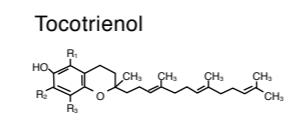
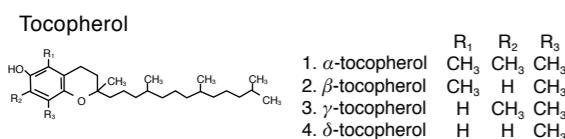


Fig. 5-41 Vitamin E in Rice Bran Oil

Conditions
Column : MCI GEL™ CMG20/C10
4.6mm I.D. \times 150mm
Eluent : Hexane-EtOH = 98/2 (vol.)
Flow rate : 0.5 mL/min
Detection : 295nm
Sample : Rice Bran Oil, 50 mg/mL
Injection : 10 μ L

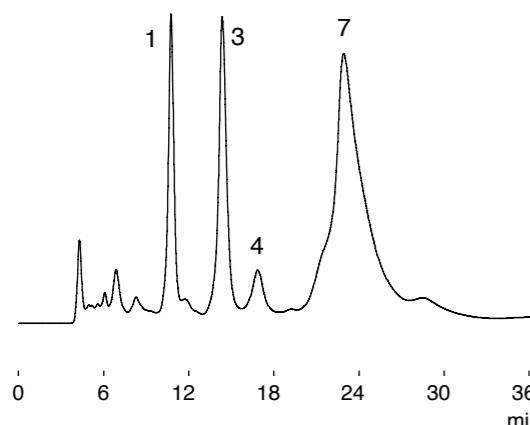


Fig. 5-42 Elution profile of Rice Bran Oil in preparative scale

Conditions
Column : MCI GEL™ CMG20/P30
20mm I.D. \times 500mm
Eluent : Hexane/C₂H₅OH = 98/2 (vol.)
Flow rate : 4.7 mL/min
Detection : 295 nm
Sample : Rice Bran Oil, 50 mg/mL
Injection : 1260 μ L

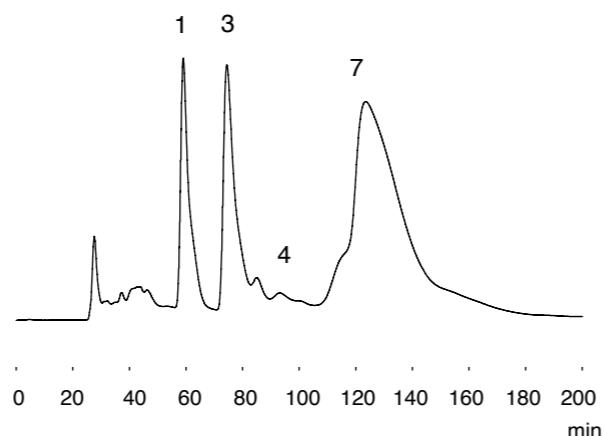


Fig. 5-43 Mixture of tocopherol and tocotrienol : Comparison with silica gel column

Conditions
Column : 1. Silica gel 5SIL, 4.6mm I.D. \times 250mm
2. MCI GEL™ CMG20/C04, 4.6mm I.D. \times 150mm
Eluent : 1. Hexane/EtOH = 99/1
2. Hexane/EtOH = 98/2
Flow rate : 1.0 mL/min
Column temp. : 25°C
Detection : UV 292nm
Sample : Mixture of tocopherol and tocotrienol
Injection : 10 μ L (1mg/mL)

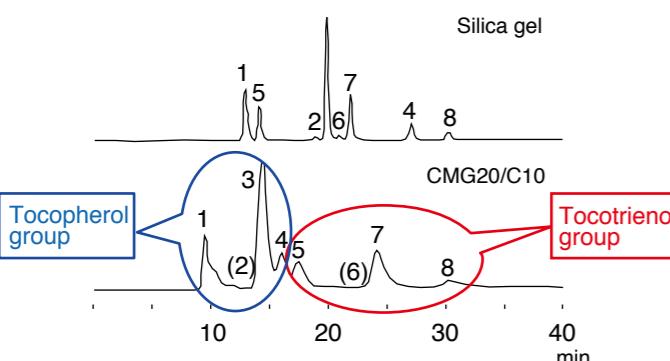
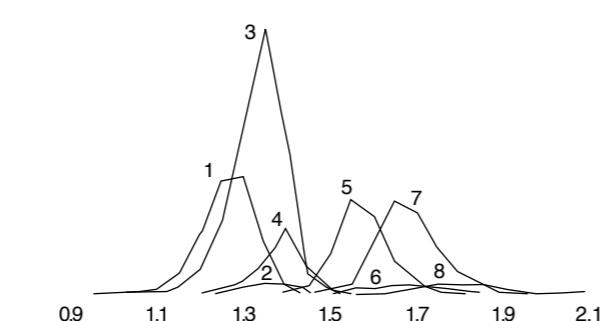


Fig. 5-44 Elution profile of tocopherol and tocotrienol in preparative scale

Conditions
Column : MCI GEL™ CMG20/P150, 41.2mm I.D. \times 550mm, \times 4
Eluent : Hexane/EtOH = 90/10
Flow rate : 49.0 mL/min (SV=1.0)
Column temp. : 25°C
Detection : UV 292nm
Sample : Mixture of tocopherol and tocotrienol
Injection : 150 mL (50g/L)



MCI GEL™

Chiral separation columns

6

○ Chiral separation columns MCI GEL™ CRS10W (DLAA)
MCI GEL™ CRS15W (LDAW)



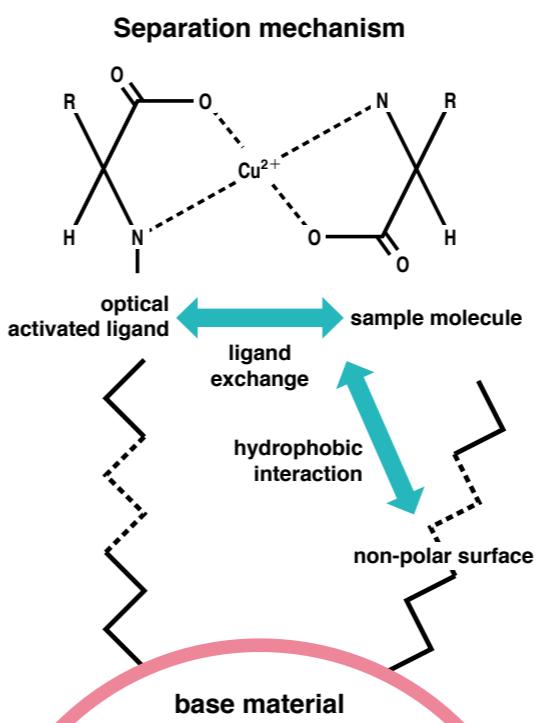
CRS10W 4.6 \times 50



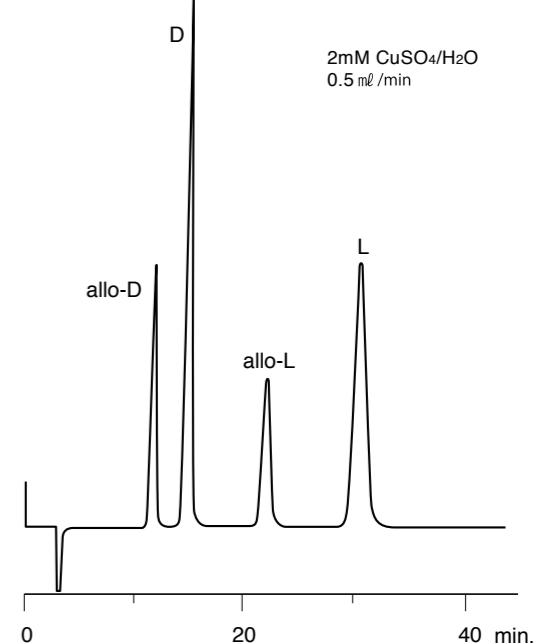
CRS15W 4.6 \times 50

MCI GEL™ column	Column dimensions	Particle size (μ m)	USP
MCI GEL™ CRS10W	4.6 \times 50mm	3	L32
MCI GEL™ CRS15W	4.6 \times 50mm	3	L32

Separation mechanism and performance of MCI GEL™ CRS series



Application of CRS10W
Fig. 6-1 DL-Isoleucine



Separation mechanism

MCI GEL™ CRS10W and its companion product MCI GEL™ CRS15W (an optical isomer of CRS10W) are based on a 3 μ m with 10nm mean pore diameter of silica gel coated with N,N-Dioctyl-L(or D)-alanine which is a novel optical activated ligand. The chiral resolution mechanism is a combination of ligand exchange and hydrophobic interaction. A copper sulfate aqueous solution is used as an eluent. Elution samples are directly detected at wave length of 254 nm because complex compound, composed of sample molecule and copper in the eluent, are object of detection. With the CRS10W, D-isomers generally elute in front of L-isomers while L-isomers elute ahead of D-isomers on the CRS15W. The hydrophobic interaction mechanism allows hydrophilic samples to elute faster than hydrophobic molecules. Long alkyl chain or aromatic compounds will elute late or require an organic solvent (CH₃CN or CH₃OH, max. of 15v/v%) to prevent adsorption onto the stationary phase.

Separation performance

- The CRS series columns separate over 20 D,L- α -Amino acids by only single column. The columns separate not only α -Amino acids but also α -Hydroxy carboxylic acids and derivative amino acids such as Acetylated amino acids.
- The columns provide excellent resolution operated at room temperature.
- The columns show high durability.

USP L32 column

Application data of CRS10W

For all chromatograms, column temperature is room temperature and wave length is 254nm.

All eluents are CuSO₄ aqueous solution except for Fig. 6-9 and Fig. 6-10.

Fig. 6-2 Separation of amino acids mixture

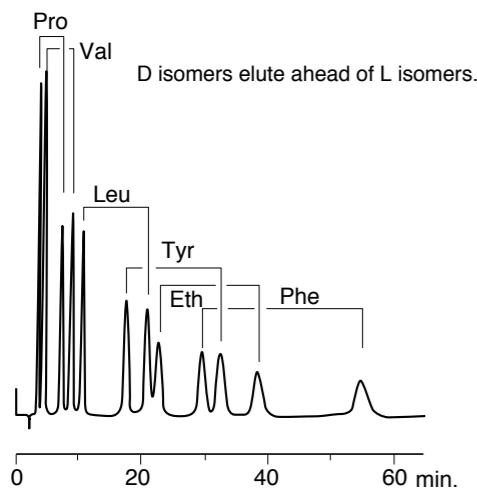


Fig. 6-3 Separation of amino acids mixture

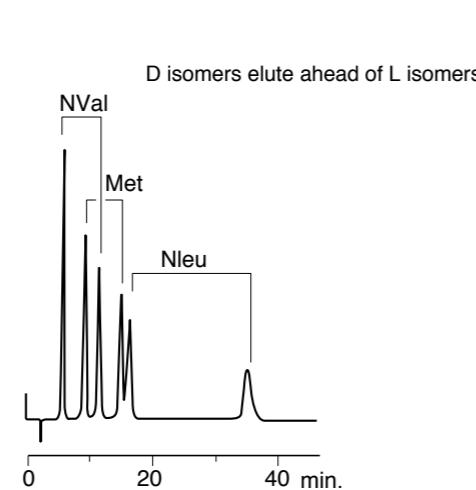


Fig. 6-4 Separation of DL-Ser.

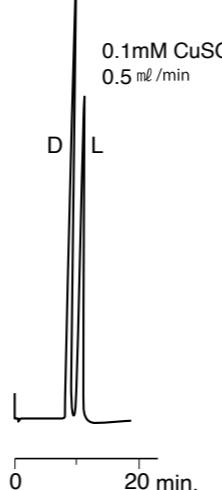


Fig. 6-5 Separation of DL-aspartic acid

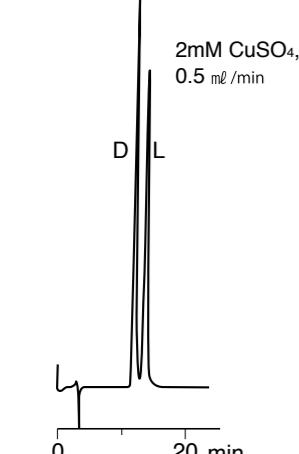


Fig. 6-6 Separation of DL-glutamic acid

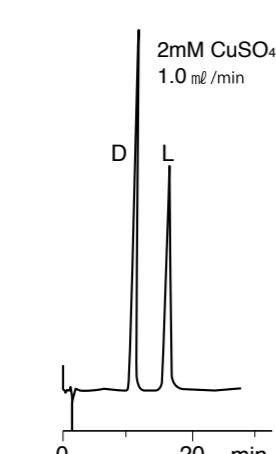


Fig. 6-7 Separation of DL-histidine

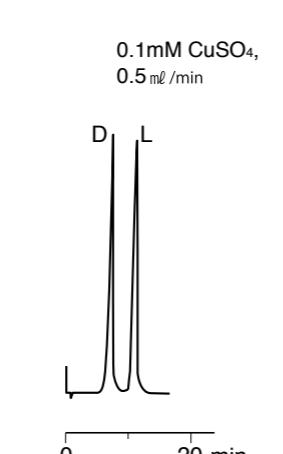


Fig. 6-8 Separation of DL-lysine

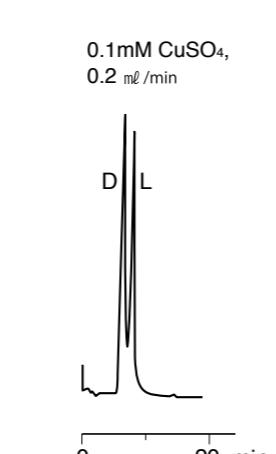


Fig. 6-9 Separation of DL-phenylalanine

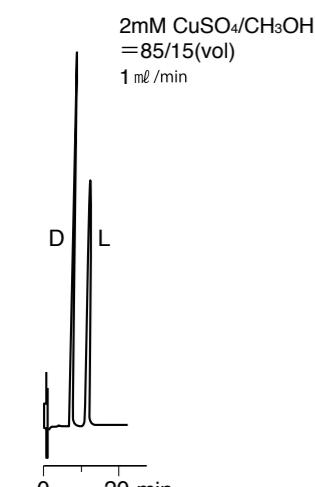


Fig. 6-10 Separation of DL-tryptophan

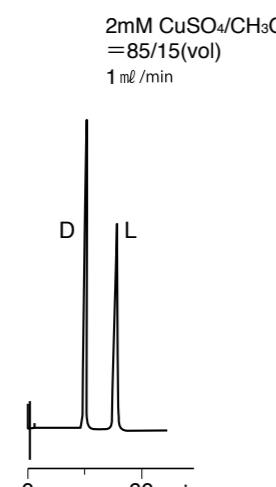


Fig. 6-11 Separation of DL-lactic acid

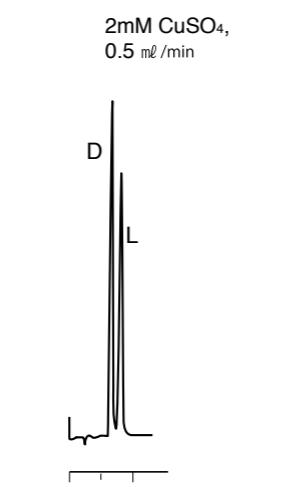
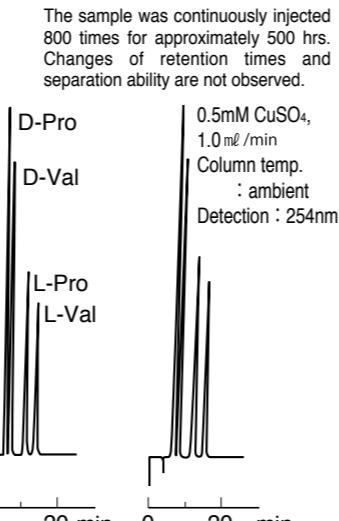


Fig. 6-12 Durability test



Application data of CRS10W

Fig. 6-13 Separation of DL- α -Phenylglycine

Conditions	
Column	: MCI GEL™ CRS10W 4.6mm I.D.×50mm
Eluent	: 2mM CuSO ₄ /CH ₃ OH=85/15
Flow rate	: 1.0 mL/min
Column temp.	: 25°C
Detection	: 254nm
Sample	: 1. D- α -Phenylglycine 2. L- α -Phenylglycine

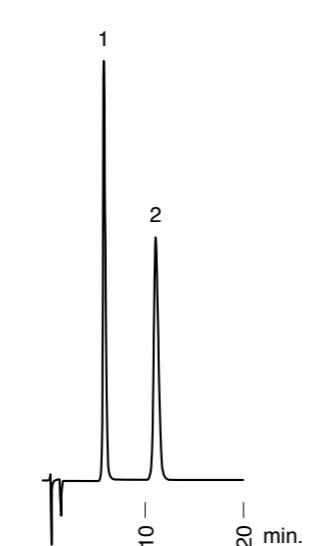


Fig. 6-14 Separation of methionine and acetylmethionine

Conditions	
Column	: MCI GEL™ CRS10W 4.6mm I.D.×50mm
Eluent	: 2mM CuSO ₄ /CH ₃ CN=90/10
Flow rate	: 1.0 mL/min
Column temp.	: 25°C
Detection	: 254nm
Sample	: 1. D-Met 2. L-Met 3. Acetyl-D-Met 4. Acetyl-L-Met

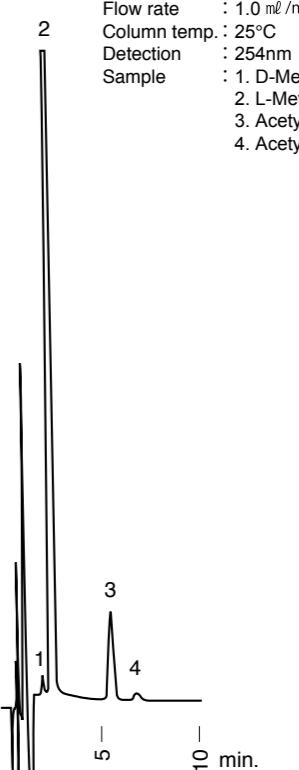


Fig. 6-15 D/L-Aspartic acid

Conditions	
Column	: MCI GEL™ CRS10W 4.6mm I.D.×50mm
Eluent	: 0.4mMCuSO ₄
Flow rate	: 1.0 mL/min
Temp.	: ambient
Detection	: UV 254nm
Sample	: D/L Aspartic acid

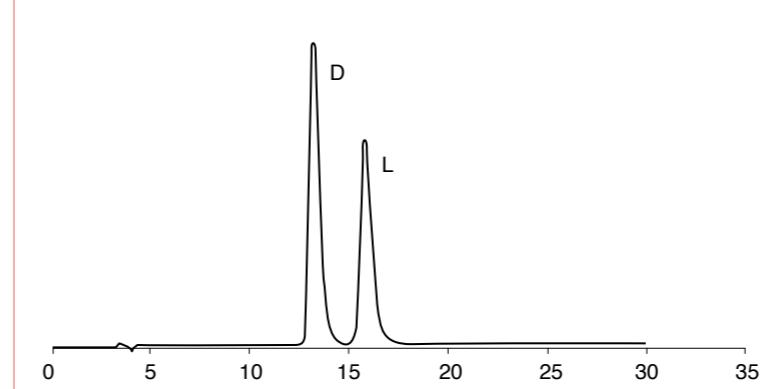
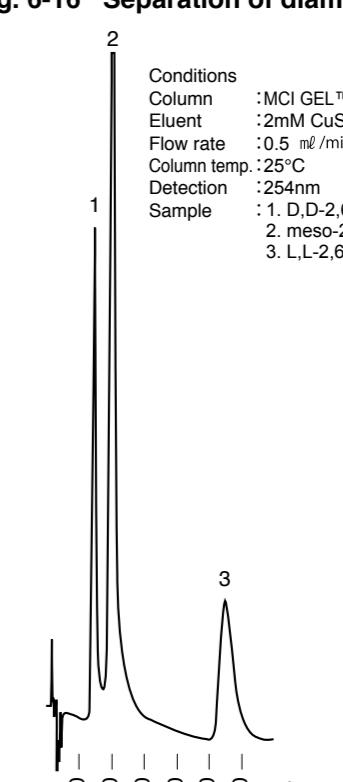


Fig. 6-16 Separation of diaminopimelic acid

Conditions	
Column	: MCI GEL™ CRS10W 4.6mm I.D.×50mm
Eluent	: 2mM CuSO ₄
Flow rate	: 0.5 mL/min
Column temp.	: 25°C
Detection	: 254nm
Sample	: 1. D,D-2,6-Diaminopimelic acid 2. meso-2,6-Diaminopimelic acid 3. L,L-2,6-Diaminopimelic acid



Application data of CRS10W

Fig. 6-17 Separation of 2-hydroxy carboxylic acids

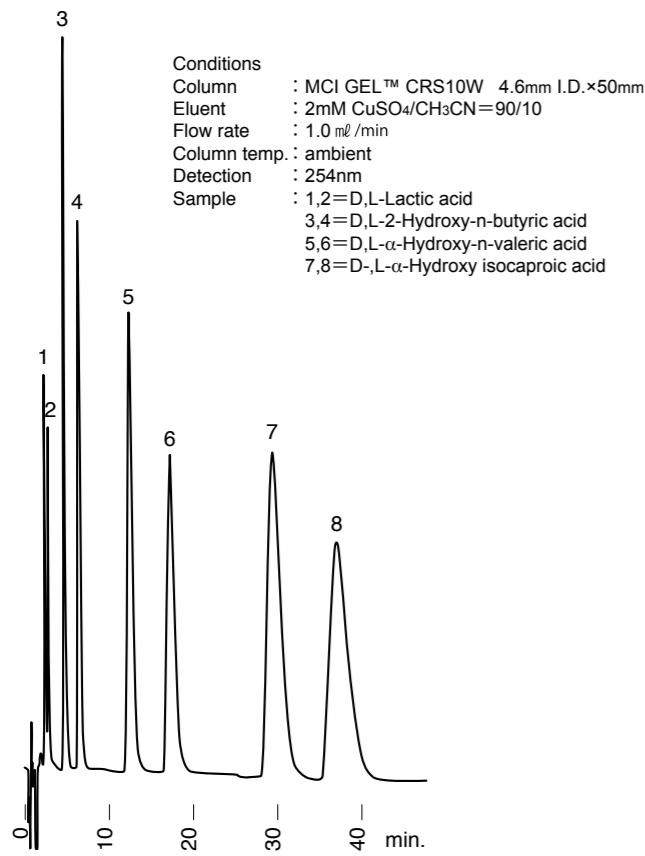


Fig. 6-18 Separation of 2-hydroxy carboxylic acids

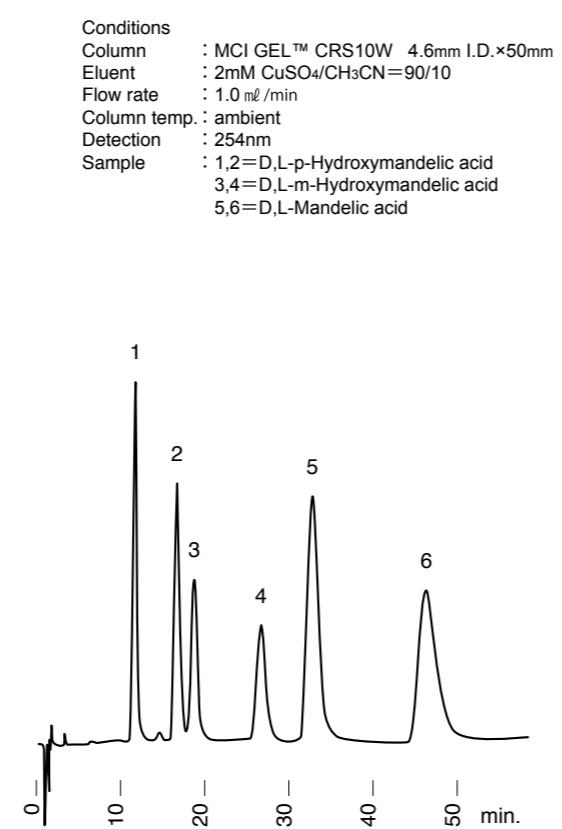


Fig. 6-19 D/L Alanine

Conditions
Column : MCI GEL™ CRS10W 4.6mm I.D.×50mm
Eluent : 0.1mMCuSO₄
Flow rate : 1.0 mL/min
Temp. : 30°C
Detection: UV 254nm
Sample : D/L Alanine

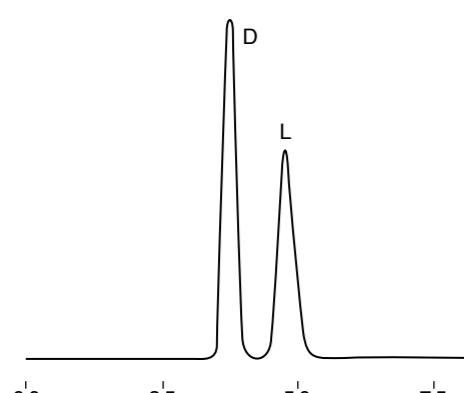
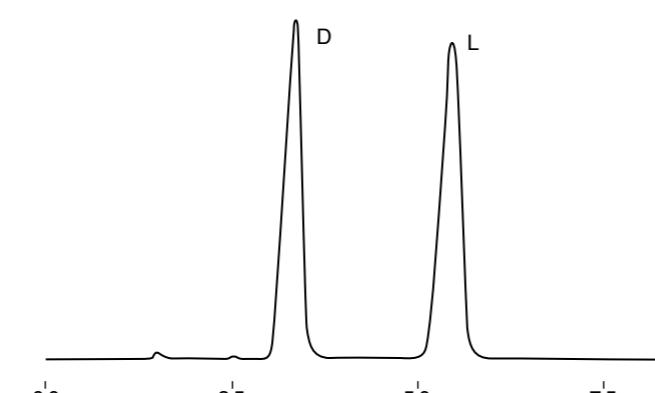


Fig. 6-20 DL-P-Hydroxyphenylglycine

Conditions
Column : MCI GEL™ CRS10W 4.6mm I.D.×50mm
Eluent : 2mMCuSO₄:MeOH=85:15
Flow rate : 1.0 mL/min
Temp. : 30°C
Detection: UV 254nm
Sample : DL-P-hydroxyphenylglycine



Comparison data of CRS10W and CRS15W

Fig. 6-21 Separation of DL-alanine

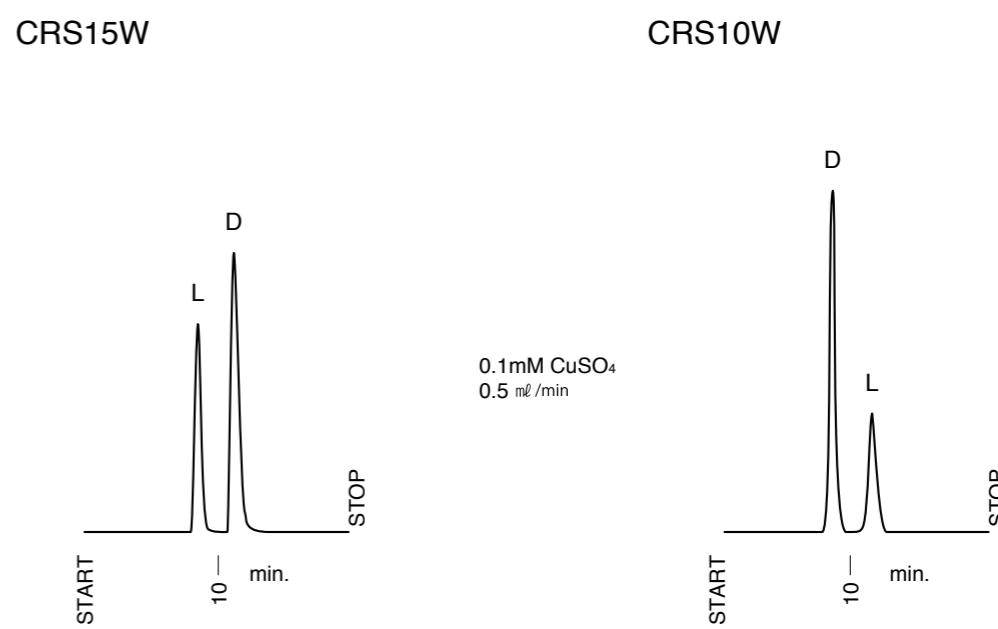
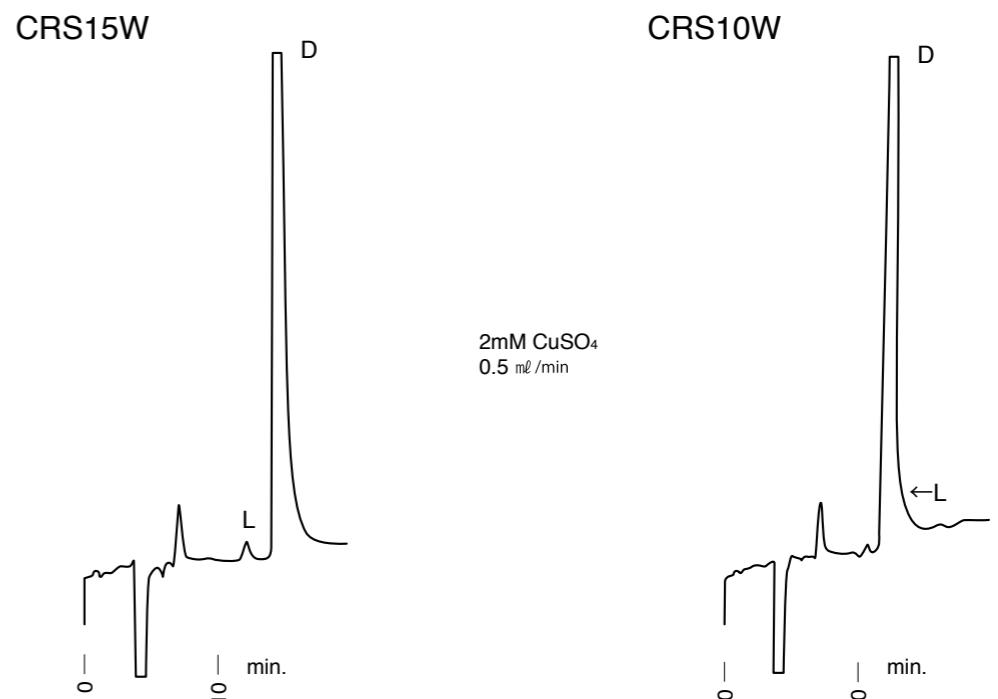


Fig. 6-22 Analysis of a trace of L-lactic acid in 50 ppm D-lactic acid

The CRS15W is recommended for analysis of a trace of L-isomer in a principal D-isomer when the CRS10W does not provide an adequate chromatogram.



Examples of chromatographic conditions and datas

	Amino acids	CuSO ₄ aq. soln. [mM]	Flow rate [mL/min]	Retention time; L-isomers [min]	Separation factor [α]	Separation rate [Rs]
1	Orn•HCl	0.1	0.2	6.8	1.26	<1
2	Lys•HCl	0.1	0.2	7.7	1.45	<1
3	Ala	0.1	0.5	11.0	1.39	1.4
4	His•HCl	0.1	0.5	10.5	1.63	1.7
5	Ser	0.1	0.5	10.1	1.25	1.0
6	Thr	0.1	0.5	11.3	1.29	1.3
7	Cit	0.5	0.5	10.4	1.75	2.3
8	Hyp	1.0	0.2	23.8	1.23	1.1
9	Pro	1.0	1.0	7.3	2.13	4.5
10	Val	1.0	1.0	8.9	2.04	5.0
11	Nval	1.0	1.0	11.5	2.07	4.7
12	Asp	2.0	0.5	13.2	1.18	0.8
13	Glu	2.0	1.0	16.2	1.54	2.3
14	Ileu(DL)	2.0	0.5	30.4	2.14	6.5
15	Ileu(allo)	2.0	0.5	21.9	1.97	6.0
16	Leu	2.0	1.0	14.6	1.97	4.6
17	Nleu	2.0	1.0	24.1	2.16	6.5
18	Met	2.0	1.0	10.3	1.64	2.6
19	Tyr	2.0	1.0	22.5	1.85	5.3
20	Eth	2.0	1.0	26.4	1.69	5.0
21	Phe	2.0	1.0	37.8	1.84	6.3

1. Column temperature; ambient Detection; 254nm
2. These are example data and do not guarantee the column specifications.
3. Improved resolution or appropriate chromatogram can be obtained by further investigating chromatographic conditions.
4. For each amino acid in the table, D-isomer elutes ahead of L-isomer except for Hydroxyproline.

Notes

1. It will take hours for equilibrium between ligand of stationary phase and copper ion of eluent. Two to three hours of conditioning the column with the eluent is advised before sample injection or after changing concentration of CuSO₄ of eluent.
2. For acidic amino acids, higher CuSO₄ concentration of eluent provides better resolution.
3. For weakly retained hydrophilic amino acids, low flow rate (0.2-0.5 mL/min) yields better resolution.
4. Peak area may decrease with continuous injection of samples, when the concentration of amino acids in sample solution is much higher than that of CuSO₄ in the eluent.
5. Please be careful not to flow both water soluble organic solvents (CH₃CN, CH₃OH, etc) and non water soluble organic solvents (n-hexane, chloroform, etc) into the column. The column will be fatally damaged and will never separate optical isomers. Please be particularly careful if HPLC equipment is used together with RP mode and NP mode.
6. Please do not use acid or alkali solutions to adjust pH of eluent. And also do not use buffer solutions. These solutions may cause forming precipitation, hence cause of blockage of the column.
7. For strongly retained hydrophobic amino acids, addition of CH₃CN or CH₃OH in the eluent enables faster elution. The concentration of these organic solvents should be below 15 v/v%.
8. DOPA and other non-polar amino acids will be strongly adsorbed on the packing material and will cause contamination of the column.
9. Regeneration of contaminated column is difficult.

MCI GEL™

7

SPE sorbent series

Solid phase extraction sorbents

For a pretreatment of analytical sample, we provide various SPE sorbents with various chemical structure, hydrophobicity, and micro-pore sizes. You can select our SPE sorbents depending on your molecule nature.

- CHP85/P120, CHP87/P120, CHPOD/P30: SPE sorbents with a controlled micro-pore size, high performance small molecule adsorption except large molecule mixture, like proteins.
- CSP800: SPE sorbents with high concentration ratio and high recovery, excellent for enrichment trace organic compounds and non-ionic substances such as trichloroethylene from environmental water. These SPE sorbents are to prepare samples for mutagenicity study or GC/MS analysis.
- SFP08/P25: SPE sorbents dedicated for small drug molecules extraction. Superior purity of this SPE extracts offers easier and faster sample preparation.
- CHL10P, CLH20P, CLB20P: SPE sorbents for rare earth metals that contains chelating functional group. CLB10P: SPE sorbents for borate, arsenic and selenium ions that contains glucamine groups on high porous ST/DVB matrix.

Material list

● Synthetic adsorbents and reversed-phase materials

Name	Mean particle size [μm]	Pore size	Surface area [m^2/g]	pH range	Typical Application
CHP85/P120	120	middle	880-940	full range	
CHP87/P120	120	small	820-910	full range	Small molecules extraction
CHPOD/P30	30	large	340-380	2~12	
CSP800	120	middle	790-920	full range	Enrichment of trace of organic compounds
SFP08/P25	25	middle	>1000	full range	Small molecules extraction

● Chelating type

Name	Functional group	Mean particle size [μm]	Ion exchange capacity [meq/mL]	Effective pH range	Typical Application
CHL10P	Iminodiacetic acid	120	>1.5	2-6	Metal Extraction
CHL20P	Polyamine	120	>1.8	2-6	Metal Extraction
CLB10P	Glucamine	120	>1.0	>3	Extraction Bron Removal

Main column				Guard/Pre-column		
Code No.	Name	Column dimensions [mm]	USP	Code No.	Name	Column dimensions [mm]
Ion exchange chromatography cation exchange resin for amino acids				0-019-01	CK10U	6×120
0-019-01	CK10U	6×120		0-033-21	AFR2-PC	6×50
Ion exchange chromatography cation exchange resin for sugars				0-009-01	CK08S	8×500
0-009-01	CK08S	8×500	L58	0-009-11	CK08SG	6×50
0-010-01 0-010-06	CK08E	8×300 7.8×300	L58	0-010-11	CK08EG	6×50
0-010-02 0-010-07	CK08EC	8×300 7.8×300	L19	0-010-12	CK08ECG	6×50
0-010-03	CK08ES	8×300		0-010-13	CK08ESG	6×50
Ion exchange chromatography cation exchange resin for carboxylic acids				0-010-05	CK08EH	8×300
0-010-05	CK08EH	8×300	L17	0-010-08		7.8×300
0-010-15	CK08EHG	6×50		0-010-15	CK08EHG	6×50
Ion exchange chromatography cation exchange resin for oligosaccharides				0-001-01	CK02A	20×250
0-001-01	CK02A	20×250	L58	0-001-11	CK02AG	8×10
0-001-02	CK02AS	20×250		0-001-12	CK02ASG	8×10
0-003-01	CK04S	10×200	L58	0-017-11 0-003-11	CK10SG CK04SG	6×50 8×10
0-003-02	CK04SS	10×200		0-017-11 0-003-12	CK10SG CK04SSG	6×50 8×10
Ion exchange chromatography anion exchange resin for carboxylic acids and sugars				0-111-01	CA08F	4.6×250
0-111-01	CA08F	4.6×250		0-111-11	CA08FG	4×10
0-119-01	CDR10	4.6×250		0-119-11	CDR10G	4×10
Ion chromatography for cations				0-034-01	SCK01	6×50
0-034-01	SCK01	6×50		0-034-21	SCK-PC	6×50
0-034-04	SCK01	4.6×150				
0-407-01	CHK45/C05	4.6×150				
Ion chromatography for anions				0-133-02	SCA04/PEEK	4.6×150
0-133-02	SCA04/PEEK	4.6×150	L31	0-133-12	SCA04G	4.6×30
				0-130-22	SCA-PC	8×10
Bioseparation for size exclusion				0-213-01	CQP06	7.5×600
0-213-01	CQP06	7.5×600	L25	0-213-11	CQP06G	4×50
0-214-01	CQP10	7.5×600	L38	0-214-11	CQP10G	4×50
0-215-01	CQP30	7.5×600	L37, 38	0-215-11	CQP30G	4×50

Main column				Main column			
Code No.	Name	Column dimensions [mm]	USP	Code No.	Name	Column dimensions [mm]	USP
Analytical and preparative chromatography columns for pharmaceutical applications [CHP column series]							
0-401-05	CHP20/C04	4.6X150	L21	0-047-21	Q01	4.6X50	
0-401-03	CHP20/C04	20X150	L21	0-047-22	Q01	4.6X100	
0-403-05	CHP20/C10	4.6X150	L21	0-047-23	Q01	7.5X100	
0-403-01	CHP20/C10	4.6X250	L21	0-047-24	Q01	11.5X100	
0-403-02	CHP20/C10	10X250	L21	0-047-31	CM01	4.6X50	
0-403-03	CHP20/C10	20X150	L21	0-047-32	CM01	4.6X100	
0-403-04	CHP20/C10	20X250	L21	0-047-33	CM01	7.5X100	
0-403-04	CHP20/C10	20X250	L21	0-047-34	CM01	11.5X100	
0-405-01	CHP07/C04	4.6X150					
0-405-04	CHP07/C04	20X200					
0-405-06	CHP07/C10	4.6X150					
0-406-01	CHP07/C10	4.6X250					
0-406-02	CHP07/C10	10X150					
0-406-03	CHP07/C10	20X150					
0-406-04	CHP07/C10	20X250					
0-402-05	CMG20/C04	4.6X150	L39				
0-402-03	CMG20/C04	20X150	L39				
0-202-06	CMG20/C10	4.6X150	L39				
0-202-05	CMG20/C10	4.6X250	L39				
0-202-02	CMG20/C10	10X250	L39				
0-202-03	CMG20/C10	20X150	L39				
0-202-04	CMG20/C10	20X250	L39				
0-404-01	CHK40/C04	4.6X150					
0-407-01	CHK45/C05	4.6X150					
0-504-01	CHPOD/C04	4.6X150					
0-504-04	CHPOD/C04	20X200					
XtalSpeed™ series							
0-047-01	DA01	4.6X50					
0-047-04	DA01	4.6X100					
0-047-02	DA01	7.5X100					
0-047-03	DA01	11.5X100					
0-047-11	SP01	4.6X50					
0-047-12	SP01	4.6X100					
0-047-13	SP01	7.5X100					
0-047-14	SP01	11.5X100					

9

MCI GEL™

MCI GEL™ material list

Characteristics

1. Excellent performance

Sphere packing and sharp particle size distribution provide high performance.

2. Persistence and highest quality

Produced with Mitsubishi Chemical's excellent technology, experience and under strict quality control.

3. Wide range of product line

MCI GEL™ packing materials include ion exchange resins (cation and anion), non-functionalized polymer used for reversed phase chromatography and other varieties of products. Also MCI GEL™ offers mean particle size of 4 µm to approximately 300 µm packing materials, this means that MCI GEL™ products are applied to analysis use and preparative use.

4. Abundant experience

Mitsubishi Chemical has been supplying packing materials for more than 50 years.

Ion exchange chromatography cation exchange resins [CK series, AFR series]

Code No.	Name	Packing size [g]	Base material	Functional group	Counter ion	Mean particle size [µm]	Cross linkage [%]	Ion exchange capacity [meq/g]	Typical Application
1-001-01	CK02A	10	ST/DVB	RSO ₃ ⁻	Na ⁺	20	2	>4.5	Oligosaccharides
1-003-01	CK04S	10	ST/DVB	RSO ₃ ⁻	Na ⁺	11	4	>4.5	Oligosaccharides
1-003-02	CK04S	25							
1-003-03	CK04S	50							
1-004-01	CK06S	10	ST/DVB	RSO ₃ ⁻	Na ⁺	11	6	>4.45	Oligosaccharides
1-004-02	CK06S	25							
1-004-03	CK06S	50							
1-009-01	CK08S	10	ST/DVB	RSO ₃ ⁻	Na ⁺	11	8	>4.40	Sugars, Carboxylic acids
1-009-02	CK08S	25							
1-009-03	CK08S	50							
1-010-01	CK08E	10	ST/DVB	RSO ₃ ⁻	Na ⁺	9	8	>4.40	Sugars, Carboxylic acids
1-010-02	CK08E	25							
1-010-03	CK08E	50							
1-013-01	CK08Y	50	ST/DVB	RSO ₃ ⁻	Na ⁺	25	8	>4.40	Sugars, Carboxylic acids
1-013-02	CK08Y	300							
1-014-01	CK08P	100 ml	ST/DVB	RSO ₃ ⁻	H ⁺	120	8	>1.8	Sugars, Carboxylic acids (meq/ml)
1-017-01	CK10S	10	ST/DVB	RSO ₃ ⁻	Na ⁺	11	10	>4.3	Carboxylic acids, Amino acids
1-017-02	CK10S	25							
1-017-03	CK10S	50							
1-018-01	CK10F	5	ST/DVB	RSO ₃ ⁻	Na ⁺	7	10	>4.3	Amino acids
1-018-02	CK10F	10							
1-019-01	CK10U	3	ST/DVB	RSO ₃ ⁻	Na ⁺	5	10	>4.3	Amino acids
1-019-03	CK10U	5							
1-019-04	CK10U	10							
1-020-05	CK10M	5	ST/DVB	RSO ₃ ⁻	Na ⁺	4	10	>4.3	Amino acids
1-020-06	CK10M	3							
1-024-02	CK12U	5	ST/DVB	RSO ₃ ⁻	Na ⁺	5	12	>4.3	Amino acids
1-021-01	CK10Y	50	ST/DVB	RSO ₃ ⁻	Na ⁺	25	10	>4.3	Amino acids
1-033-01	AFR2	5	ST/DVB	RSO ₃ ⁻	H ⁺	25	-	>2.7	Ammonia trap

Abbreviation; ST/DVB = Styrene-divinylbenzene copolymer

Column selection
guide

Ion exchange columns
and materials

Ion chromatography
columns and materials

Bioseparation columns
and materials

Analytical and preparative
chromatography columns and materials
for pharmaceutical applications

Chiral separation
columns

SPE sorbent series

MCI GEL™ column list

MCI GEL™ material list

Compounds index

10

Column selection
guide

1

Ion exchange columns
and materials

2

Ion chromatography
columns and materials

3

Bioseparation columns
and materials

4

Analytical and preparative
chromatography columns and materials
for pharmaceutical applications

5

Chiral separation
columns

6

SPE sorbent series

7

MCI GEL™ column list

8

MCI GEL™ material list

9

Compounds index

10

63

64

● Ion exchange chromatography anion exchange resins [CA series, CDR series]

Code No.	Name	Packing size [g]	Base material	Functional group	Counter ion	Mean particle size [μm]	Cross linkage [%]	Ion exchange capacity [meq./ml]	Typical Application
1-104-01	CA06S	10	ST/DVB	QA	Cl^-	11	6	>1.2	Sugars, Carboxylic acids
1-104-02	CA06S	25							
1-104-03	CA06S	50							
1-109-01	CA08S	10	ST/DVB	QA	Cl^-	11	8	>1.2	Sugars, Carboxylic acids
1-109-02	CA08S	25							
1-109-03	CA08S	50							
1-111-01	CA08F	5	ST/DVB	QA	Cl^-	7	8	>1.2	Sugars, Carboxylic acids
1-111-02	CA08F	10							
1-112-01	CA08Y	50							
1-113-01	CA08P	100 ml	ST/DVB	QA	Cl^-	120	8	>1.3	Sugars, Carboxylic acids
1-116-01	CA10S	10							
1-116-02	CA10S	25							
1-116-03	CA10S	50							
1-119-01	CDR10	7	ST/DVB	QA	Cl^-	7	-	>0.3	Nucleic acids, Sugars
1-119-02	CDR10	14							

Abbreviations : ST/DVB=styrene-divinyl benzene copolymer QA : Quaternary ammonium

● Ion chromatography materials [SCA, SCK series]

Code No.	Name	Packing size [g]	Base material	Functional group	Counter ion	Mean particle size [μm]	Ion exchange capacity [$\mu\text{eq/g}$]	Typical Application
1-034-01	SCK01	5	ST/DVB	RSO ³⁻	H ⁺	11	25	Cation analysis
1-034-02	SCK01	10						
1-133-01	SCA04	5	HMA	QA	Cl ⁻	5	30	Anion analysis
1-133-02	SCA04	10						

Abbreviations; ST/DVB = Styrene-divinylbenzene copolymer HMA = Polyhydroxymethacrylate QA = Quaternary ammonium

● Bioseparation columns -Size exclusion chromatography materials- [CQP series]

Code No.	Name	Packing size [ml]	Base material	Mean particle size [μm]	Pore size [nm]	Exclusion limit	Typical Application
1-222-01	CQP30P	100	HMA	30	60	1×10^6	Water soluble polymer

Abbreviation; HMA = Polyhydroxymethacrylate

● Bioseparation columns -Ion exchange materials- [CQA series, CQK series]

Code No.	Name	Packing size [ml]	Base material	Functional group	Counter ion	Mean particle size [μm]	Pore size [nm]	pH range	Typical Application
1-127-01	CQA31P	100	HMA	DEAE	Cl ⁻	30	60	<11	Proteins
1-131-01	CQA35P	100	HMA	QA	Cl ⁻	30	60	2~12	Proteins
1-037-01	CQK30P	100	HMA	SP	Na ⁺	30	60	1~13	Proteins
1-039-01	CQK31P	100	HMA	CM	Na ⁺	30	60	>4	Proteins

Abbreviations; HMA = Polyhydroxymethacrylate SP = Sulfopropyl CM = Carboxymethyl DEAE = Diethylaminoethyl QA = Quaternary ammonium

● Analytical and preparative chromatography materials for pharmaceutical applications [CHP material series]

Code No.	Product Name	Packing size [ml]	Base material	Mean particle size [μm]	Pore size [nm]	pH range	Typical Application
1-307-06	CHP20/P20	25	ST/DVB	20	45	full range	Reversed-phase chromatography
1-307-07	CHP20/P20	100					
1-307-08	CHP20/P20	1,000					
1-305-06	CHP20/P30	25	ST/DVB	30	45	full range	Reversed-phase chromatography
1-305-07	CHP20/P30	100					
1-305-08	CHP20/P30	1,000					
1-310-01	CHP20/P50	100g	ST/DVB	50	45	full range	Reversed-phase chromatography
1-313-02	CHP20/P70	500	ST/DVB	70	45	full range	Reversed-phase chromatography
1-313-03	CHP20/P70	1,000					
1-313-04	CHP20/P70	10,000					
1-311-01	CHP20/P120	100	ST/DVB	120	45	full range	Reversed-phase chromatography
1-311-02	CHP20/P120	500					
1-311-03	CHP20/P120	1,000					
1-311-04	CHP20/P120	10,000					
1-311-05	CHP20/P120	50,000					
1-304-06	CHP50/P20	25	ST/DVB	20	25	full range	Reversed-phase chromatography
1-304-07	CHP50/P20	100					
1-304-08	CHP50/P20	1,000					
1-303-06	CHP50/P30	25	ST/DVB	30	25	full range	Reversed-phase chromatography
1-303-07	CHP50/P30	100					
1-303-08	CHP50/P30	1,000					
1-312-01	CSP50/P10	10g	ST/DVB	10	25	full range	Reversed-phase chromatography
1-312-03	CSP50/P10	1,000					
1-314-02	CHP07/P120	100	ST/DVB	120	25	full range	Reversed-phase chromatography
1-314-03	CHP07/P120	1,000					
1-314-04	CHP07/P120	10,000					
1-314-05	CHP07/P120	50,000					
1-309-01	CMG20/P10	10g	MA	10	25	2~12	Reversed-phase chromatography
1-309-03	CMG20/P10	1,000					
1-306-06	CMG20/P30	25	MA	30	25	2~12	Reversed-phase chromatography
1-306-07	CMG20/P30	100					
1-306-08	CMG20/P30	1,000					
1-308-02	CMG20/P150	100	MA	150	25	2~12	Reversed-phase chromatography
1-308-03	CMG20/P150	1,000					
1-308-04	CMG20/P150	10,000					
1-308-05	CMG20/P150	50,000					

Abbreviations; MA=Polymethacrylate ST/DVB=Styrene-divinylbenzene copolymer

● Synthetic adsorbent and reversed-phase materials

Code No.	Product Name	Packing size	Mean particle size [μm]	Pore size	pH range	Typical Application
1-315-02	CHP85/P120	100mL	120	Middle	Full range	Small molecules extraction
1-316-02	CHP87/P120	100mL	120	Small	Full range	</td

10

MCI GEL™

Compounds index

Column selection

1

Ion exchange columns and materials

2

Ion chromatography columns and materials

3

Bioseparation columns and materials

4

Analytical and preparative chromatography columns and materials for pharmaceutical applications

5

Chiral separation columns

6

SPE sorbent series

7

MCI GEL™ column list

8

MCI GEL™ material list

9

Compounds index

10

	Compound	Classification	MCI GEL™ column	Figure	Page
1	Acetic acid	Acetic acid	CK08EH	2-13	11
2	Acetic acid	Acetic acid	CK08EH	2-16	12
3	Acetic acid	Acetic acid	CK08EH	2-18	12
4	Acetic acid	Acetic acid	CA08F	2-39	19
5	Acetic acid	Acetic acid	CA08F	2-40	20
6	Acetic acid	Acetic acid	CA08F	2-41	20
7	Acetic acid	Acetic acid	CA08F	2-42	20
8	N-Acetylgalactosamine	N-Acetylgalactosamine	CK08EH	2-14	11
9	N-Acetylglucosamine	N-Acetylglucosamine	CK08EH	2-14	11
10	Acetyl-D-Met.	Acetyl-D-Met.	CRS10W	6-14	56
11	Acetyl-L-Met.	Acetyl-L-Met.	CRS10W	6-14	56
12	Adenine	Nucleic base	CDR10	2-43	21
13	Adenine	Nucleic base	CHK40/C04	5-25	45
14	Adenine	Nucleic base	CHK45/C05	5-33	48
15	Adenosine	Nucleoside	SCK01	3-6	24
16	Adenosine	Nucleoside	CHK40/C04	5-25	45
17	Adenosine	Nucleoside	CHK45/C05	5-33	48
18	Adonitol	Adonitol	CK08EC	2-1	7
19	5'-ADP	Nucleotide	CDR10	2-43	21
20	Alanine	Alanine	CK10U	2-32	17
21	Alanine	Amino acid	CK10U	2-36	18
22	β-Alanine	β-Alanine	CK10U	2-33	18
23	D-Alanine	D-Alanine	CRS10W/CRS15W	6-21	58
24	D-Alanine	D-Amino acid	CRS10W	6-19	57
25	L-Alanine	L-Alanine	CRS10W/CRS15W	6-21	58
26	L-Alanine	L-Amino acid	CRS10W	6-19	57
27	γ-Aminobutyric acid	γ-Aminobutyric acid	CK10U	2-34	18
28	6-Aminopenicillanic acid	6-Aminopenicillanic acid	CHP50/P20	5-35	50
29	Ammonia	Ammonia	SCK01	3-2	24
30	Ammonium ion	Cation	SCK01	3-1	24
31	Ammonium ion	Cation	SCK01	3-3	24
32	Ammonium ion	Cation	CHK45/C05	3-10	25
33	2'-AMP	Nucleotide	CDR10	2-43	21
34	2'-AMP	Nucleotide	CDR10	2-44	21
35	3'-AMP	Nucleotide	CDR10	2-44	21
36	5'-AMP	Nucleotide	CDR10	2-43	21
37	5'-AMP	Nucleotide	CDR10	2-44	21
38	AMP	Nucleotide	CHK45/C05	5-33	48
39	Amphotericin B	Amphotericin B	CHP20/C10	5-17	41
40	Angiotensin II	Angiotensin II	CMG20/C04	5-12	40
41	Antipyrine	Antipyrine	CMG20/C04	5-10	39
42	Arginine	Arginine	CK10U	2-32	17
43	Ascorbic acid	Carboxylic acid	CA08F	2-41	20
44	Aspartic acid	Aspartic acid	CK10U	2-32	17
45	D-Aspartic acid	D-Aspartic acid	CRS10W	6-5	55
46	D-Aspartic acid	D-Amino acid	CRS10W	6-15	56
47	L-Aspartic acid	L-Aspartic acid	CRS10W	6-5	55
48	L-Aspartic acid	L-Amino acid	CRS10W	6-15	56
49	Aspirin	Aspirin	CMG20/C04	5-10	39
50	5'-ATP	Nucleotide	CDR10	2-43	21
51	Barium ion	Barium ion	SCK01	3-7	25
52	Bromide ion	Bromide ion	SCA04	3-11	26
53	Bromide ion	Bromide ion	SCA04	3-12	26
54	Buserelin	Buserelin	CHP20/C10	5-22	44
55	Buserelin	Buserelin	CMG20/C10	5-22	44
56	n-Butyl alcohol	n-Butyl alcohol	CK08EH	2-15	11
57	sec-Butyl alcohol	sec-Butyl alcohol	CK08EH	2-15	11
58	Cadmium ion	Cadmium ion	SCK01	3-8	25
59	Caffeine	Caffeine	CHP20/C04	5-4	36
60	Caffeine	Caffeine	CMG20/C04	5-10	39

	Compound	Classification	MCI GEL™ column	Figure	Page
61	Caffeine	Caffeine	CHP50/P20	5-36	50
62	Calcium ion	Calcium ion	SCK01	3-7	25
63	Calcium ion	Calcium ion	SCK01	3-8	25
64	Calcium ion	Calcium ion	SCK01	3-9	25
65	Calcium ion	Cation	CHK45/C05	3-10	25
66	Candesartan	Candesartan	CHK40/C04	5-28	47
67	Capsaicin	Alkaloid	CMG20/C10	5-40	52
68	Carbonate ion	Carbonate ion	SCA04	3-14	27
69	Catechin	Catechin	CHP50/P20	5-36	50
70	Cellulose	Cellulose	CA08F	2-38	19
71	Cellulose	Cellulose	CDR10	2-45	22
72	Cesium ion	Cation	SCK01	3-1	24
73	Chloride ion	Anion	SCA04	3-11	26
74	Chloride ion	Anion	SCA04	3-12	26
75	Chloride ion	Anion	SCA04	3-13	26
76	Chloride ion	Anion	SCA04	3-14	27
77	Chloride ion	Anion	SCA04	3-16	27
78	Chloroacetic acid	Carboxylic acid	CK08EH	2-16	12
79	Chloroacetic acid	Carboxylic acid	CK08EH	2-18	12
80	Cholic acid	Bile acid	CHP20/C04	5-7	37
81	α-Chymotrypsinogen A	Protein	CMG20/C04	5-13	40
82	α-Chymotrypsinogen A	Protein	CMG20/C10	5-18	42
83	Cinchonine	Cinchona alkaloid	CHP20/C04	5-5	36
84	Citric acid	Carboxylic acid	CK08EH	2-13	11
85	Citric acid	Carboxylic acid	CA08F	2-39	19
86	Citric acid	Carboxylic acid	CA08F	2-40	20
87	2'-CMP	Nucleotide	CDR10	2-44	21
88	3'-CMP	Nucleotide	CDR10	2-44	21
89	5'-CMP	Nucleotide	CDR10	2-43	21
90	5'-CMP	Nucleotide	CDR10	2-44	21
91	Cobalt ion	Cation	SCK01	3-8	25
92	Conalbumin	Protein	CHP20/C10	5-19	42
93	Corticosterone	Adrenal cortical hormone	CHP20/C04	5-9	38
94	Crocin	Crocin	CMG20/P30	5-39	52
95	3',5'-Cyclic AMP	Nucleotide	CDR10	2-43	21
96	Cystine	Amino acid	CK10U	2-32	17
97	Cytidine	Nucleoside	SCK01	3-6	24
98	Cytidine	Nucleoside	CHK40/C04	5-25	45
99	Cytidine	Nucleoside	CHK45/C05	5-33	48
100	Cytosine	Nucleic base	CDR10	2-43	21
101	Cytosine	Nucleic base	CHK40/C04	5-25	45
102	Cytosine	Nucleic base	CHK45/C05	5-33	48
103	Cytochrome c	Protein	CQP30	4-6	32
104	Cytochrome c	Protein	CMG20/C04	5-13	40
105	Cytochrome c	Protein	CMG20/C10	5-18	42
106	β-Citronellol	Perfume	CMG20/C10	5-26	46
107	Deoxycholic acid	Bile acid	CHP20/C04	5-7	37
108	11-Deoxycortisol	Adrenal cortical hormone	CHP20/C04	5-9	38
109	Deoxyribose	Deoxysugar	CA08F	2-38	19
110	Deoxyribose	Deoxysugar	CDR10	2-45	22
111	D,D-2,6-Diaminopimelic acid	D,D-Diamino carboxylic acid	CRS10W	6-16	56
112	L,L-2,6-Diaminopimelic acid	L,L-Diamino carboxylic acid	CRS10W	6-16	56
113	meso-2,6-Diaminopimelic acid	meso-Diamino carboxylic acid	CRS10W	6-16	56
114	Dibutyl phthalate	Aromatic ester	CHP50/P20	5-34	50
115	Dichloroacetic acid	Carboxylic acid	CK08EH	2-16	12
116	Dichloroacetic acid	Carboxylic acid	CK08EH	2-18	12
117	Diethylene glycol	Polyalcohol	CK08EH	2-17	12
118	Diethyl phthalate	Aromatic ester	CHP20/C04	5-3	36

	Compound	Classification	MCI GEL™ column	Figure	Page
1	Dopamine	Catecholamine	CHP20/C04	5-2	36
126	Epicatechin	Catechol	CHP50/P20	5-36	50
127	Epicatechin gallate	Catechol	CHP50/P20	5-36	50
128	Epigallocatechin	Catechol	CHP50/P20	5-36	50
129	Epigallocatechin gallate	Catechol	CHP50/P20	5-36	50
130	Epigallocatechin gallate	Catechol	CHP50/P20	5-36	50
131	Epinephrine	Catecholamine	CHP20/C04	5-2	36
132	Erythritol	Sugar alcohol	CK08EC	2-8	8
133	Erythritol	Sugar alcohol	CK08EC	2-9	9
134	Erythritol	Sugar alcohol	CK08EC	2-10	9
135	meso-Erythritol	Sugar alcohol	CK08EC	2-1	7
136	D-Ethionine	D-Amino acid	CRS10W	6-2	55
137	L-Ethionine	L-Amino acid	CRS10W	6-2	55
138	Ethyl alcohol	Alcohol	CK08EC	2-8	8
139	Ethyl alcohol	Alcohol	CK08EH	2-15	11
140	Ethyl alcohol	Alcohol	CK08EC	2-9	9
141	Ethylene glycol	Polyalcohol	CK08EH	2-15	11
142	Ethylene glycol	Polyalcohol	CK08EH	2-17	12
143	Ferritin	Protein	CQP30	4-6	32
144	Filipin	Antibiotic	CHP20/C10	5-17	41
145	Fluoride ion	Anion	SCA04	3-11	26
146	Fluoride ion	Anion	SCA04	3-12	26
147	Formic acid	Carboxylic acid	CK08EH	2-13	11
148	Formic acid	Carboxylic acid	CA08F	2-39	19
149	Formic acid	Carboxylic acid	CA08F	2-41	20
150	Fructose	Sugar	CK08EC	2-1	7
151	Fructose	Sugar	CK08EC	2-2	7
152	Fructose	Sugar	CK08EC	2-4	8
153	Fructose	Sugar	CK08EC	2-5	8
154	Fructose	Sugar	CK08EC	2-6	8
155	Fructose	Sugar	CK08EC	2-7	8
156	Fructose	Sugar	CK08EC	2-8	8
157	Fructose	Sugar	CK08EC	2-9	9
158	Fructose	Sugar	CK08EC	2-10	9
159	Fructose	Sugar	CK08EC	2-11	10
160	Fructose	Sugar	CK04S	2-28	16
161	Fructose	Sugar	CK04S	2-29	16
162	Fructose	Sugar	CK04S	2-30	16
163	Fructose	Sugar	CA08F	2-38	19
164	Fructose	Sugar	CDR10	2-45	22
165	Fructo-oligosaccharide	Fructo-oligosaccharide	CK04S	2-30	16
166	Fucose	Sugar	CA08F	2-38	19
167	Galactose	Sugar	CK08EC	2-3	7
168	Galactose	Sugar	CK08EC	2-11	10
169	Galactose	Sugar	CK06SC	2-19	12
170	Galactose	Sugar	CA08F	2-38	19
171	Galactose	Sugar	CDR10	2-45	22
172	Gallocatechin	Catechol	CHP50/P20	5-36	50
173	GDP	Nucleotide	CHK45/C05	5-33	48
174	Gentiobiose	Disaccharide	CK08EC	2-1	7
175	Gentiobiose	Disaccharide	CK08EC	2-10	9
176	Geraniol	Perfume	CMG20/C10	5-26	46
177	Ghrelin human	Peptide	CMG20/C10	5-21	43
178	Ghrelin rat	Peptide	CMG20/C10	5-21	43
179	Gluconic acid	Carboxylic acid	CA08F	2-40	20
180	Gluconic acid	Carboxylic acid	CA08F	2-42	20
181	Gluconic acid	Carboxylic acid	CQP06	4-7	32
182	Glucose	Sugar	CK08EC	2-1	7
183	Glucose	Sugar	CK08EC	2-2	7
184	Glucose	Sugar	CK08EC	2-4	8
185	Glucose	Sugar	CK08EC	2-5	8
186	Glucose	Sugar	CK08EC	2-6	8
187	Glucose	Sugar	CK08EC	2-7	8
188	Glucose	Sugar	CK08EC	2-8	8
189	Glucose	Sugar	CK08EC	2-9	9
190	Glucose	Sugar	CK08EC	2-10	9

	Compound	Classification	MCI GEL™ column	Figure	Page
191	Glucose	Sugar	CK08E	2-12	10
192	Glucose	Sugar	CK08EH	2-14	11
193	Glucose	Sugar	CK04S	2-28	16
194	Glucose	Sugar	CK04S	2-29	16
195	Glucose	Sugar	CK04S	2-30	16
196	Glucose	Sugar	CA08F	2-38	19
197	Glucose	Sugar	CDR10	2-45	22
198	Glucose	Sugar	CQP06	4-7	32
199	Glutamic acid	Amino acid	CK10U	2-32	17
200	D-Glutamic acid	D-Amino acid	CRS10W	6-6	55
201	L-Glutamic acid	L-Amino acid	CRS10W	6-6	55
202	Glycerol	Polyalcohol	CK08EC	2-8	8
203	Glycerol	Polyalcohol	CK08EC	2-9	9
204	Glycerol	Polyalcohol	CK08EH	2-15	11
205	Glycine	Amino acid	CK10U	2-32	17
206	Glycolic acid	Carboxylic acid	CK08EH	2-13	11
207	Glycolic acid	Carboxylic acid	CK08EH	2-18	12
208	Glycyrrhizic acid	Glycyrrhizic acid	CHP20/C04	5-8	38
209	Gly-Tyr	Peptide	CMG20/C04	5-12	40
210	3'-GMP	Nucleotide	CDR10	2-44	21
211	5'-GMP	Nucleotide	CDR10	2-44	21
212	5'-GTP	Nucleotide	CDR10	2-43	21
213	Guanosine	Nucleoside	SCK01	3-6	24
214	Guanosine	Nucleoside	CHK40/C04	5-25	45
215	Guanosine	Nucleoside	CHK40/C04	5-25	45
216	Guanosine	Nucleoside	CHK45/C05	5-33	48
217	Guanosine	Nucleoside	CHK45/C05	5-33	48
218	Histidine	Amino acid	CK10U	2-32	17
219	D-Histidine	D-Amino acid	CRS10W	6-7	55
220	L-Histidine	L-Amino acid	CRS10W	6-7	55
221	Hemoglobin A0	Protein	SP01	4-2	30
222	Hemoglobin A1c	Protein	SP01	4-2	30
223	5-HPA	Amino acid	CK10U	2-35	18
224	Hydrocortisone	Adrenal cortical hormone	CHP20/C04	5-9	38
225	5-Hydroxytryptophan	Amino acid	CHP20/C04	5-2	36
226	D-2-Hydroxy-n-butrylic acid	D- α -Hydroxycarboxylic acid	CRS10W	6-17	57
227	L-2-Hydroxy-n-butrylic acid	L- α -Hydroxycarboxylic acid	CRS10W	6-17	57
228	D- α -Hydroxy isocaproic acid	D- α -Hydroxycarboxylic acid	CRS10W	6-17	57
229	L- α -Hydroxy isocaproic acid	L- α -Hydroxycarboxylic acid	CRS10W	6-17	57
230	5-hydroxy lysine	Amino acid	CK10U	2-37	18
231	D- α -Hydroxy-n-valeric acid	D- α -Hydroxycarboxylic acid	CRS10W	6-17	57
232	L- α -Hydroxy-n-valeric acid	L- α -Hydroxycarboxylic acid	CRS10W	6-17	57
233	D-m-Hydroxymandelic acid	D- α -Hydroxycarboxylic acid	CRS10W	6-18	57
234	L-m-Hydroxymandelic acid	L- α -Hydroxycarboxylic acid	CRS10W	6-18	57
235	D-p-Hydroxymandelic acid	D- α -Hydroxycarboxylic acid	CRS10W	6-18	57
236	L-p-Hydroxymandelic acid	L- α -Hydroxycarboxylic acid	CRS10W	6-18	57
237	D-P-Hydroxy phenylglycine	D-Amino acid	CRS10W	6-20	57
238	L-P-Hydroxy phenylglycine	L-Amino acid	CRS10W	6-20	57
239	Hydroxy proline	Amino acid	CK10U	2-35	18
240	Hypoxanthine	6-Hydroxypurine	CHP20/C04	5-6	37
241	Hypoxanthine	Nucleic base	CHK40/C04	5-25	45
242	Hypoxanthine	Nucleic base	CHK45/C05	5-33	48
243	5'-IMP	Nucleotide	CDR10	2-44	21
244	Inosine	Nucleoside	CHK40/C04	5-25	45
245	Inosine	Nucleoside	CHK45/C05	5-33	48
246	Insulin human recombinant	Peptide	CHP20/C10	5-20	43
247	Insulin glargine	Peptide	CHP20/C10	5-20	43
248	Insulin human recombinant	Peptide	CMG20/C10	5-20	43
249	Insulin glargine	Peptide	CMG20/C10	5-20	43
250	Irbesartan	Sartan	CHK40/C04	5-28	47
251	Isoleucine	Amino acid	CK10U	2-32	17
252	Isoleucine	Amino acid	CK10U	2-36	18
253	D-Isoleucine	D-Amino acid	CRS10W	6-1	54
254	L-Isoleucine	L-Amino acid	CRS10W	6-1	54
255	allo-D-Isoleucine	D-Amino acid	CRS10W	6-1	54

	Compound	Classification	MCI GEL™ column	Figure	Page
1	256 allo-L-Isoleucine	L-Amino acid	CRS10W	6-1	54
ion exchange columns and materials	257 Isopropyl alcohol	Alcohol	CK08EH	2-15	11
2	258 Kinase	Enzyme	DA01	4-3	30
ion chromatography columns and materials	259 Lactic acid	Carboxylic acid	CK08EH	2-13	11
3	260 Lactic acid	Carboxylic acid	CA08F	2-39	19
Bioseparation columns and materials	261 Lactic acid	Carboxylic acid	CA08F	2-41	20
4	262 Lactic acid	Carboxylic acid	CA08F	2-42	20
Analytical and preparative chromatography columns and materials for pharmaceutical applications	263 D-Lactic acid	D- α -Hydroxycarboxylic acid	CRS10W	6-11	55
5	264 L-Lactic acid	L- α -Hydroxycarboxylic acid	CRS10W	6-11	55
Chiral separation columns	265 D-Lactic acid	D- α -Hydroxycarboxylic acid	CRS10W	6-17	57
6	266 L-Lactic acid	L- α -Hydroxycarboxylic acid	CRS10W	6-17	57
SPE sorbent series	267 D-Lactic acid	D- α -Hydroxycarboxylic acid	CRS10W/CRS15W	6-22	58
7	268 L-Lactic acid	L- α -Hydroxycarboxylic acid	CRS10W/CRS15W	6-22	58
MCI GEL™ column list	269 β -Lactoglobulin	Protein	CMG20/C10	5-18	42
8	270 Lactose	Disaccharide	CK08EC	2-1	7
MCI GEL™ material list	271 Lactose	Disaccharide	CK08EC	2-3	7
9	272 Lactose	Disaccharide	CK06SC	2-19	12
Compounds index	273 Lactose	Disaccharide	CA08F	2-38	19
10	274 Lactose	Disaccharide	CDR10	2-45	22
	275 Lactose monohydrate	Disaccharide	CK08EC	2-10	9
	276 Lactulose	Disaccharide	CK08EC	2-3	7
	277 Lactulose	Disaccharide	CK06SC	2-19	12
	278 Leucine	Amino acid	CK10U	2-32	17
	279 Leucine	Amino acid	CK10U	2-36	18
	280 Nor-Leucine	Amino acid	CK10U	2-36	18
	281 D-Leucine	D-Amino acid	CRS10W	6-2	55
	282 L-Leucine	L-Amino acid	CRS10W	6-2	55
	283 Leu-Enkephalin	Peptide	CMG20/C04	5-12	40
	284 Leuprorelin	Peptide	CHP20/C10	5-22	44
	285 Leuprorelin	Peptide	CMG20/C10	5-22	44
	286 LH-RH human	Peptide	CHP20/C10	5-22	44
	287 LH-RH human	Peptide	CMG20/C10	5-22	44
	288 LH-RH salmon	Peptide	CHP20/C10	5-22	44
	289 LH-RH salmon	Peptide	CMG20/C10	5-22	44
	290 Linalool	Perfume	CMG20/C10	5-26	46
	291 Linalool	Perfume	CMG20/C10	5-27	46
	292 Linalyl acetate	Perfume	CMG20/C10	5-26	46
	293 Lithium ion	Cation	SCK01	3-1	24
	294 Lithium ion	Cation	CHK45/C05	3-10	25
	295 Losartan	Sartan	CHK40/C04	5-28	47
	296 Lysine	Amino acid	CK10U	2-32	17
	297 Lysine	Amino acid	CK10U	2-37	18
	298 D-Lysine	D-Amino acid	CRS10W	6-8	55
	299 L-Lysine	L-Amino acid	CRS10W	6-8	55
	300 Magnesium ion	Cation	SCK01	3-7	25
	301 Magnesium ion	Cation	SCK01	3-9	25
	302 Magnesium ion	Cation	CHK45/C05	3-10	25
	303 Malic acid	Carboxylic acid	CK08EH	2-13	11
	304 Malic acid	Carboxylic acid	CA08F	2-39	19
	305 Malic acid	Carboxylic acid	CA08F	2-41	20
	306 Malonic acid	Carboxylic acid	CK08EH	2-13	11
	307 Malonic acid	Carboxylic acid	CA08F	2-39	19
	308 Maltose	Disaccharide	CA08F	2-38	19
	309 Maltose	Disaccharide	CDR10	2-45	22
	310 Maltose	Disaccharide	CK08E	2-12	10
	311 Maltotriose	Disaccharide	CK08E	2-12	10
	312 D-Mandelic acid	D- α -Hydroxycarboxylic acid	CRS10W	6-18	57
	313 L-Mandelic acid	L- α -Hydroxycarboxylic acid	CRS10W	6-18	57
	314 Manganese ion	Cation	SCK01	3-8	25
	315 Mannitol	Sugar alcohol	CK08EC	2-1	7
	316 Mannitol	Sugar alcohol	CK08EC	2-8	8
	317 Mannitol	Sugar alcohol	CK08EC	2-9	9
	318 Mannitol	Sugar alcohol	CK08EC	2-10	9
	319 Mannose	Sugar	CK08EC	2-1	7
	320 Mannose	Sugar	CK08EC	2-10	9

	Compound	Classification	MCI GEL™ column	Figure	Page
1	321 Mannose	Sugar	CA08F	2-38	19
ion exchange columns and materials	322 Mannose	Sugar	CDR10	2-45	22
2	323 Melibiose	Disaccharide	CA08F	2-38	19
ion chromatography columns and materials	324 Melibiose	Disaccharide	CDR10	2-45	22
3	325 Met-Enkephalin	Peptide	CMG20/C04	5-12	40
Bioseparation columns and materials	326 Methionine	Amino acid	CK10U	2-32	17
4	327 D-Methionine	D-Amino acid	CRS10W	6-3	55
Analytical and preparative chromatography columns and materials for pharmaceutical applications	328 L-Methionine	L-Amino acid	CRS10W	6-3	55
5	329 D-Methionine	D-Amino acid	CRS10W	6-14	56
Chiral separation columns	330 L-Methionine	L-Amino acid	CRS10W	6-14	56
6	331 Methyl alcohol	Alcohol	CK08EH	2-15	11
SPE sorbent series	332 Methylamine	Amine	SCK01	3-2	24
7	333 γ -methyl leucine	Amino acid	CK10U	2-36	18
MCI GEL™ column list	334 Mevastatin	Medicine	CHP20/C10	5-16	41
8	335 Myoglobin	Protein	CQP30	4-6	32
MCI GEL™ material list	336 Nerol	Perfume	CMG20/C10	5-26	46
9	337 Nitrate ion	Anion	SCA04	3-11	26
Compounds index	338 Nitrate ion	Anion	SCA04	3-12	26
10	339 Nitrate ion	Anion	SCA04	3-13	26
	340 Nitrate ion	Anion	SCA04	3-14	27
	341 Nitrate ion	Anion	SCA04	3-16	27
	342 Nitrate ion	Anion	SCA04	3-11	26
	343 Nitrate ion	Anion	SCA04	3-12	26
	344 D-Norleucine	D-Amino acid	CRS10W	6-3	55
	345 L-Norleucine	L-Amino acid	CRS10W	6-3	55
	346 D-Norvaline	D-Amino acid	CRS10W	6-3	55
	347 L-Norvaline	L-Amino acid	CRS10W	6-3	55
	348 Nystatin	Antibiotic	CHP20/C10	5-17	41
	349 Oligosaccharide	Dp1-Dp9	CK04S	2-21	15
	350 Oligosaccharide	Dp1-Dp13	CK04SS	2-22	15
	351 Oligosaccharide	Dp1-Dp16	CK02A	2-23	15
	352 Oligosaccharide	Dp1-Dp20	CK02AS	2-24	15
	353 Oligosaccharide	Dp1-Dp7	CK04S	2-25	16
	354 Oligosaccharide	Dp1-Dp7	CK04SS	2-26	16
	355 Oligosaccharide	Dp1-Dp7	CK02AS	2-27	16
	356 Ornithine	Amino acid	CK10U	2-37	18
	357 Orotic acid	Carboxylic acid	CHP20/C04	5-6	37
	358 Ovalbumin	Protein	CQP30	4-6	32
	359 Oxalic acid	Carboxylic acid	CK08EH	2-13	11
	360 PEG MW 145,000	PEG	CQP30	4-5	32
	361 PEG MW 40,000	PEG	CQP30	4-5	32
	362 PEG MW 6,000	PEG	CQP30	4-5	32
	363 Penicillin G	Antibiotic	CHP50/P20	5-35	50
	364 Penicillin V	Antibiotic	CHP50/P20	5-35	50
	365 Phenacetin	Medicine	CMG20/C04	5-10	39
	366 Phenylalanine	Amino acid	CK10U	2-32	17
	367 Phenylalanine	Amino acid	CK10U	2-37	18
	368 D-Phenylalanine	D-Amino acid	CRS10W	6-2	55
	369 L-Phenylalanine	L-Amino acid	CRS10W	6-2	55
	370 D-Phenylalanine	D-Amino acid	CRS10W	6-9	55
	371 L-Phenylalanine	L-Amino acid	CRS10W	6-9	55
	372 D- α -Phenylglycine	D-Amino acid	CRS10W	6-13	56
	373 L- α -Phenylglycine	L-Amino acid	CRS10W	6-13	56
	374 Phosphate ion	Anion	SCA04	3-11	26
	375 Pipecolic acid	Amino acid	CK10U	2-35	18
	376 Polyphenol 60	Polyphenol	CHP07/C04	5-29	47
	377 Polyphenol 60	Polyphenol	CHP20/C04	5-30	47
	378 Potassium ion	Cation	SCK01	3-1	24
	379 Potassium ion	Cation	SCK01	3-3	24
	380 Potassium ion	Cation	SCK01	3-4	24
	381 Potassium ion	Cation	SCK01	3-5	24
	382 Potassium ion	Cation	CHK45/C05	3-10	25
	383 Pravastatin Na	Medicine	CHP20/C10	5-16	41
	384 Procainamide	Anesthetic	CMG20/C04	5-14	40
</td					

	Compound	Classification	MCI GEL™ column	Figure	Page
1	Proline	Amino acid	CK10U	2-32	17
2	Proline	Amino acid	CK10U	2-35	18
3	D-Proline	D-Amino acid	CRS10W	6-2	55
4	L-Proline	L-Amino acid	CRS10W	6-2	55
5	n-Propyl alcohol	Alcohol	CK08EH	2-15	11
6	Pyruvic acid	Carboxylic acid	CA08F	2-41	20
7	Quinine	Cinchona alkaloid	CHP20/C04	5-5	36
8	Rhamnose	Sugar	CA08F	2-38	19
9	Rhamnose	Sugar	CDR10	2-45	22
10	Ribitole(Adnitole)	Sugar alcohol	CK08EC	2-10	9
11	Ribonuclease A	Protein	CMG20/C04	5-13	40
12	Ribonuclease A	Protein	CMG20/C10	5-18	42
13	Ribonuclease A	Protein	CHP20/C10	5-19	42
14	Ribose	Sugar	CK08EC	2-1	7
15	Ribose	Sugar	CK08EC	2-10	9
16	Ribose	Sugar	CA08F	2-38	19
17	Ribose	Sugar	CDR10	2-45	22
18	Rituximab	Monoclonal antibody	SP01	4-1	30
19	Rubidium ion	Cation	SCK01	3-1	24
20	Salicin	Phenol glycoside	CK08EC	2-1	7
21	Salicin	Phenol glycoside	CK08EC	2-10	9
22	Sennoside A	Sennoside A	CHP20/C10	5-37	51
23	Sennoside B	Sennoside B	CHP20/C10	5-37	51
24	Sennoside A	Sennoside A	CHP20/P20	5-37	51
25	Sennoside B	Sennoside B	CHP20/P20	5-37	51
26	Sennoside A	Sennoside A	CHP20/P30	5-37	51
27	Sennoside B	Sennoside B	CHP20/P30	5-37	51
28	Sennoside A	Sennoside A	CHP20/P30	5-38	51
29	Serine	Amino acid	CK10U	2-32	17
30	D-Serine	D-Amino acid	CRS10W	6-4	55
31	L-Serine	L-Amino acid	CRS10W	6-4	55
32	Serotonin	Catecholamine	CHP20/C04	5-2	36
33	Sifuvirtide	Peptide	CMG20/C10	5-23	44
34	Simvastatin	Medicine	CHP20/C10	5-16	41
35	Sodium ion	Cation	SCK01	3-1	24
36	Sodium ion	Cation	SCK01	3-3	24
37	Sodium ion	Cation	SCK01	3-4	24
38	Sodium ion	Cation	SCK01	3-5	24
39	Sodium ion	Cation	CHK45/C05	3-10	25
40	Sorbitol	Sugar alcohol	CK08EC	2-2	7
41	ssRNA	RNA	CHP20/C10	5-24	45
42	Stachyose	Tetrasaccharide	CK08EC	2-1	7
43	Stachyose hydrate	Sugar	CK08EC	2-10	9
44	Strontium ion	Cation	SCK01	3-7	25
45	Strontium ion	Cation	SCK01	3-8	25
46	Succinic acid	Carboxylic acid	CA08F	2-41	20
47	Succinylsulfathiazole	Sulfa drug	CMG20/C04	5-11	39
48	Sucrose	Disaccharide	CK08EC	2-2	7
49	Sucrose	Disaccharide	CK08EC	2-4	8
50	Sucrose	Disaccharide	CK08EC	2-5	8
51	Sucrose	Disaccharide	CK08EC	2-11	10
52	Sucrose	Disaccharide	CK04S	2-30	16
53	Sulfate ion	Anion	SCA04	3-11	26
54	Sulfate ion	Anion	SCA04	3-12	26
55	Sulfate ion	Anion	SCA04	3-13	26
56	Sulfamerazine	Sulfa drug	CMG20/C04	5-11	39
57	Sulfanilamide	Sulfa drug	CMG20/C04	5-11	39
58	Sulfathiazole	Sulfa drug	CMG20/C04	5-11	39
59	Tartaric acid	Carboxylic acid	CK08EH	2-13	11
60	Tartaric acid	Carboxylic acid	CA08F	2-39	19
61	Tartaric acid	Carboxylic acid	CA08F	2-41	20
62	Tert-leucine	Amino acid	CK10U	2-36	18
63	Theobromine	Purine alkaloid	CHP20/C04	5-4	36
64	Theophylline	Purine alkaloid	CHP20/C04	5-4	36
65	Theophylline	Purine alkaloid	CHP20/C04	5-6	37

	Compound	Classification	MCI GEL™ column	Figure	Page
1	Thiocyanic ion	Anion	SCA04	3-15	27
2	Thiosulfuric ion	Anion	SCA04	3-15	27
3	Threonine	Amino acid	CK10U	2-32	17
4	Thymidine	Nucleoside	CHK40/C04	5-25	45
5	Thymidine	Nucleoside	CHK45/C05	5-33	48
6	Thymine	Nucleic base	CDR10	2-43	21
7	Thymine	Nucleic base	CHK40/C04	5-25	45
8	D- α -Tocopherol	Vitamin	CMG20/C04	5-41	53
9	D- γ -Tocopherol	Vitamin	CMG20/C04	5-41	53
10	D- δ -Tocopherol	Vitamin	CMG20/C04	5-41	53
11	D- α -Tocopherol	Vitamin	CMG20/P30	5-42	53
12	D- γ -Tocopherol	Vitamin	CMG20/P30	5-42	53
13	D- δ -Tocopherol	Vitamin	CMG20/P30	5-42	53
14	D- α -Tocopherol	Vitamin	CMG20/C04	5-43	53
15	D- β -Tocopherol	Vitamin	CMG20/C04	5-43	53
16	D- γ -Tocopherol	Vitamin	CMG20/C04	5-43	53
17	D- δ -Tocopherol	Vitamin	CMG20/C04	5-43	53
18	D- α -Tocopherol	Vitamin	CMG20/P150	5-44	53
19	D- β -Tocopherol	Vitamin	CMG20/P150	5-44	53
20	D- γ -Tocopherol	Vitamin	CMG20/P150	5-44	53
21	D- δ -Tocopherol	Vitamin	CMG20/P150	5-44	53
22	D- α -Tocotrienol	Vitamin	CMG20/C10	5-41	53
23	D- γ -Tocotrienol	Vitamin	CMG20/C10	5-41	53
24	D- δ -Tocotrienol	Vitamin	CMG20/C10	5-41	53
25	D- γ -Tocotrienol	Vitamin	CMG20/C04	5-41	53
26	D- δ -Tocotrienol	Vitamin	CMG20/C04	5-41	53
27	D- α -Tocotrienol	Vitamin	CMG20/P30	5-42	53
28	D- α -Tocotrienol	Vitamin	CMG20/C04	5-43	53
29	D- β -Tocotrienol	Vitamin	CMG20/C04	5-43	53
30	D- γ -Tocotrienol	Vitamin	CMG20/C04	5-43	53
31	D- δ -Tocotrienol	Vitamin	CMG20/C04	5-43	53
32	D- γ -Tocotrienol	Vitamin	CMG20/C04	5-43	53
33	D- δ -Tocotrienol	Vitamin	CMG20/C04	5-43	53
34	D-TPN	Nucleotide	CDR10	2-43	21
35	Transferrin	Protein	CMG20/C10	5-18	42
36	Trichloroacetic acid	Carboxylic acid	CK08EH	2-16	12
37	Triethylene glycol	Polyalcohol	CK08EH	2-17	12
38	Trimethylamine	Amine	SCK01	3-2	24
39	TritonX-100	Surfactant	CHPOD/04	5-31	48
40	TritonX-100	Surfactant	ODS-1HU	5-32	48
41	Tryptophan	Amino acid	CHP20/C04	5-2	36
42	D-Tryptophan	D-Amino acid	CRS10W	6-10	55
43	L-Tryptophan	L-Amino acid	CRS10W	6-10	55
44	Tyrosine	Amino acid	CK10U	2-32	17
45	Tyrosine	Amino acid	CK10U	2-37	18
46	D-Tyrosine	D-Amino acid	CRS10W	6-2	55
47	L-Tyrosine	L-Amino acid	CRS10W	6-2	55
48	3'-UMP	Nucleotide	CDR10	2-44	21
49	5'-UMP	Nucleotide	CDR10	2-44	21
50	UMP	Nucleotide	CHK45/C05	5-33	48
51	Uracil	Nucleic base	CDR10	2-43	21
52	Uracil	Nucleic base	CHK40/C04	5-25	45
53	Uracil	Nucleic base	CHK45/C05	5-33	48
54	Uric acid	2,6,8-Trioxypurine	CHP20/C04	5-6	37
55	Uridine	Nucleoside	SCK01	3-6	24
56	Uridine	Nucleoside	CHK40/C04	5-25	45
57	Uridine	Nucleoside	CHK45/C05	5-33	48
58	Urine	Urine	CDR10	2-46	22
59	Ursodeoxycholic acid	Bile acid	CHP20/C04	5-7	37
60	Valine	Amino acid	CK10U	2-32	17
61	Valine	Amino acid	CK10U	2-33	18
62	Valine	Amino acid	CK10U	2-36	18
63	D-Valine	D-Amino acid	CRS10W	6-2	55

	Compound	Classification	MCI GEL™ column	Figure	Page

<tbl

	Compound	Classification	MCI GEL™ column	Figure	Page
516	L-Valine	L-Amino acid	CRS10W	6-2	55
517	Valsartan	Sartan	CHK40/C04	5-28	47
518	Vitamin B3	Water soluble vitamin	CMG20/C04	5-15	40
519	Vitamin B6	Water soluble vitamin	CMG20/C04	5-15	40
520	Vitamin B12	Water soluble vitamin	CMG20/C04	5-15	40
521	Vitamin C	Water soluble vitamin	CMG20/C04	5-15	40
522	Xanthine	2,6-Dihydroxypurine	CHP20/C04	5-6	37
523	Xanthine	Nucleic base	CHK40/C04	5-25	45
524	Xanthine	Nucleic base	CHK45/C05	5-33	48
525	Xanthine oxidase	Nucleic base	CHK45/C05	5-33	48
526	Xanthosine	Nucleoside	CHK40/C04	5-25	45
527	Xylitol	Sugar alcohol	CK08EC	2-1	7
528	Xylitol	Sugar alcohol	CK08EC	2-10	9
529	Xylose	Sugar	CA08F	2-38	19
530	Xylose	Sugar	CDR10	2-45	22
531	Zinc ion	Cation	SCK01	3-7	25

Limited warranty

Mitsubishi Chemical Corporation warrants that its pre-packed columns (including separation media products) shall meet published specifications at the time of shipment from Mitsubishi Chemical Corporation. Because of the susceptibility of these products to deterioration, all warranty claims must be made within the stipulated in the listed sales office. All claims shall be deemed waived in the event the purchaser fails to notify the company within the period.

Conditions

A. The products in this brochure are for laboratory or manufacturing use. They are not intended for drug, medicine, food additive or household use. Compliance with local and government regulations concerning their use is the responsibility of the purchaser.

B. Voiding of warranty :

This warranty is null and void if any product has been (1) altered or modified such that its stability or reliability is any way affected ; (2) misused ; or (3) damaged by abuse, negligence or accident. The term "misuse" includes, but is not limited to, use not in compliance with the "Column Handling Instructions".

C. Limitations and Exclusions :

All recommendations, information and descriptions supplied by Mitsubishi Chemical Corporation with respect to any product in this brochure are believed to be accurate and reliable, but do not constitute warranties. The sole liability of Mitsubishi Chemical Corporation for any breach of warranty is limited to replacement, or at the sole option of Mitsubishi Chemical Corporation a refund of the purchase price.

Changes

All specifications, quantities, designs and prices are subject to change without notice.

General

Neither this publication, nor any products in this brochure shall be construed as recommending the infringement of any patent, nor extending any license, express or implied, nor assuming any liability under any issued or pending patent. The data presented herein have been carefully compiled from our records which we believe to be accurate and reliable. We make, however, no warranties or representations with respect hereto, nor is freedom from any patent to be inferred.



Related Product Lineup

DIAION™
SEPABEADES™
MabSpeed™
ChromSpeed™

Ion Exchange Resins (DIAION™)

Strongly Acidic Cation Exchange Resins
(Gel, Porous, Highly Porous Type and Industrial Chromatography)
DIAION™ SK, UBK, PK series

Weakly Acidic Cation Exchange Resins
(Methacrylic and Acrylic Type)
DIAION™ WK series

Strongly Basic Anion Exchange Resins
(Gel, Porous, and Highly Porous Type)
DIAION™ SA, UBA, PA, HPA series

Weakly Basic Anion Exchange Resins
(Acrylic, Polyamine, and Dimethylamine Type)
DIAION™ WA series

Chelating Resins (DIAION™)

Iminodiacetate type, Polyamine Type, and Glucamine Type
DIAION™ CR series

Synthetic Adsorbents (DIAION™/SEPABEADS™)

Aromatic Type, Modified Aromatic Type, and Methacrylic Type
DIAION™ HP series
SEPABEADES

Bioseparation Media

Affinity Chromatography Resins
MabSpeed™ RP series

Strongly Acidic Cation Exchange Resins
ChromSpeed™ S series

Weakly Acidic Cation Exchange Resins
ChromSpeed™ CM series

Strongly Basic Anion Exchange Resins
ChromSpeed™ Q series

Weakly Basic Anion Exchange Resins
ChromSpeed™ DA series

Screening Columns
MabSpeed™, ChromSpeed™